

Roles of galectins in chronic inflammatory microenvironments

**Greg Parsonage,
Emily Trebilcock,
Marta A Toscano,
Germán A Bianco,
Juan M Ilarregui,
Christopher D Buckley &
Gabriel A Rabinovich†**
*Hospital de Clínicas "José de San Martín", Rabinovich,
División Inmunogenética,
Facultad de Medicina,
Universidad de Buenos Aires,
Av. Córdoba 2351, 3
Piso (C1120), Ciudad de Buenos Aires, Argentina
Tel.: +54 115 950 8755;
Fax: +54 115 950 8758;
gabyrabi@ciudad.com.ar*

Lectins are multifunctional carbohydrate-binding proteins that can recognize various carbohydrates on cell surfaces and extracellular matrix, and are involved in several biological processes. Galectins, a family of animal lectins with affinity for β -galactoside-containing oligosaccharides, are expressed by several cells of the immune system and tissue-resident stromal cells. Increasingly, experimental evidence indicates that galectins might play critical regulatory roles in cancer, fibrosis and chronic inflammatory disorders, such as rheumatoid arthritis. In this review, we summarize recent developments in our understanding of the galectins' roles within particular cells, and in the broader context of the inflammatory or tumor microenvironments. This body of knowledge, documenting the coming-of-age of galectins as potential immunosuppressive agents or targets for anti-inflammatory drugs, represents a sound basis to further explore their immunoregulatory properties in the development of novel therapies for autoimmune diseases and chronic inflammation.

Galectins, a subfamily of the animal lectins, are evolutionarily conserved carbohydrate-binding proteins [1]. Members of the galectin family are defined by a conserved carbohydrate-recognition domain (CRD) with a canonical amino acid sequence and affinity for β -galactosides [2]. To date, 15 mammalian galectins have been identified, 11 of which have human orthologues (Table 1). These can be subdivided into three groups; single-CRD galectins; tandem-CRD galectins, and the unique 'chimera-type' galectin-3, which contains a single CRD fused to unusual tandem repeats of short amino acid stretches [1,3].

Some single-CRD galectins can exist as dimers, tandem-CRD galectins have two carbohydrate binding sites and galectin-3 can form oligomers when it binds to multivalent carbohydrates [4,5]. Galectins, like antibodies, can therefore establish ordered arrays of complexes with increased avidity when they bind to multivalent glycosylated proteins [6,7].

Galectins can be found inside and outside cells and have distinct functions in each location [8]. Whether endogenously expressed, or rapidly internalized from the cell surface, galectins have been implicated in important intracellular functions, such as pre-mRNA splicing, regulation of cell growth, cell cycle progression and protein sorting [9–14]. Furthermore, the cytoplasmic-nuclear transport of galectin-3 appears to be regulated by unknown chaperone factors [15] and modulated by neighboring leucine-rich nuclear export signals [16,17].

Although galectins do not contain signal peptides to direct them through the classical endoplasmic reticulum (ER)–Golgi apparatus secretory system, they can be secreted by other unorthodox secretory pathways [18–21]. Once outside the cell, galectins bind to and crosslink multiple molecules found on the cell surface or in the extracellular matrix (ECM) that display appropriate galactose-containing oligosaccharides. In this way, galectins may exert autocrine and paracrine effects to regulate the inflammatory response within tissue microenvironments [3,22,23].

General considerations for extracellular galectin signaling

Cross-linkage of cell-surface receptors by galectins can trigger transmembrane signaling events through which diverse processes such as apoptosis, cytokine secretion and cell migration are modulated. However, highly significant factors that determine the responsiveness of cells to galectin-mediated signals include the repertoire of potentially glycosylated molecules expressed on the cell surface, and the activities of specific glycosyltransferases that are responsible for producing galectin ligands. These variables can dramatically change according to the developmental stage and activation status of cells [24].

In addition to producing galectin ligands, glycosyltransferases can also effectively mask galectin saccharide ligands. For example, the addition of α 2,6-linked sialic acids to lactosamine units by the α 2,6-sialyltransferase I (*ST6Gal-I*) has been shown to block galectin-1 binding [25].

Keywords: apoptosis, cancer, chronic inflammation, fibroblast, fibrosis, galectin, immunosuppression, neutrophil, rheumatoid arthritis, T cell

future
medicine

On the other hand, the Core-2- β -1,6-*N*-acetylglucosaminyltransferase (C2GnT), creates branched polysaccharide structures, which galectin-1 recognizes on T-cell surface glycoproteins, such as CD45 [26].

