Evaluation of Edible Oyster Mushroom (*Pleurotus ostreatus*) on Oxidative Stress and Neurological Cognitive Disorder in Streptozotocin –Diabetic Rats

Abstract

*Pleurotus ostreatus* is drawn into lime light for the treatment of Diabetes mellitus for the last few years, but now it is evidently proven for the recovery of the diabetes mellitus-induced memory impairments in streptozotocin induced diabetic rats. Oxidative stress, neurological disorders, cognitive and spatial learning associated to Diabetic-Alzheimer’s is irreversibly inhibited by *P. ostreatus*. The availability and cost effectively makes a stepping stone for the middle to low income people to rely on *P. ostreatus* rather than of costly medications for the treatment of neurological and cognitive disorders associated with Diabetes. The research was done to identify a potent free radical bioactive compound against cognitive impairment in streptozotocin –diabetic rats. Five weeks after diabetic induction, *P. ostreatus* extract was administered orally (2 mg/l). The cognitive, cerebral and perceptive behaviour was examined with T Maze (Radial Arm Maze), Morris Water Maze (MWM), Novel object recognition task in wistar rats (male). Besides oxidative stress parameters like Lipid peroxidation (LPO), FRAP (Ferric reducing ability of plasma), antioxidant assay and Thiol assay to determine total Thiol group in the blood was carried out to determine the neurological disorders associated with brain. *P. ostreatus* showed significant improvements in Spatial and Cognitive disorders when compared to the diabetic and healthy control. Also the FRAP and thiol group in blood showed tremendous increase due to the *P. ostreatus* proving the effectiveness of *Pleurotus* extract on Cognitive impairment and Oxidative stress in Diabetic mellitus.

Graphical Abstract

Keywords: *P. ostreatus* • Oxidative stress • Diabetic • Streptozotocin • Histopathology

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Introduction

Diabetes mellitus, a blooming disorder linked with Cognitive disorder related neurological inefficiency [1-3]. Cellular machinery oxidation in human body is by the continuous exposure to different types of molecules that enumerates reactive species known as free radicals (ROS/RNS) [4]. These Free radical arbitrated imbalances in systemic brain manifestation with oxidative stress share a foremost part in septohippocampial dysfunction in diabetes [5, 6]. Autoxidation of glucose, glycation of protein and polyol pathway are the cellular mechanisms that seem to be complicated in diabetes [1]. In some cases this free radicals causes necrosis and ultimately to the cell death [7]. Cellular functions are protected by antioxidant enzymes which maintain in vivo homeostasis throughout oxidative stress [6]. Brain is extremely prone to overproduction of ROS and faulty antioxidant and leading to neurodegeneration and association cognitive decline in aging and Alzheimer’s disease [7-9]. AD is allied to pathogenetic mechanisms with multiple aetiologies. Free radical prompted oxidative stress, diminishing of cerebral energy metabolism along with excitotoxic events are most promising events for cognitive disorder studies [10,11]. Ageing and age associated neurodegenerative disorders such as Alzheimer’s disease (AD) and Parkinson’s disease stay concomitant with varying degrees of cognitive impairment leading to indisposition [12] and neuronal oxidative stress which leads to behavioural effects [13]. Patients by diabetes mellitus are reported with tripping enhancements of spatial impairment functioning [14,15]. Previous evidence suggests that the hyperglycemia as a “toxic” effect prominent to diabetic end-organ damage to brain in Type I and Type II diabetes [16]. Diabetic patients may express cognitive deficits even at their young age even more worse during elderly [17]. Recent epidemiological studies even report relation flanked by diabetes and dementia. Cognitive discrepancies are also reported in rat replicas of diabetes. Learning deficits are seen in streptozotocin convinced diabetic rats which is preventable, but not fully reversible with insulin treatment [16]. Cerebral dysfunction pathogenesis in diabetes is yet to be explicates [17]. Age-associated Perceptive debility and AD can be initially treated with antioxidant drugs [18]. Mild to modest diminishing of intellectual functioning (DM) are seen in patients with diabetes mellitus (DM) [12,13]. Hyperglycemia is considered as the major precursor for End organ damage to brain in Type I and Type II diabetes [14]. Elderly people are marked with intellectual deficits associated to young diabetic adults [15]. Diabetes and Dementia are even correlated in current research for example perceptive scarcities are also reported in rat models of diabetes [16]. Diabetic patients with Cerebral dysfunction pathogenesis is an emerging era for study of Intellectual and perceptive shortages [17]. Thus it is estimated that age-associated cognitive decline such as AD could be prevented or treated using antioxidant drugs [18]. Edible mushrooms is considered equally similar to meat for vegan in the modern world [18]. P. ostreatus is primarily consumed for its taste, fiberiness and nutritive value, besides it is also used industrially as a bioremediator [19,20]. The active principles of P. ostreatus are isolated for direct use as drugs, lead compounds or pharmacological agents [21], Interleukin-12 production, nitric oxide synthase activation and iron chelating properties are mediated by the bioactive compounds derived from mushrooms [22,23]. Great threat to human life by AD lingers to escalation and thus the pursuit for AD drugs takes a compelling urgency [24]. Studies are still being conducted in lots of countries to explore the utilization and consuming strategy of mushrooms and their metabolites for human ailments [25]. The antitumor, anti-diabetic, anti-mutagenic, anti-proliferative, anticarcinogenic properties and neuroprotective in models of degenerative are the most attractive features of edible oyster mushrooms. These properties are thought to be mediated by the phenols by means of b-carotene, tocopherols, and total polyphenols [26]. Therefore the current research investigated the consequences of P. ostreatus on oxidative damage and spatial cognition in streptozotocin-convincing diabetic rats.

