

The efficacy of herbal products as antioxidant in cushioning the effects of short-term exposure of female rats to low dose of environmental toxicity

Uchewa O^{*1} & Ezugworie JO²

¹Department of Anatomy, Faculty of Basic medical Sciences, Federal University Ndufu-Alike Ikwo (FUNAI), Ebonyi State, Nigeria

²Department of Anatomy, Faculty of Basic medical Sciences, University of Nigeria Enugu Campus (UNEC), Enugu State, Nigeria

*Author for Correspondence:
euchewa1@gmail.com

Introduction: Natural products can be more active than we consider them. Some herbs used by man can boost the body's antioxidant level. This herb that serves as vegetable have also been identified as a cure some illnesses and also restore the integrity of the organs of the body.

Objective: This research investigated the efficacy of as a natural antioxidant for reducing toxicity.

Materials and Methods: Twenty five (25) rats with average weight of 145 g were randomly assigned into five groups and received standard rat's diet and allowed water ad libitum. Group A (negative control) received normal saline. Group B and C received Lead acetate solution (1.5 mg/kg) in addition to the extract serving as low and high dose groups respectively. Group D (positive control) received Lead acetate solution only while group E was given ethanolic extract only to ascertain if it has any deleterious effects.

Results: The animals showed an increase in both the body, uterine and ovarian weight when compared to group D. The SOD activity reduced in both B and C while increasing in D but E had a normal SOD level. On examination of the ovary and uterus microscopically, the alterations seen in the photomicrophic plates includes damages and reduced folliculogenesis with a marked increase in the number of atretic follicles, oedema, necrotic zones, etc. These changes were restored to near normal following the administration of the extract.

Conclusion: The finding of this research shows that natural products are capable of removing oxidative radical produced in the body thereby detoxifying the body and reproductive organs.

Keywords: antioxidant • toxicity • ogbu • infertility • ovary

Introduction

The female reproductive system and human fertility is adversely affected by exposure to environmental toxicants. In this regard, much attention has been paid to toxic environmental factors that cause ovarian toxicity and there have been some attentions on toxic environmental factors that cause both ovarian and uterine toxicity [1-3]. According to Sullivan [4] "we know of relatively few chemicals that have adverse effects on reproduction but we are ignorant of more than 90% of industrial chemicals to which people are exposed". Reproductive toxicity is the adverse effects of chemicals on gonadal structures and functions, alterations

in fertility and impaired gametes functions [5-8].

Infertility as a common problem affects perhaps one in every six couples [9]. A number of studies have suggested that male problems is the commonest single known cause of infertility even though diagnostic problems make it difficult to establish the extent to which the females are involved [10]. The use of plant extract as fertility booster in human is now on the increase because of the shifting of the attention from synthetic drugs to natural plant products [11]. Accessibility, availability and affordability are some of the factors that have prompted over 80% of the populace in the developing countries to

continue using medicinal plant products in handling primary medical problems [12].

In the African continent, various plants have been identified to pose fertility regulating properties and quite a few of them have been tested and proven for such effects [13-16]. This informed the need to test *F. vogelii* a woody tree or shrub in the family Moraceae [17] in treating the reproductive organs. This tree is called "figs tree" in English. The fruit of most other species are also edible though they are usually of only local economic importance or eaten as bush food [18]. The leave of *F. vogelii* is used locally as vegetables. The Igbo refers to *F. vogelii* as ogbu. This research aims at investigating the role of *F. vogelii* as an antioxidant in reducing toxicity.

Materials and Methods

Plant collection and preparation of the extracts

The plant material was collected from Enyibichiri Ndufu-Alike in Ikwo Local Government Area of Ebonyi State where it grows in abundant and used by the inhabitants as vegetable. The leaves were washed and dried in ventilated room. Thereafter, it was crushed into powder using pestle and mortar and passed through mesh sieve to get the fine powder which was divided into portions 'A' and 'B'. Portion 'A', five hundred grams (500 g) of the powder was weighed using an electronic weighing balance and soaked in 1200 mL of ethanol (powder/solvent) respectively. The mixture was agitated using an electric blender to enhance proper mixing of the solvent with the powder and then poured into air-tight plastic container. The container, with the mixture was kept in the refrigerator for 48 hours. The mixture was filtered, first, with cheese cloth, and then with Whatman No 1 filters paper (24 cm). The filtrates were separated and concentrated in vacuum using Rotary Evaporator to 10% of their original volumes at 37°C-40°C. This was heated to dryness using water bath. This was stored until it was required for use.

Phytochemical screening

The phytochemical screening was performed using 500 g of the powder representing portion 'B' of the extract. This portion was subjected to quantitative phytochemical screening using the method described by Prashant, et al. [19].

Ethical approval

The ethical approval for this work was sorted and obtained from the Ethical Committee of the Faculty of Basic Medical Sciences, University of Nigeria Nsukka, Enugu Campus (UNEC). There was a strict adherence to International Guidelines for the use of animals in research studies based on European Commission Directive on the use of animals [20].

Experimental design and animal grouping

There twenty five (25) female Wistar rats with average weight of 145 g was procured and maintained in the Animal House of Anatomy Department of Faculty of Basic Medical Sciences of the University of Nigeria, Enugu Campus (UNEC). The animals were housed in netted cages, fed with grower's mash and allowed water ad libitum with acclimatization period of one week (7 days) was observed. There were randomly assigned into five groups (A, B, C, D and E). The rats received standard rat's diet containing 0.5% NaCl, 16.8% protein and 46.5% dietary fat, 35.7%, vitamins, other micro nutrients and tap water ad libitum [21], throughout the period of the study. Group A served as negative control and received normal saline to expose them to the same stress as the other groups. Groups B and C received 1.5 mg/kg Lead acetate solution daily [22] and 24 hours later ethanolic extract of *F. vogelii* was administered. Group B served as the low dose group and received 100 mg/kg of the ethanolic extract while C served as the high dose group and received 300 mg/kg of the ethanolic extract. Group D served as the positive control group and received Lead acetate solution. The rats in this group received 1.5 mg/kg of Lead acetate solution [23]. Group E received only the aqueous extract of *F. vogelii* and it serves to ascertain if the extract has any deleterious effects in the rats (**FIGURE 1**).