Undoubtedly, one of the most intriguing findings is the fact that individual galectins can exert contrasting effects on the same target cells, depending on the activation or differentiation state of these cells. In this regard, galectins-1 and -3 have been shown to promote survival or death, activation or silencing and differentiation or proliferation on particular leukocyte subsets [27–31]. The intimate mechanisms of these contrasting effects still remain to be elucidated.

Some galectins, such as galectins-1 and -3, appear to be expressed ubiquitously whereas others, such as galectins-2, -4, -7 and -13, have a more restricted tissue localization [1,3,32]. The expression of the galectins themselves is modulated during the activation and differentiation of immune cells and changes under different physiological, pathological or *in vitro* conditions [22]. For instance, galectins-1 and -3 are upregulated following activation of differentiation in macrophages [19,33,34], T cells [35–37] and fibroblasts [38]. In addition, recent evidence indicates that galectin-1 is overexpressed during the expansion of CD4⁺ CD25⁺-regulatory T cells [39], suggesting a potential role for this protein in the establishment of peripheral tolerance. Furthermore, galectin-9 expression can be upregulated by proinflammatory cytokines, including interleukin (IL)-1 β and interferon

(IFN)- γ [40,41], and galectin-12 expression can be downregulated by different stimuli, including isoproterenol, tumor necrosis factor (TNF)- α and dexamethasone [42]. In addition, different members of the family can be up- or downregulated during myeloid differentiation into the monocyte, eosinophil or neutrophil lineages [43].

Functions of the tandem-CRD galectins

Although galectin-1 (single-CRD galectin) and galectin-3 (chimera-type galectin) are the most extensively studied members of the galectin family, it is gradually becoming evident that other galectins can also modulate innate and acquired immune responses. Examples include the abilities of the tandem-CRD-type galectin-8 to activate microbial killing machinery in neutrophils [44], and of galectin-9 to act as an eosinophil-specific chemoattractant [45]. In addition, galectin-9 can induce the maturation of monocyte-derived human dendritic cells, providing a link between innate and adaptive immunity [46].

Another tandem-CRD-type family member, galectin-4, has been found to play a key role in CD4⁺ T-cell activation in intestinal inflammation [47]. Epithelial cell-derived galectin-4 stimulates an increase of IL-6 production in CD4⁺ T cells and exacerbates chronic colitis. Discussion of this finding is worthwhile in terms of the different roles played by individual members of the same galectin subfamily in activating or silencing pathogenic T-cell responses.

Table 1. Mammalian galectin family and their subgroups.

Single-CRD (monomeric or dimeric)	Two-CRDs in tandem	Chimera-type
Galectin-1		
Galectin-2		
		Galectin-3
	Galectin-4	
Galectin-5 (found in rat)*		
	Galectin-6 (found in chicken and mouse)*	
Galectin-7		
	Galectin-8	
	Galectin-9	
Galectin-10		
Galectin-11 (found in sheep)*		
	Galectin-12	
Galectin-13		
Galectin-14		

*These galectins have no reported human ortholog.
CRD: Carbohydrate recognition domain.

In this regard, immunosuppressive functions for tandem-CRD galectins have also been described. For example, galectin-9 induces apoptosis of murine thymocytes [48] and peripheral CD4⁺ and CD8⁺ T-cell death through a Ca²⁺-calpain-caspase-1 signaling pathway [49]. Interestingly, in a very elegant study, Zhu and colleagues recently showed that galectin-9 is a ligand of Tim-3, a T-helper (Th)1-specific cell-surface molecule. The authors showed that galectin-9 specifically kills Tim-3 expressing IFN- γ -producing cells [50]. Interestingly, this immunosuppressive effect has clear consequences in silencing Th1 responses *in vivo* [50]. This effect will be more extensively discussed in the next sections.

Regarding other members of this family, it has been demonstrated that galectin-8 can induce either growth arrest or apoptosis of tumor cell lines depending upon the activities of cyclin-dependent kinase inhibitors and c-Jun N-terminal kinase (JNK) [51]. Finally, the tightly restricted expression of galectin-12 in adipocytes has also been shown to regulate cell-cycle progression and apoptosis [52,53].

Pro-inflammatory functions of galectin-3
Studies of acute peritonitis in galectin-3-deficient mice have provided significant support for the pro-inflammatory role of galectin-3 [54,55]. Following thioglycolate administration into the peritoneum, fewer granulocytes could be recovered from galectin-3-deficient mice than from wild-type controls. Furthermore, galectin-3 has been shown to promote neutrophil adhesion to laminin and endothelial cells *in vitro* [56,57]. Karlsson and colleagues showed that galectin-3 is able to induce activation of the superoxide-producing NADPH oxidase in primed neutrophils [58]. In this regard, we have recently demonstrated that galectin-3 and soluble fibrinogen together regulate neutrophil activation, degranulation and survival [59].