Methodology

Identification and handling of sample

P. ostreatus was purchase from Green Grow Cultivation Unit, Big Shop, Ooty, Tamilnadu and authentication was done by Dr. Annamalai, Plant Biologist, and Coimbatore. It was reserved in a hygenic, sterilized pliable container and sealed. The fruiting hyphae was washed and fragmented and finally dried in incubator at 38°C.

Preparation of extract

P. ostreatus extract was prepared by drying at 38°C for 36 h. The dehydrated sections was weighed and grounded prior to extraction. Dried sample (50 gm) was extracted with 400 ml of 100% ethanol for 9 h using soxhlet apparatus. Rotary evaporator was used in vacuo to at 40°C [27].

Refinement of bioactive compound

HPTLC: Bioactive compounds were determined
qualitatively by High Performance Thin Layer Chromatography (HPTLC). The presence of Alkaloids, Flavonoids, Steroids was established with Dragendorff’s reagent by Keller-Killani test (Table 1 and Figure 1) [28].

**Chemicals and drugs**

Dithiononitrobenzoic acid (DTNB), Tris base, 1,1,3,3'-tetraethoxypropane (MDA), 2-thiobarbituric acid (TBA), trichloroacetic acid (TCA), n-butanol, 2,4,6-tripyridyl-s-triazine (TPTZ), from Merck Chemical Co. (Tehran), Streptozotocin Sigma Chemical Co and *P. ostreatus* were used in this study.

**Animals and experimental design**

Male Wistar rats weighing approximately 180±10 g acquired from the Pasteur Institute Coonoor, India. Temperature maintenance was about of 25±2°C and 12/12 h of light–dark cycle. Experiments were conducted with guidelines and norms of Animal Ethics Committee. The experimental rats were assigned into five groups; each group contained 8-10 animals:

(i) Control rats (C); (ii) Control with Streptozotocin (C+D); (iii) *P. ostreatus* treated control rats (C+E); (iii) diabetic rats treated with *P. ostreatus* (D+E); (iv) diabetic rats treated with Std Glibenciclamide drug).

**Table 1: Preliminary Phytochemical Screening of Pleurotus ostreatus Extract by HPTLC**

<table>
<thead>
<tr>
<th>TEST FOR EXTRACT</th>
<th>INFERENCE</th>
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<tbody>
<tr>
<td>Alkaloid</td>
<td>+</td>
</tr>
<tr>
<td>Flavonoid</td>
<td>+</td>
</tr>
<tr>
<td>Steroid</td>
<td>+</td>
</tr>
<tr>
<td>Terpenoid</td>
<td>+</td>
</tr>
</tbody>
</table>

+ = Presence    - = Absence

**After Derivitization**

- Alkaloids (Day light)
- Flavonoid CV 366nm
- Steroid UV 366nm
- Terpenoid UV 366nm

*Figure 1:* TLC plates showing the presence of Alkaloid, Flavonoid, Steroid and Terpenoid after derivitization process
A single intraperitoneal dose of streptozotocin (STZ) (50 mg (kg body weight)$^{-1}$) dissolved in 0.2 mL of normal saline was used to induce diabetes in rats. After few days the stasis of confirmation of diabetes was done by tail vein identifying the amount of glucose in the tail vein. A rat with blood glucose levels of 250 mg/dL was set to be diabetic. After diabetes induction, *P. ostreatus* extract was directed orally to rats for 8 weeks (3 mg /L) [29].