Ovarian SOD activities

After the experiment one of the ovaries was harvested and homogenized with mortar and pestles [24] and then preserved with phosphate buffer. The preserved homogenate was centrifuged and kept under refrigeration overnight before it was sent to a laboratory for antioxidant check. Superoxide dismutase activity was measured according to the method of Rukmini et al. [25] as described by Winterboun et al. [26]. The principle of the assay was based on

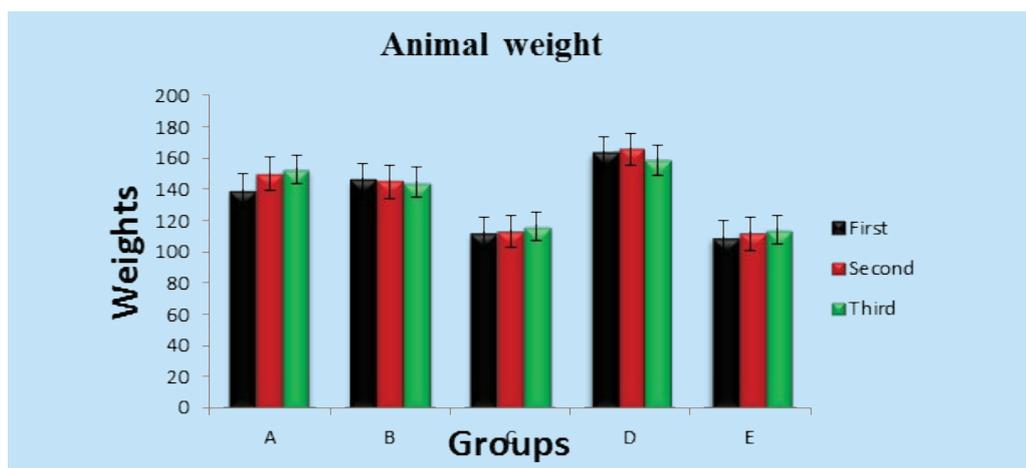


Figure 1. Change in the weight of the animals during the experimental period

the ability of SOD to inhibit the reduction of nitro-blue tetrazolium (NBT). Briefly, the reaction mixture contained 2.7 ml of 0.067 M phosphate buffer, pH 7.8, 0.05 ml of 0.12 mM riboflavin, 0.1 ml of 1.5 mM NBT, 0.05 ml of 0.01 M methionine and 0.1 ml of enzyme samples. Uniform illumination of the tubes was ensured by placing it in air aluminum foil in a box with a 15 W fluorescent lamp for 10 minutes. Control without the enzyme source was included. The absorbance was measured at 560 nm [27]. One unit of SOD was defined as the amount of enzyme required to inhibit the reduction of NBT by 50% under the specific conditions.

Histological study

At the end of the experiment, the rats were starved overnight and anaesthetized with chloroform and then decapitated by Schoenwolf et al. [28]. The animals were sacrificed, dissected then the uterus and ovary removed, weighed and quickly fixed in bouin's fluid for routine histological procedures. The tissues were processed and embedded in paraffin wax to obtain thin sections.

Statistical analysis

The Data were analyzed using descriptive statistics and presented as mean \pm standard error of mean (SEM). Inferential statistics of paired student's t-Test was adopted and the statistical significance level was established at a value of $P \leq 0.05$ (*) or $P \leq 0.01$ (**) with the aid of statistical software package (SPSS) version 20.

Results

The results such as weights, SOD activities,

and the histological studies are summarized below in tabular form, charts and photomicrographs.

TABLE 1 shows weight change of the animals that received aqueous extract of *F. vogelii*. During the experiment, group A showed significant increase in weight at $P \leq 0.05$ and $P \leq 0.01$ in second and third weeks respectively. Group D recorded weight loss in the second week but a significant reduction in third week. There was a progressive weight loss in group B but only significant in third week.

TABLE 2 below shows the mean weight of the uterus in both the control and experimental groups. Group D when compared to group A recorded a significant weight loss in their uterine weight as shown in the table below. All the other groups when compared to group D (positive control) showed a statistically significant uterine weight gain (**FIGURE 2**).

TABLE 3 below shows the mean weight of the ovary in the control and experimental groups. As shown, group D (positive control) recorded a significant ovarian weight loss ($P \leq 0.01$) when compared to normal control (group A). All the other groups when compared to group D (positive control) showed a statistically significant ($P \leq 0.01$) ovarian weight gain (**FIGURE 3**).

Ovarian SOD activities

The SOD results as presented in **TABLE 4** below shows that positive control (group D) have a significant increase in SOD activity when compared to the negative control (group A). The curative groups showed reduction in SOD activity level but not

Table 1. Table showing the weight (g) of animals that received aqueous extract of *F. vogelii* and lead acetate solution during the period of the research

S.No	Groups	No of Animals	Weeks (Mean ± SEM)		
			First	Second	Third
1	A	5	139.25 ± 12.07	150.05 ± 12.52*	152.83 ± 13.85**
2	B	5	145.90 ± 12.91	144.78 ± 16.20+	144.60 ± 16.12+
3	C	5	111.78 ± 5.50	113.05 ± 9.97**	116.03 ± 9.02**
4	D	5	163.65 ± 12.15	165.55 ± 5.14	158.78 ± 5.71+
5	E	5	109.13 ± 0.87	111.75 ± 0.66	114.13 ± 0.96

*significant increase in weight at $P \leq 0.05$
 **significant increase in weight at $P \leq 0.01$
 +significant reduction in weight at $P \leq 0.01$
 S.No=Serial number

Table 2. Mean weight (mg) of uterus of control and experimental groups of adult female Wistar rats

S.No	Groups	No. of Animals	Mean ± SEM (mg)
1	A	5	521.00 ± 0.50
2	B	5	511.71 ± 1.50*
3	C	5	510.13 ± 5.00*
4	D	5	463.50 ± 5.00**
5	E	5	527.11 ± 5.00****

**Significant reduction in uterine weight when compared to A ($P \leq 0.01$ and $P \leq 0.05$)
 *Significant uterine weight gain when compared to D ($P \leq 0.01$)
 ***Significant uterine weight gain when compared to D ($P \leq 0.01$)
 **** Significant uterine weight gain when compared to A ($P \leq 0.01$)
 S.No=Serial number