An essential role for galectin-3 in the phagocytic function of macrophages has been reported by Liu and coworkers [60]. 3 years ago the same group also demonstrated that galectin-3 can promote chemotaxis of human monocytes through interaction with a G-protein coupled receptor [61].

Consistent with its pro-inflammatory function, galectin-3 promotes dendritic cell-naïve T-cell interactions in lymph nodes [62]. This molecule is also a critical intracellular mediator of IL-4-induced survival and differentiation of B cells into

a memory phenotype [63]. Therefore, it seems evident that galectin-3 plays a critical role in the regulation of the inflammatory response.

Interestingly, depending on whether the protein is found in the intracellular compartment or extracellularly, galectin-3 can have dramatically different functions. Fukumori and colleagues demonstrated that extracellular galectin-3 could induce apoptosis in human T cells, binding mainly to CD7 and β 1-integrin and resulting in activation of the mitochondrial death pathway [64]. The proapoptotic effect of galectin-3 was recently confirmed by the groups of Liu and Baum who showed that galectin-3 and galectin-1 can induce cell death through binding to a different set of glycoreceptors [30]. Contrarily, galectin-3 overexpression studies by Yang and colleagues demonstrated that T-cell transfectants exhibited faster growth than control cells and were protected from apoptosis induced through death receptors and mitochondrial routes [11]. Interestingly, over expressed galectin-3 appeared to interact with Bcl-2. Similarly, Hahn and colleagues demonstrated that galectin-1-induced cell death could be inhibited by intracellular expression of galectin-3 [65]. The antiapoptotic activity of intracellular galectin-3 has been implicated in pathological situations, including rheumatoid arthritis (RA) [66], lymphomas [67] and other types of cancer [68].

Galectin-3 in rheumatoid arthritis, bone development & fibrosis

Two studies have independently found increased expression of galectin-3 and galectin-3-binding protein (G3BP) in cells from RA patients. Using subtractive cDNA hybridization, Seki and colleagues found that G3BP was one of 11 genes that were expressed at significantly higher levels in cultured synovial fibroblasts from RA patients than in osteoarthritis patient fibroblasts [69]. Accordingly, Ohshima and colleagues have reported that both galectin-3 and G3BP are abundantly expressed in RA patient synovia, particularly at sites of cartilage invasion [66]. Both proteins could also be found in synovial fluid. Furthermore, galectin-3 expression appears to be associated with the expression of L1 retrotransposable elements [70] and can be induced by the adhesion of synovial fibroblasts to cartilage oligomeric matrix protein [71].

Interestingly, Colnot and colleagues found that galectin-3-deficient mice displayed accelerated apoptosis or terminal differentiation of

chondrocytes in the hypertrophic zones of developing long bones [72]. Furthermore, evidence from Ortega and coworkers suggested that, in matrix metalloproteinase (MMP)-9-deficient mice, excess extracellular galectin-3 can accumulate, potentially leading to increased recruitment of monocytes and the extended survival of osteoclasts [73].

Elegant work by Henderson and colleagues has recently revealed a crucial role of galectin-3 in the fibrotic response to tissue injury [13]. This is consistent with the fact that galectin-3 is an immediate early gene, with a serum responsive element in its promoter [74]. Galectin-3 deficiency in mice drastically reduced renal, pulmonary and hepatic fibrosis by preventing the differentiation of myofibroblasts [13]. Despite comparable levels of transforming growth factor (TGF)- β in injured livers of both wild-type and galectin-3 null animals, and intact Smad-2 and -3 activation in isolated hepatic stellate cells, galectin-3 proved necessary for TGF- β -induced procollagen-I and α -smooth muscle actin expression. Since exogenously added galectin-3 was rapidly internalized, the authors favor a hypothesis in which the essential role of galectin-3 is intracellular, although the protein is likely to be delivered in an autocrine and paracrine fashion.