**Behavioral tests**

**Wisdom and recollection:** Morris water maze (MWM) Neurological disorders with cognitive components like AD and Schizophrenia are studied by knowledge and recollecting pattern in animal models [13-15]. MWM is one among them in identifying the cognitive ailments allied with neuron stimulated brain reactions. In this research 5-day training was conducted for animals. After every 60 hr 4 dimensional memory retaining capability was tested. It was conducted in square shaped tank with the dimension of 60 × 120 cm (L X B). This tank was with water at a depth of 25 cm. Extra cues were also attached. The acrylic sheet platform was submerged in the water tank at a deepness of 2 cm as an escalator for the rats to come out after the trial of escape latency test. The four quadrants were marked and to the each quadrant rats were allowed to swim to find the sheet for escape latency. The time taken by each rats were noted down along with the error time taken when the rats exploited the same quadrant. The animal which remained exploring even after 1 minute was noted by the experimenter. No trial period was performed since sociability relies directly on cognitive disorders. This assessment was conducted for the animals to identify the empty boxes and the boxes with a social partner of the same breed or different breed. The time taken by the each animal to recognize the empty box was noted as well as error time was also noted when the same empty boxes was rechecked again and again [33,34].

**Sociability:** Sociability is a novel method to study the dejection and violent actions. Besides these there is a detail platform to study autism. In this basic test of sociability the repeated time the rat applies exploring a new impetus and the error time for an empty box was noted by the experimenter. No trial period was performed since sociability relies directly on cognitive disorders. This assessment was conducted for the animals to identify the empty boxes and the boxes with a social partner of the same breed or different breed. The time taken by the each animal to recognize the empty box was noted as well as error time was also noted when the same empty boxes was rechecked again and again [31,32].

**Simple visual task:** On week 8 after STZ-administration, performance on a ‘simple visual task’ was evaluated. In this task the performance of identification, behavior, recognition with same breed was identified for the animals with diabetic and normal healthy control rats. This comparative study elucidates the memory errors, working memory errors, comparative errors and so on with the help of longitudinal study pattern.

**Biochemical analysis**

**Estimation of oxidative stress parameters:** The superoxide dismutase (SOD), glutathione peroxidase (GSH-Px), catalase (TAC) activities, and glutathione (GSH) level in hepatic and pancreatic tissue were measured using commercially available kits (Sigma Aldrich, Chennai). Lipid peroxidation was measured as malondialdehyde (MDA) level in hepatic and pancreatic tissue according to Jain's method [35].

**Assay of cellular lipid peroxidation (LPO):** Lipid peroxidation was measured using TBA. The rat plasma sections were mixed with TCA (20) and the precipitate was collected and dispersed in Sulphuric acid (0.05
M). Sodium sulfate 2 M was added and heated for 30 minutes in boiling water bath. The absorbance of the pink complex was measured at 532 nm [36].

**Assay of total antioxidant capacity (TAC):** TAC was measured by ferric reducing ability of plasma (FRAP) method. This method is based on the ability of plasma in reducing Fe$^{3+}$ to Fe$^{2+}$ in the presence of TPTZ. The reaction of Fe$^{2+}$ and TPTZ gives a complex with blue color and maximum absorbance in 593 nm [37].

**Statistical analysis**

Readings was observed in Standard deviation using ANOVA table linked to Dunkan test. Statistically significant variations are compared as follows: control versus *P. ostreatus* canned control; control versus diabetic and diabetic versus *P. ostreatus* treated diabetic rats. Results were considered significantly different if p<0.05.

**Results**

**Effect of *P. ostreatus* extracts on the behaviour patterns and antioxidant assay**

Administration of *P. ostreatus* decreased escape latency, travelled distance and speed in *P. ostreatus* control and diabetic treated *P. ostreatus* vs. control and diabetic groups (Table 2) and (Figure 1 and 2).

**Effect of *P. ostreatus* extracts on blood glucose levels of rats**

Induction of diabetes mellitus was confirmed by blood glucose value above 250 mg/dL. The streptozotocin-induced diabetic rats showed consistent fasting hyperglycemia throughout the study. As shown in result section, *P. ostreatus* treatment to diabetic rats significantly reduced the blood glucose level. Diabetic animals also showed signs of polyuria, polydipsia and polyphagia.