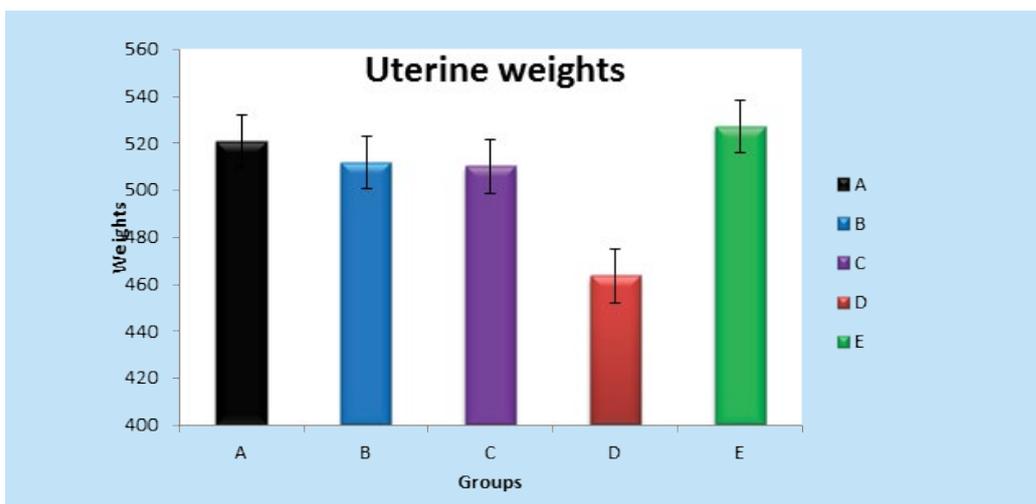


Figure 2. The Uterine weight (mg) of the control and experimental groups

statistically significant when compared to the positive control (group D).

Phytochemical analysis

The following chemicals were screened for and the results are as presented below in **TABLE 5.**

Histological studies

The main histological alterations observed in female rat ovaries and uterus after inducing toxicity with lead acetate solution and treating with aqueous and ethanolic extracts of *F. vogelii* are presented below as they appear in H&E stain, 60X and 150X.

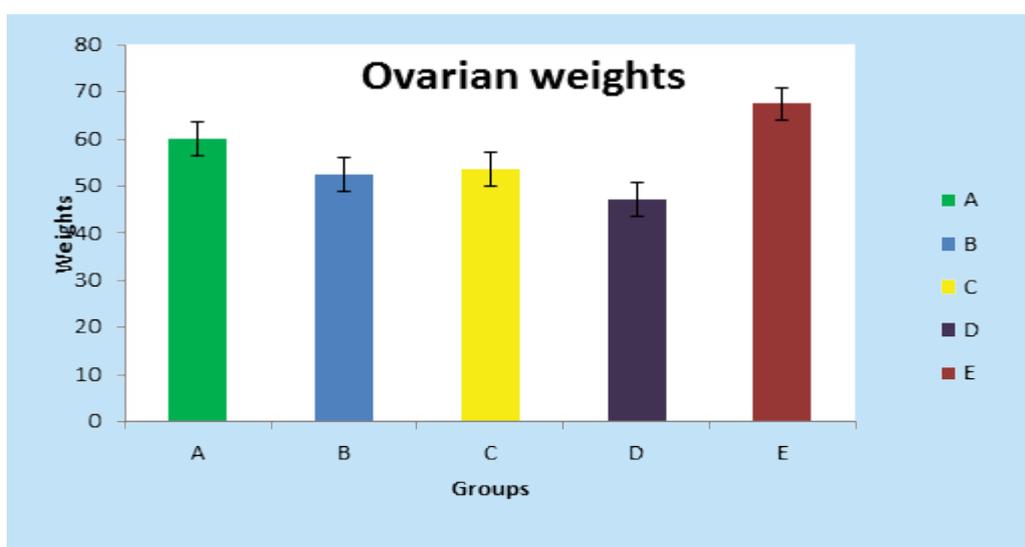
Table 3. Mean weight (mg) of Ovaries of control and experimental groups of adult female Wistar rats

S.No	Groups	No. of Animals	Mean \pm SEM (mg)
1	A	5	60.10 \pm 0.50
2	B	5	52.40 \pm 0.50d
3	C	5	53.50 \pm 0.50d
4	D	5	47.10 \pm 2.00**
5	E	5	67.43 \pm 0.51***

*** Significant increase in ovarian weight when compared to group A ($P \leq 0.01$)

** Significant reduction in ovarian weight when compared to group A ($P \leq 0.01$) d Significant increase in ovarian weight when compared to group D ($P \leq 0.01$)

S.No - Serial Number

**Figure 3.** The Ovarian weight of the control and experimental groups**Table 4. Activities of ovarian enzyme Superoxide Dismutase (SOD) of the experimental**

S.No	Groups	No. of animals	Mean \pm SEM ($\mu\text{g}/\text{mg}$)
1	A	5	20.00 \pm 0.38
2	B	5	26.00 \pm 1.77
3	C	5	26.89 \pm 1.73
4	D	5	27.15 \pm 5.22***
5	E	5	21.16 \pm 0.60

***Significant increase in SOD activity when compared to group A ($P \leq 0.01$ and $P \leq 0.05$)

** Significant reduction in SOD activity when compared to group D ($P \leq 0.01$)

S.No-Serial Number

Discussion

The wake of 21st century gave rise to the recognition of herbal medicine more than ever and *F. vogelii* is a vegetable that has been used locally to treat some illnesses and also implicated by so many researchers as an herbal remedy for some human diseases [29]. The positive change in body weight in any research is a useful indicator of favourable effect of herbal medicine [30,31] and is also an important constituent in the study of safety of any therapeutic agent [32].

In this research, the animals in negative group recorded a significant increase in body weight in the second and third week at $P \leq 0.01$ and $P \leq 0.05$ respectively while lead acetate group showed a little weight increase in the second week and a significant weight loss in the third week agreeing with Dumetrescu et al. [33]. The little increase in weight might be due to the fact that lead needed time to cause enough damage that could lead to weight loss, while the subsequent weight loss could have resulted from the deleterious

Table 5. Phytochemical composition of ethanolic extracts of dried leave of the extract

S.No	Parameter	Present	Absent	Moderate	High
1	Saponins				***
2	Alkaloids	*			
3	Phlobatannins		-		
4	Flavonoids				***
5	Coumarins			**	
6	Steroids			**	
7	Reducing Sugar		-		
8	Triterpenoids	*			
9	Flavons		-		
10	Carotenoids			**	
11	Polyphenols				***
12	Xathoxyllines		-		
13	Cardiac glycosides	*			