Immunosuppressive & anti-inflammatory functions of galectins

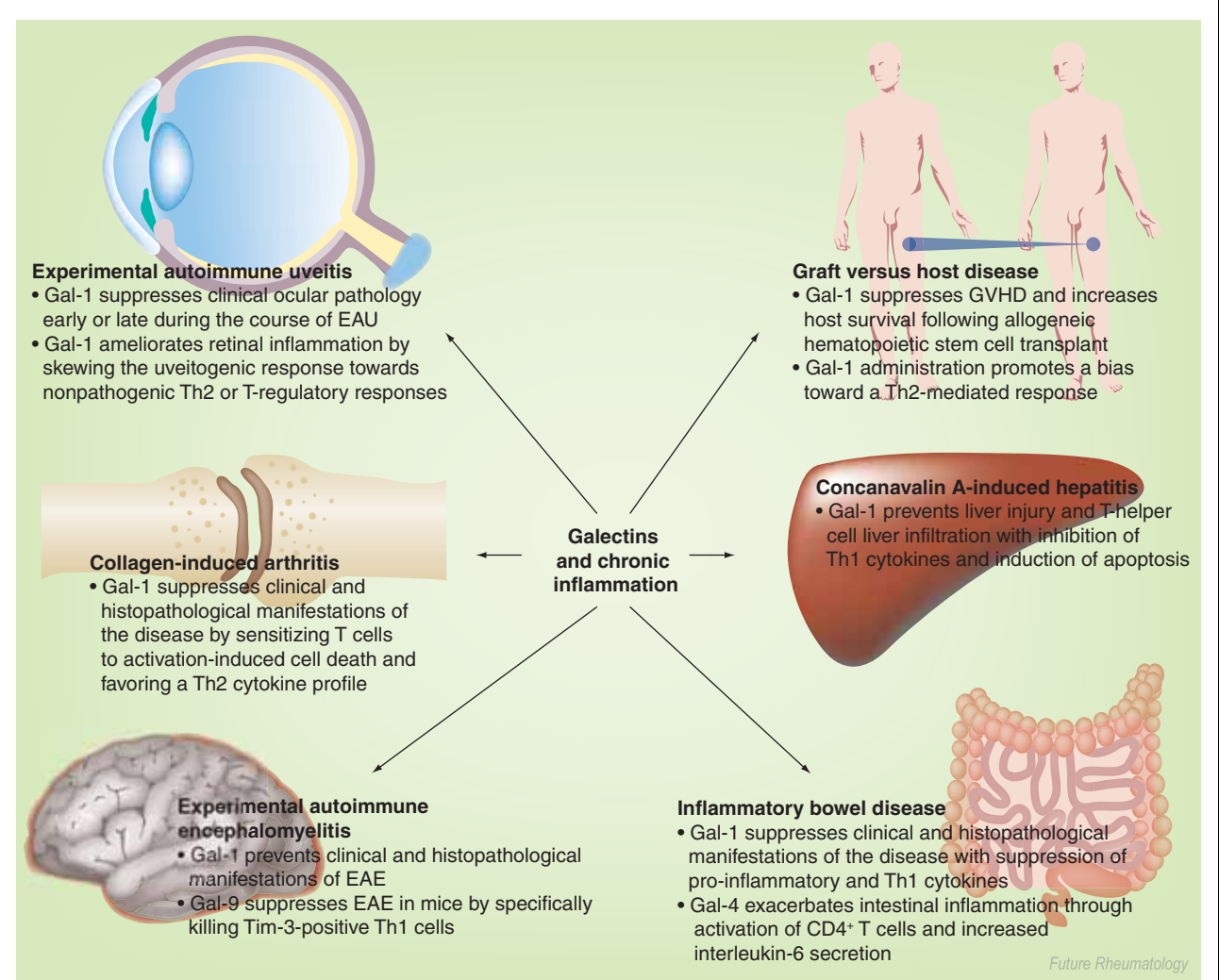
In general, galectin-1, a prototypical single-CRD galectin, displays pro-apoptotic and anti-inflammatory properties (Figure 1). Accordingly, we found that pretreatment of rats with galectin-1 suppressed the acute inflammatory response and inhibited neutrophil extravasation induced by bee venom phospholipase A₂ [75]. Furthermore, arachidonic acid release and nitric oxide production from activated macrophages was inhibited [75,76]. In this regard, La and colleagues demonstrated that, at remarkably low doses (20 pmol/mouse), galectin-1 could inhibit neutrophil chemotaxis and transendothelial migration [77]. The authors speculate that local galectin-1 release from endothelial cells at inflammatory sites may be a crucial negative feedback mechanism to prevent excessive neutrophil recruitment. In addition, it has been reported that exogenous galectin-1 causes phosphatidylserine exposure and phagocytic uptake of activated neutrophils [78]. Therefore, it seems that galectin-1 can display a wide variety of anti-inflammatory effects on different immune cell types.