**Impression of *P. ostreatus* extracts on the oxidative stress parameters in hepatic and pancreatic tissue**

Malondialdehyde (MDA) level in hepatic and pancreatic tissues showed markable increase in the Diabetic group whereas at the same time there was tremendous decrease in antioxidant level (GSH) and antioxidant enzyme activity (SOD, CAT, GSH-Px) when compared to healthy control (P<0.01). *P. ostreatus* treatment and standard glibenciclamide treatment inhibited the formation of MDA and raised antioxidant hormone (GSH) and antioxidant enzyme activity ((SOD, CAT, GSH-Px) (Table 3).

**Effect of *P. ostreatus* on immunohistochemical staining of iNOS**

Histopathological analysis of the harvested pancreas of streptozotocin induced diabetic mice was fixed in 10% formalin for 48 h, revealed remarkable changes versus the control mice. These changes included periportal fatty infiltration with focal necrosis of hepatocytes. The normal healthy control pancreatic tissue sections showed immunoreactive B cells (Figures 3 and 4). Diabetic mice treated with *P. ostreatus* extract (Figure 5) revealed a remarkable improvement of hepatic tissues where reduced necrosis was observed and the cellular arrangement of hepatocytes were found to be normal. Treatment with *P. ostreatus* extract brought back the cellular arrangement to near normal.

**Discussion**

The current research validated that the administration of *P. ostreatus* improves the performance in sociability, novel object recognition, water maze tasks, simple visual task and reactive oxygen and relative stress parameters such as lipid peroxidation and antioxidant activity.

**Table 2: Acute toxicity Studies of Pleurotus ostreatus Extract**

<table>
<thead>
<tr>
<th>Factors Analyzed</th>
<th>50 mg/kg</th>
<th>100 mg/kg</th>
<th>150 mg/kg</th>
<th>200 mg/kg</th>
<th>250 mg/kg</th>
<th>300 mg/kg</th>
<th>350 mg/kg</th>
<th>400 mg/kg</th>
<th>450 mg/kg</th>
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<tr>
<td>Changes on Mucous membrane</td>
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<td>+</td>
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</table>
Figure 2: Effect of Pleurotus ostreatus on the Behaviour of experimental animals

***Significantly different from control and diabetic group at P<0.5

Table 3: Effect of Pleurotus ostreatus extract on the oxidative stress parameters in Hepatic and Pancreatic Tissue