* = Indicates the presence of phytochemicals
 - = Indicates the absence of phytochemicals
 ** = Shows moderate concentration level of phytochemicals
 *** = Shows high concentration level of phytochemicals

effect of lead on the organs at this stage. Among the experimental groups some showed a significant increase in body weight at $P \leq 0.05$ in the second week while others showed significant increase at $P \leq 0.01$ in the third week. The increase in weight is one of the major pointers that the leave of *F. vogelii* has a nutritive and therapeutic value. The significant ($P \leq 0.01$ and $P \leq 0.05$) decrease in the uterine and ovarian weights of the animals in positive control group agrees. The above findings agree with the report of Durgesh and Lata, [34] where they stated that "the animals that received only lead acetate experienced a significant reduction in uterine weight." The extract group only recorded a significant ($P \leq 0.01$) uterine and ovarian weight gain compared to the negative control group. This organ weight gain could be an insight into the therapeutic benefits of extract. As shown in **TABLE 4** above, positive control group showed a significant ($P < 0.01$) increase in superoxide dismutase (SOD) assumed to be as a result of the inflammation caused by the induced toxicity compared to the negative control group. The rats treated with the extract recorded a decrease in ovarian SOD when compared to that of the positive control group. The group that received only the extract showed SOD closer to the negative control which might be a pointer that *F.vogelii* is a good antioxidant capable of removing free radicals produced due to inflammation by the presence of toxins. These free radicals cause SOD level in the body

to rise above normal [35] (**FIGURE 4**).

After a quantitative phytochemical screening of the ethanolic extract, the following chemicals were present: saponins, flavonoids, alkaloids, triterpenoids, polyphenols, coumarins, steroids, carotenoids and cardiac glycosides. Various phytosteroids (plant steroids) have been shown to promote fertility [36,37]. Flavonoids present in the leave of plants have been shown to possess many pharmacological properties such as: anti-oxidant activities, anti-inflammatory activities, anti-cancer activities and anti-microbial effects. This is in agreement with the work done by Uche and Obianime [38]. Flavonoids might have contributed to the extracts healing (anti-inflammatory) properties as observed from the histological studies [39,40]. *F. vogelii* is very rich in triterpenoids, a compound which plays a very active role in wound healing [41]. According to Krishnaiah et al. [42], triterpenoids are shown to strengthen the skin, increase the concentration of antioxidants in wounds, and restore inflamed tissues by increasing blood supply (**FIGURES 5-8**).

Microscopical examinations of various parts of the ovary and uterus for histological alterations showed numerous damages and reduction in the formation of follicles with a marked increase in the number of atretic follicles which is in agreement with Igile et al. [29], Patrick [43] and Junaid et al. [44]. The major structural changes in the organs

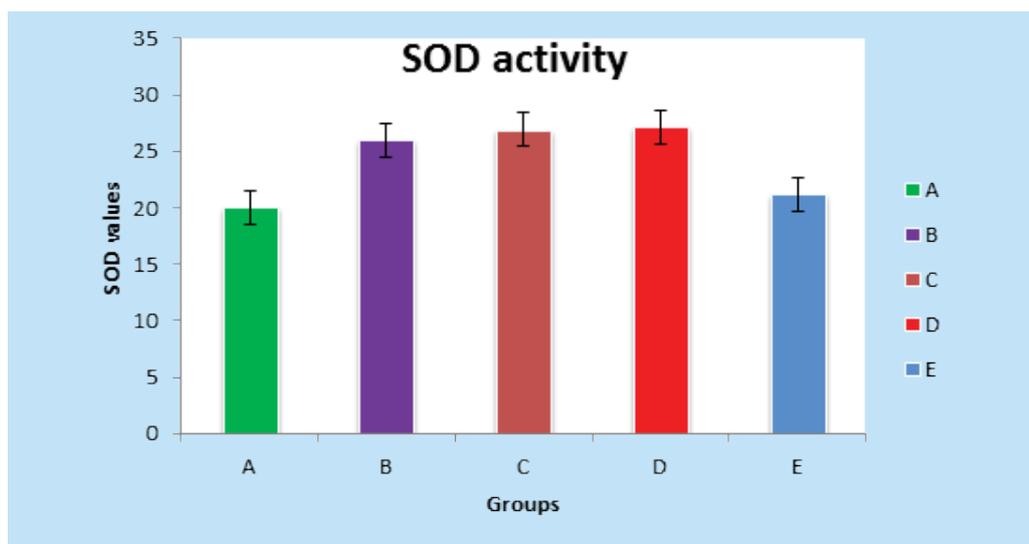


Figure 4. The MEAN \pm SEM antioxidant activity of the Superoxide Dismutase (SOD) enzyme of the control and experimental groups.

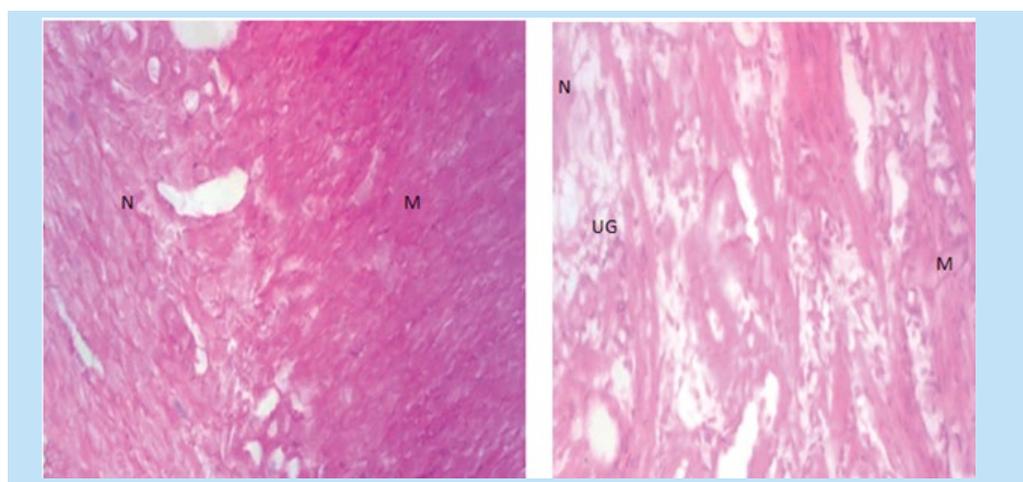


Figure 5. Uterus from the group A (negative control) reveals the presence of N: Normal mucosa structure, M: Myometrium and UG: uterine glands. H&E 100X and 200X

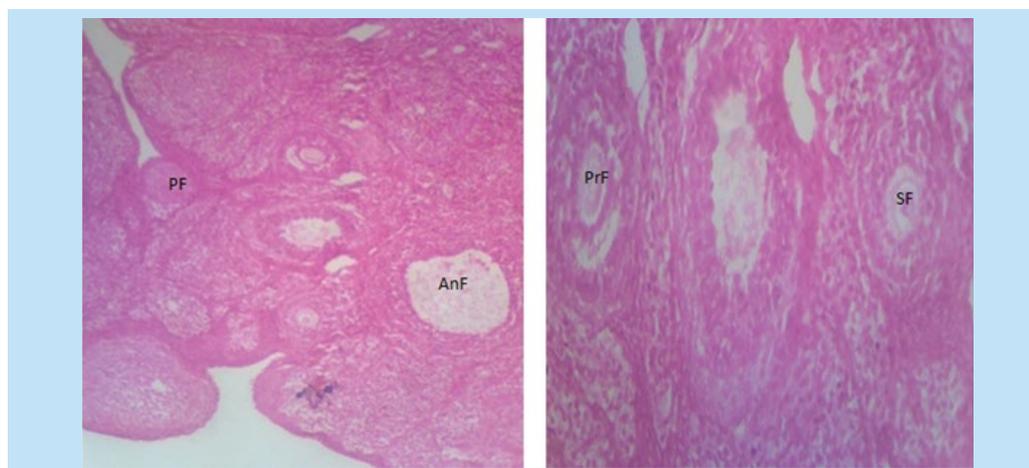


Figure 6. Ovaries from negative control group showing ovarian follicles at different stages of development, PF: Primordial follicles, AnF: Antral follicles, PrF: Primary follicle and SF: Secondary follicle; H&E 100X and 200X

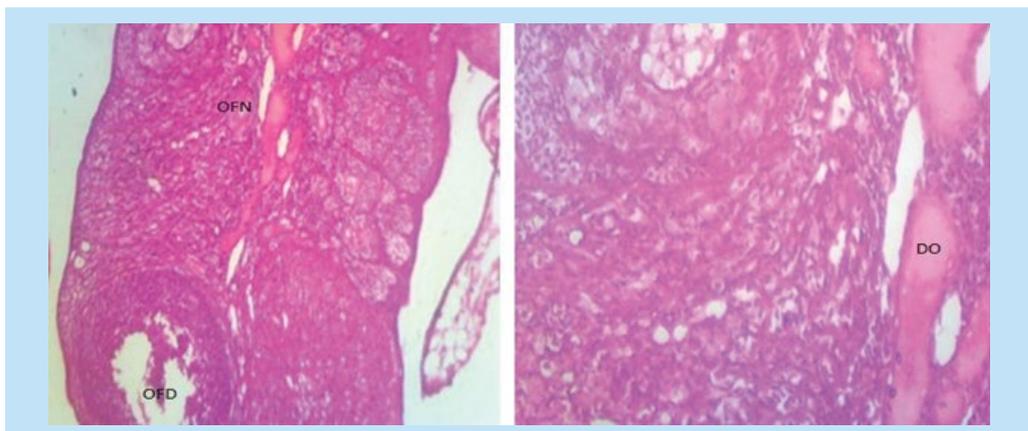


Figure 7. Group B Ovary showing PF: Primordial follicle, DO: Diffused oedema, OES: Optical empty spaces and F: follicles; H&E. 100X and 200X

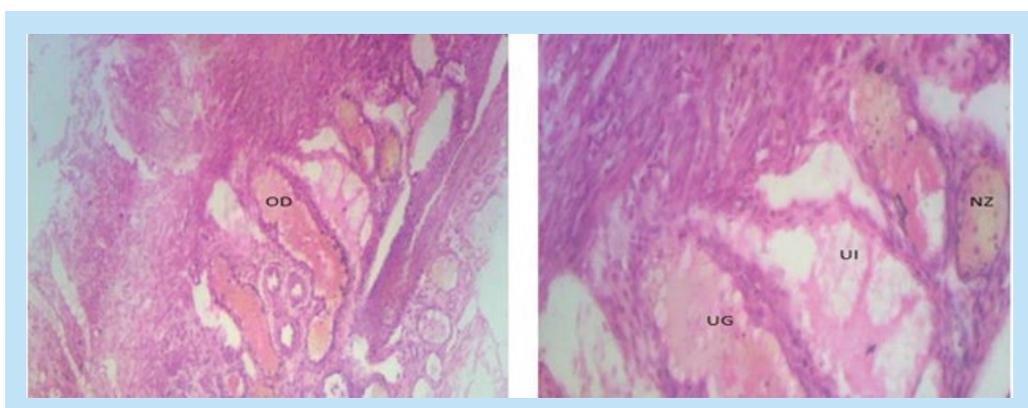


Figure 8. Group B showing OD: Edema, UI: Uterine injury and UG: uterine glands, H&E 100X and 200X

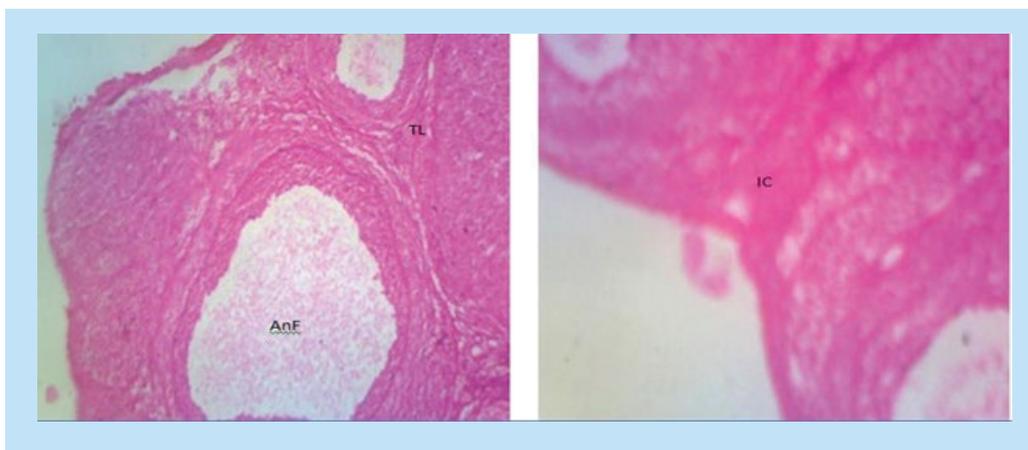


Figure 9. Group C Ovary showing TL: theca lutea, AnF: Antral follicle and IC: interstitial cells; H&E 100X and 200X.