<table>
<thead>
<tr>
<th>PARAMETERS</th>
<th>C</th>
<th>C+D</th>
<th>C+E</th>
<th>D+E</th>
<th>D+G</th>
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<tr>
<td><strong>Liver</strong></td>
<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>GSH (µg/mg protein)</td>
<td>512.36±91.25</td>
<td>363.72±52.3</td>
<td>529.36±100**</td>
<td>451±89.44***</td>
<td>482.09±106.30</td>
</tr>
<tr>
<td>CAT (U/mg Protein)</td>
<td>360.6±18.36</td>
<td>318.29±61.2</td>
<td>368.23±22.8**</td>
<td>322±17.55***</td>
<td>344.8±28.91</td>
</tr>
<tr>
<td>SOD(U/mg Protein)</td>
<td>540±24.21</td>
<td>214.72±41.3</td>
<td>593.72±31.7**</td>
<td>412±35.33***</td>
<td>523.36±36.6</td>
</tr>
<tr>
<td>GSH-PX(U/mg Protein)</td>
<td>301±100</td>
<td>2149±638</td>
<td>3162±37.23**</td>
<td>2769±252***</td>
<td>2808±212</td>
</tr>
<tr>
<td>MDA(Mol/mg Protein)</td>
<td>8.05±0.98</td>
<td>18.12±1.92</td>
<td>9.01±1.02**</td>
<td>10.01±1.01***</td>
<td>9.93±1.52</td>
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<tr>
<td><strong>Pancreas</strong></td>
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<tr>
<td>GSH (µg/mg protein)</td>
<td>456.03±71.36</td>
<td>313.08±52.3</td>
<td>493.12±51.3**</td>
<td>360±57***</td>
<td>406.16±56.38</td>
</tr>
<tr>
<td>CAT (U/mg Protein)</td>
<td>278.02±17.62</td>
<td>159.13±13.32</td>
<td>286.33±28.3**</td>
<td>210±23***</td>
<td>274.30±21.94</td>
</tr>
<tr>
<td>SOD(U/mg Protein)</td>
<td>396±28.73</td>
<td>186.78±43.65</td>
<td>386.31±31.11***</td>
<td>349±37.5***</td>
<td>364.52±40.23</td>
</tr>
<tr>
<td>GSH-PX(U/mg Protein)</td>
<td>636±92.73</td>
<td>431±78</td>
<td>703.12±96**</td>
<td>598±94***</td>
<td>530±68</td>
</tr>
<tr>
<td>MDA(Mol/mg Protein)</td>
<td>7.33±1.08</td>
<td>16.13±1.36</td>
<td>7.50±19**</td>
<td>10.01±1.61***</td>
<td>8.23±1.4</td>
</tr>
</tbody>
</table>
of the *P. ostreatus* extract in blood correlates with the MWM score. Thus, *P. ostreatus* may be involved in protecting against neuronal degenerative disorders and ROS. In this study, (30 days) administration of *P. ostreatus* decreased the plasma oxidative status. An increase in the production of free radicals exacerbates the neurodegenerative process by deteriorating cellular enzymes [38]. *P. ostreatus* extract activated the antioxidant enzymes [39] when compared to the effects of essential oil of *Satureja khuzestanica* on the oxidative stress in experimental hyperthyroid male rat [40]. Complete oxidative stress can be controlled by long-term intake of mushrooms [41-42]. Memory deficits contributed by ROS and cytokines released from activated microglia and astrocytes [43,44]. Bindhu et al. reported that the *P. ostreatus* reduces the elevated blood glucose level in both type 1 and type 2 diabetic animals [45]. *Pleurotus* mushrooms enhances basal and insulin-roused glucose uptake [46], inhibit intestinal glucose uptake by sodium-dependent glucose transporter SGLT1 and mimic insulin by decreasing the countenance of genes that control gluconeogenesis [47]. Consumption of aqueous and hydroalcoholic extracts of *B. vulgaris* for 30 consecutive days as dose of 200 mg/kg can beneficially affect the hepatic liver enzymes, especially ALT, due to the liver regeneration and providing the source of the liver enzyme [48]. The intake of *P. ostreatus* caused a momentous rise in body weight and reduction in food and water intake in diabetic rats. This could possibly improve glycemic control produced by *P. ostreatus* in diabetic rats [49].

The increase in thio barbituric acid-reactive substances (TBARS), an index of lipid peroxidation in the diabetic rats might be due to increased levels of oxygen free radicals. In animal studies, mushroom polyphenol administration was shown to decrease serum TBARS level due to its potential antioxidant activity [50]. Similarly, *T. foenum-graecum* seeds are more potent than *G. officinalis* leaves with regards to hypoglycaemic properties, but body weight-reducing properties were similar between these two plant species. These results indicated that these two plant species can be used as an herbal treatment for DM and weight gain [51]. Behavioral protocol and increase in task complexity as accentuated due to performance deficits in diabetic rats. This protocol was depicted by study conducted in STZ-diabetic mice for normal acquisition and relatively simple tasks [52]. Oral and enema forms of hydroalcoholic extract of *Calendula. G. officinalis* can be offered as are potential therapeutic agents for Ulcerative
colitis (UC) induced in rats and even oxidative stress parameters was also reversed by CO administration [53]. The accentuation of the deficits by variations in the behavioral protocol may reflect an inability of the diabetic rats to adapt their behavioral strategies to the MWM [54]. S. khuzestanica have a lipid-lowering and antihyperglycemic property in male rats as an animal model for human [55]. Additionally STZ did not induce toxic influences on brain. Neurodegenerative diseases for instance AD which is allied to oxidative damage and neurotoxicity could be treated with the prophylactic means of P. ostreatus. Aторвастатин can decrease the histopathological lesions especially in liver, if used alone or in combination with T. vulgaris extract [56]. The use of Carum carvi L. (caraway) hydroalcoholic extract (CHE) in topical form may be associated with reduced intensity of oral mucositis OM. This may be due to appropriate antibacterial activity and terpinene contents [57]. All these above research proves P. ostreatus plays a vital role in controlling oxidative stress.

Long-term administration of an amount of P. ostreatus that could prompt antioxidative effects might probably deal with age-related declines in memory and learning ability in humans. In the process of aging, LPO accumulates and induces disorders of cellular functions [58]. Memory–related learning ability declines with aging [59]. Therefore, the continuous intake of, P. ostreatus might promote healthy aging of the brain in older individual.

**Financial & competing interest’s disclosure**

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