(ovaries and uterus) include diffuse oedema, necrosis, optical empty spaces, denudation, folliculogenesis and atretic follicles in accordance with Durgesh and Lata, [34], Taupeau et al. [45] and Dumetrescu et al. [46]. The organs also looked more vascularized the extract groups which indicated a restoration in the blood supply to the organs that was lost due to the lead induced toxicity. According to Shah et al. [47], oral

administration of high doses of lead, leads to reduced number of ovarian follicles and an increased number of atretic follicles as was also observed in this study. The effects of lead on reproductive systems of female rats are complex as seen in this study. The uterus of animals in negative control group presented a normal endometrium, intact uterine glands and unaltered stroma in agreement with Dumetrescu et al. [46] report of 2005. The

ovaries also showed normal histoarchitecture with ovarian follicles at various stages of development. Ovaries of the animals that received only lead acetate presented a fatty change (a reversible change that occurs in organs due to toxicity) appearance and focal loss of tissue (optical empty spaces) [46]. This fatty change occurred in the organ due to toxicity. The damage might be as a result of the lead effect on blood vessels leading to tissue necrosis. Lead also interferes with several enzymatic steps in the haem pathway [48]. The above changes were also reported by Dumitrescu et al. [33] and Dumitrescu et al. [46]. The animals in experimental groups showed particularly an interesting observation about the dynamics of reactions to the presence of extract in their systems. The ovary of low dose of the extract showed some developing follicles with evidence of lead poisoning shown by fatty changes. The uterus has little damaged areas which might

be due to the healing effect of the extract in the body against the toxicity induced by the lead acetate [46]. The antioxidant property of the extract is implicated in the control or prevention of inflammation. The granuloma (a disease condition due to inflammation or infections) formed is not pronounced. The higher dose engendered folliculogenesis as evidenced in the ovaries. The oedema and denudations observed were not as prominent as in the low dose group. The uterine glands were better healed than those in the low dose group. There was a marked increase in the perfusion of the organs (ovary and uterus) which might be due to the extract's ability to restore the blood supply to the organs (**FIGURES 9-12**).

In summary, our results demonstrated that of *F. vogelii* leaves are very good as an alternative herbal medicine. Reproductive toxicity of

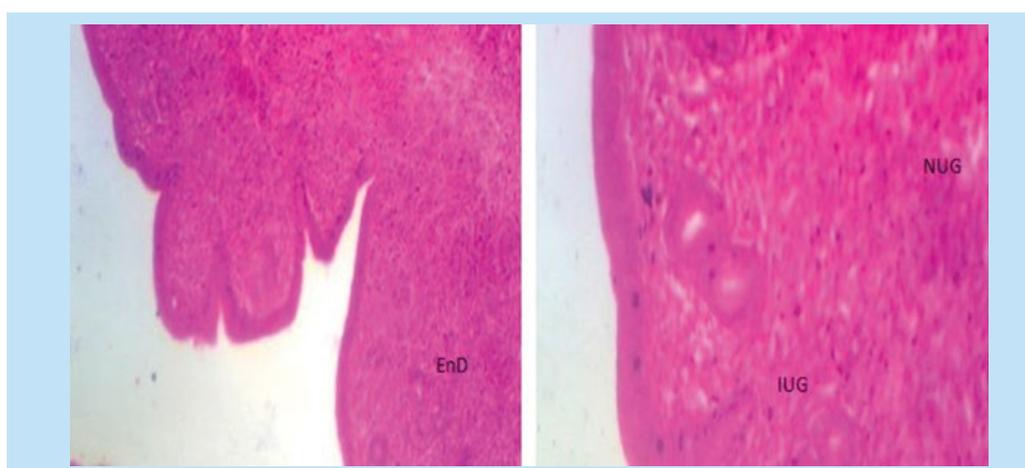


Figure 10. Group C Uterus showing EnD: Endometrial denudation, NUG: Necrotic uterine glands and IUG: Invaginated uterine gland; H&E 100X and 200X

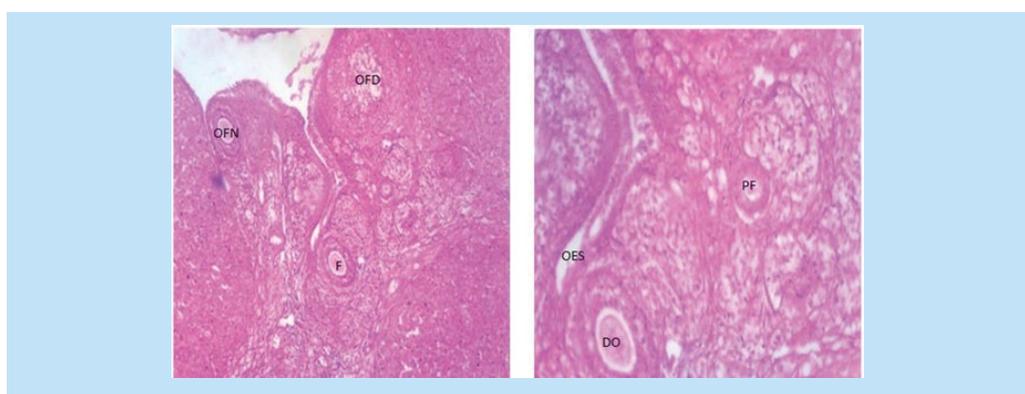


Figure 11. Group D Ovary showing OES: optical empty spaces; PF: primordial ovarian follicles; OFD: ovarian follicle denudation; DO: diffuse oedemas; OFN: Ovarian follicles necrosis; F: Follicles at different stages of development; H&E 100X and 200X.

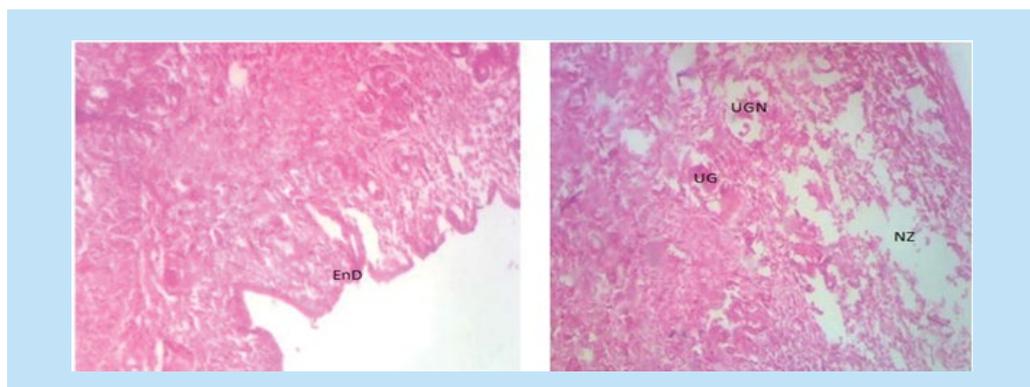


Figure 12. Group D Uterus showing NZ: Necrotic zone, UG: Uterine glands, UGN: Necrotic Uterine Gland and EnD: endometrial denudation; H&E 100X and 200X.

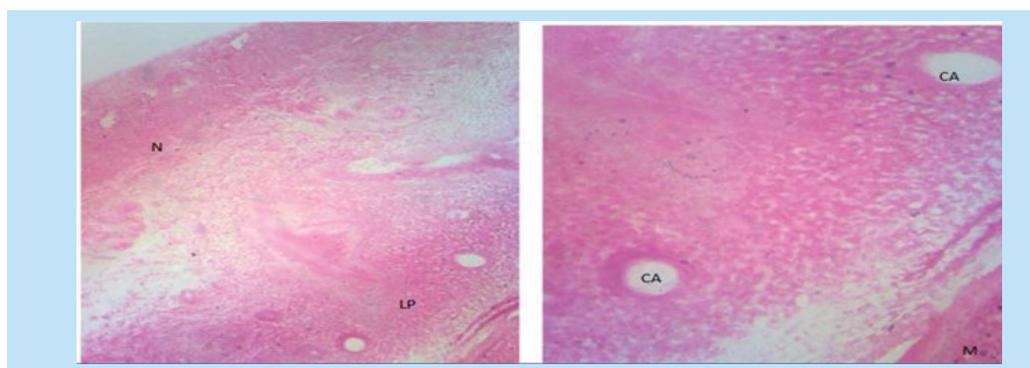


Figure 13. Group F uterus showing N: Normal mucosa, LP: Lamina propria, CA: Coiled arteries and M: Myometrium; H&E 100X and 200X.

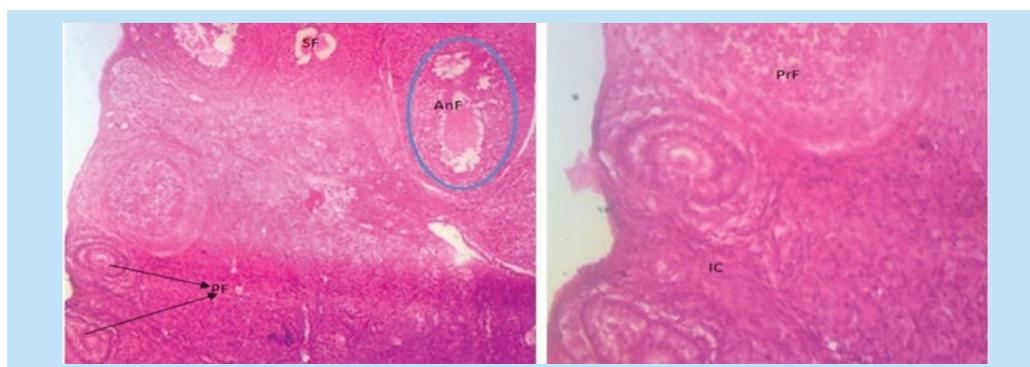


Figure 14. Group F ovary showing SF: Secondary follicle, AnF: Antral follicle, PF: primordial follicles and IC: Interstitial cells; H&E 100X and 200X.

lead in female rats can be reversed to normal using this extract and there was in the free radicals by the antioxidant by interfering with the reactive oxygen species produced by lead toxicity as evidenced by the reduction in the ovarian SOD in **TABLE 4** above. We also discovered that the extract suppressed and healed the damages caused on the tissues confirming its effectiveness in the treatment of reproductive organ alterations (**FIGURES 13 and 14**).

References

1. Hruska KS, Furth PA, Seifer DB *et al.*

Environmental factors in infertility. *Clin. Obstet. Gynecol.* 43(4), 821-829 (2000).

2. Brunton LL, Goodman LS, Blumenthal D *et al.* Principles of toxicology. McGraw-Hill Professional (2007).

3. Casarett LJ, Doull J, Klaassen CD. Casarett and Doull's toxicology: the basic science of poisons. McGraw-Hill, New York (2008).

4. Sullivan FM. Impacts of the environment on reproduction from conception to parturition. *Env. Health. Perspec.* 101(2), 13-18 (1993).

5. Hodge AT. Roman aqueducts & water supply. Bristol Classical Press (2002).
6. Timbrell J. Introduction to toxicology. CRC Press (2001).
7. Brent J, Wallace K, Burkhart KK *et al.* Critical care toxicology: diagnosis and management of the critically poisoned patient. Gulf Professional Publishing (2005).
8. Kosnett MJ. Heavy metal intoxication and chelators. Basic and clinical pharmacology. McGraw-Hill, New York (2007).
9. Oladimeji SO, Igbalaye JO, Coleshowers CL. Immunological biomarkers determined in female rats administered with pro-fertility extract of *Anthocleista vogelii*. *J. Nat. Sci. Res.* 4(8), 113-122 (2014).
10. Van den Eede B. Investigation and treatment of infertile couples: ESHRE guidelines for good clinical and laboratory practice. *Hum. Reprod.* 10(5), 1246-1271 (1995).
11. Dada AA and Ajilore VO. Use of ethanol extracts of *Garcinia kola* as fertility enhancer in female catfish *Clarias gariepinus* broodstock. *Int. J. Fish. Aquacul.* 1(1), 005-010 (2009).
12. Watcho P, Ngadjui E, Alango NP *et al.* Reproductive effects of *Ficus asperifolia* (Moraceae) in female rats. *Afr. Health Sci.* 9(1), 49-53 (2009).
13. Baker DA, Hameed C, Tejani N *et al.* Lymphocyte subsets in women on low dose oral contraceptives. *Contraception* 32(4), 377-382 (1985).
14. Telefo PB, Moundipa PF, Tchouanguép FM. Oestrogenicity and effect on hepatic metabolism of the aqueous extract of the leaf mixture of *Aloe buettneri*, *Dicliptera verticillata*, *Hibiscus macranthus* and *Justicia insularis*. *Fitoterapia.* 73(6), 472-478 (2002).
15. Ganguly M, Borthakur M, Devi N *et al.* Antifertility activity of the methanolic leaf extract of *Cissampelos pareira* in female albino mice. *J. Ethnopharmacol.* 111(3), 688-691 (2007).
16. Cherdshewasart W, Nimsakul N. Clinical trial of *Butea supeba*, an alternative herbal treatment for erectile dysfunction. *Asian. J. Androl.* 5(3), 243-246 (2003).
17. Berg CC, Corner E.J.H. Moraceae. In: Flora Malesiana Ser. 17(2), (2005).
18. Egbuna PAC, Joshua PE, Chigbo MU. Anti-hepatotoxic effects of *ficus vogelii* ethanol leaf extract on the liver function indices of Ccl4-induced hepatotoxicity in rats. *J. Am. Sci.* 7(6), (2011).
19. Tiwari P, Kumar B, Kaur M *et al.* Phytochemical screening and extraction: a review. *Int. Pharm. Sci.* 1(1), 98-106 (2011).
20. European Commission. Directive 2010/63/EU of the European parliament and the council on the protection of animals used for scientific purposes. Brussels, Belgium (2010).
21. Krinke GJ. The laboratory rat. Academic press, New York (2002).
22. Dafalla HAA. Studies on the constituents of *Ficus capensis* (Thumb). *Pak. J. Soc. Sci.* 3(5), 751-754 (2005).
23. Benoff S, Auburn K, Marmar JL *et al.* Link between low-dose environmentally relevant cadmium exposures and asthenozoospermia in a rat model. *Fert. Ster.* 89(2), 73-79 (2008).
24. Akunna GG, Saalu LC, Ogunlade B *et al.* Consumption of bay leaf (a food spice) may be a safe and effective treatment for male infertility resulting from partial ligation of the left renal vein in wistar rat: study suggest. *Am. J. Res. Comm.* 1(3), 123-142 (2013).
25. Rukmini MS, D'souza B, D'souza V. Superoxide dismutase and catalase activities and their correlation with malondialdehyde in schizophrenic patients. *Ind. J. Clin. Biochem.* 19(2), 114 (2004).
26. Winterbourn CC, Hawkins RE, Brian M *et al.* The estimation of red cell superoxide dismutase activity. *J. Lab. Clin. Med.* 85(2), 337-341(1975).
27. Adenuga GA, Adegbesan BO, Adebayo OL. Antioxidant defense of zinc acetate supplementation on the brain of protein-undernourished rats. *Int. J. Biol. Chem. Sci.* 3(1), 152-155 (2009).
28. Schoenwolf GC, Bleyl SB, Brauer PR *et al.* Larsen's human embryology. Elsevier (2015).

29. Igile GO, Utin IC, Iwara IA *et al.* Ethanolic extract of *Ficus Vogelii* ameliorates dyslipidemia in diabetic albino wistar rats. *Int. J. Curr. Res. Biosci. Plant. Biol.* 2(8), 87-96 (2015).
30. OA, Al-Shabanah, El-Hadiyah TM *et al.* Effect of prolonged vigabatrin treatment on hematological and biochemical parameters in plasma, liver and kidney of Swiss albino mice. *Sci. Pharm.* 70(2), 135-145 (2002).
31. Alli LA, Oluwole OP, Aniah JA *et al.* Histopatologic effects of aqueous root extract of *Acacia nilotica* on rat liver, kidney and lungs. *J. Anatomical. Sci.* 6(1), 12-18 (2015).
32. OECD. The OECD guideline for testing of chemical usage in experiment. The organization of economic co-operation development. Paris (2001).
33. Dumitrescu E, Chiurciu V, Muselin F *et al.* Effects of long-term exposure of female rats to low levels of lead: ovary and uterus histological architecture changes. *Turkish. J. Biol.* 39(2), 284-289 (2015).
34. Sharma DN, Bhattacharya L. Role of Vitamin E on anti folliculogenesis effects of lead acetate on diameter of follicles containing ovarian tissue of swiss albino mice. *Global. J. Biol. Agri. Health. Sci.* 3(1), 322-325 (2014).
35. Sirisha N, Sreenivasulu M, Sangeeta K *et al.* Antioxidant properties of *Ficus* species-a review. *Int. J. Pharm.Tech. Res.* 2(4), 2174-2182 (2010).
36. Ruiz-Luna AC, Salazar S, Aspajo NJ *et al.* *Lepidium meyenii* (Maca) increases litter size in normal adult female mice. *Rep. Biol. Endocrinol.* 3(1), 16 (2005).
37. Iwara IA, Igile GO, Uboh FE *et al.* Hypoglycemic and hypolipidemic potentials of extract of *vernonia calvaona* on alloxan-induced diabetic albino wistar rats. *Eur. J. Med. Plants.* 8(2), 78-86 (2015).
38. Uche FI, Obianime AW. The effects of Vitamin E on Vanadium Pentoxide toxicity of the biochemical parameters of male guinea pigs. *J. Appl. Sci. Env. Manag.* 12(3), 101-105 (2008).
39. Okwu DE, Josiah C. Evaluation of the chemical composition of two nigerian medicinal plants. *Afr. J. Biotech.* 5(4), 357-361 (2006).
40. Uche FI, Obianime AW, Gogo-Abite M. Effects of vanadium pentoxide on the histological and sperm parameters of male guinea pigs. *J. Appl. Sci. Env. Manag.* 12(3), 107-115 (2008).
41. Sotheeswaran S, Doyle M, Aalbersberg W. Medicinal plants in the south pacific. WHO Regional Publications. Philippines (1998).
42. Krishnaiah D, Devi T, Bono A *et al.* Studies on phytochemical constituents of six Malaysian medicinal plants. *J. Med. Plants. Res.* 3(2), 067-072 (2009).
43. Patrick L. Lead toxicity part II: The role of free radical damage and the use of antioxidants in the pathology and treatment of lead toxicity. *Altern. Med. Rev.* 11, 114-127 (2006).
44. Junaid M, Chowdhuri DK, Saxena R. Lead-induced changes in ovarian follicular development and maturation in mice. *J. Toxicol. Env. Health.* 50(1), 31-40 (1997).
45. Taupeau C, Poupon J, Nomé F *et al.* Lead accumulation in the mouse ovary after treatment-induced follicular atresia. *Reprod. Toxicol.* 15(4), 385-391 (2001).
46. Dumitrescu E, Cristina RT, Muselin F. Reproductive biology study of dynamics of female sexual hormones: a 12-month exposure to lead acetate rat model. *Turk. J. Biol.* 38(5), 581-585 (2014).
47. Shah A, Mian M, Khan S *et al.* Correlation of blood lead levels with atresia of ovarian follicles of albino mice. *Ann. Pak. Inst. Med. Sci.* 4, 188-192 (2008).
48. <https://www.atsdr.cdc.gov/toxprofiles/TP.asp?id=96&tid=22>.