In this regard, accumulating evidence suggests that galectins might have particularly important roles in the regulation of cell survival in the immune system [79]. Recent work by Endharti and colleagues showed that secretion of galectin-1 by stromal cells supports the survival of naive T cells without promoting proliferation [29]. However, galectin-1 has been reported to induce apoptosis and regulate cell growth in developing thymocytes and activated T cells [34,80,81,82]. In this regard, recent evidence indicates that dendritic cells engineered to over-express galectin-1 can promote contrasting effects on resting and activated T cells, either promoting activation or apoptosis [31]. The fact that galectin-1 is expressed by activated but not resting T cells may point towards an autocrine suicide mechanism, similar to that reported for Fas ligand expression, to prevent excessive T-cell clone expansion after the completion of an ongoing immune response [83].

Perhaps intriguingly, many reports of the pro-apoptotic effect of galectin-1 have used micromolar concentrations of the protein, which are unlikely to exist in biological fluids *in vivo*. However, recent evidence indicates that the more moderate amounts of galectin-1 secreted by most T cells is, in fact, sufficient to kill T cells when the galectin is displayed in the context of ECM glycoproteins [84]. In this regard, the same group recently showed that endothelial cell expression of galectin-1 can also inhibit T-cell transendothelial migration in a manner independent of its pro-apoptotic properties [85].

Several glycosylated proteins on the surface of activated T cells are reported to be crucial receptors for galectins, including CD2, 7, 43 and 45 [86–88]. Galvan and coworkers showed that T cells expressing CD45, but lacking the C2GnT glycosyltransferase, become resistant to galectin-1-induced cell death [26]. Furthermore, a recent report by Lanteri and colleagues is noteworthy in this context – the authors conclude that, during HIV-1 infection, T cells become increasingly susceptible to galectin-1-induced cell death due to altered cell-surface molecule glycosylation [89]. CD7 appears to be a particularly critical receptor for galectin-1-induced cell death. T lymphocytes from patients with mycosis fungoides/Sezary syndrome that lack CD7 expression were demonstrated to be insensitive to galectin-1-triggered death [90]. Interestingly, very recent evidence in neoplastic T-cell lymphoma indicates that haploinsufficiency of C2GnT results in altered

Figure 1. Immunosuppressive effects of galectins in different experimental models of T-cell dependent chronic inflammation and autoimmunity.



The effects of galectins in different murine models of autoimmune disease and inflammation, including collagen-induced arthritis (CIA), EAE, inflammatory bowel disease (IBD), concanavalin A-induced hepatitis, EAU and GVHD. Gal-1 exerts immunosuppressive effects through induction of T-cell apoptosis, modulation of the Th1/Th2 cytokine balance, inhibition of leukocyte migration and generation of regulatory T cells. On the other hand, Gal-9 negatively regulates Th1-mediated responses through selective killing of Tim-3-positive IFN- γ -producing Th1 cells. In addition, Gal-9 can modulate nephrotoxic nephritis through regulation of cell cycle-dependent kinases. EAE: Experimental autoimmune encephalomyelitis; EAU: Experimental autoimmune uveitis; GVHD: Graft versus host disease; Th: T helper.

cellular glycosylation and resistance to galectin-1-induced cell death. These results identify a potentially novel escape mechanism displayed by T-lymphoma cells to survive in galectin-1 enriched microenvironments [91].

Whilst several classical apoptotic signal transduction events have been documented during galectin-1-induced cell death, including caspase activation and cytochrome c release [28], alternative death pathways and apoptotic end points appear to be triggered in different T-cell types [65]. Indeed, apoptosis may only partially explain the immunosuppressive properties of

galectin-1: the T cells that escape apoptosis may instead be subject to suppression of pro-inflammatory cytokine secretion [92,93] or even targeted for phagocytic removal [78].

Miceli and collaborators demonstrated that galectin-1 induces partial T-cell receptor (TCR)- γ chain phosphorylation, antagonizing full signals through the TCR and costimulatory receptors, but allowing partial TCR-mediated responses, such as CD69 upregulation and apoptosis [94,95]. Furthermore, in a very elegant study, Demetriou and colleagues demonstrated that galectin-3 may also restrict

signal transduction initiated by TCR complexes [96]. Hypothetically, the authors argued, the lateral mobility of TCR complexes might be restrained by multivalent complexes of galectin-3 and TCR. Mice deficient in a crucial enzyme in the N-glycosylation pathway (β 1,6 N-acetylglucosaminyltransferase or Mgat5), showed an increased susceptibility to autoimmunity [96]. Thus, galectin-1 and galectin-3 may share an ability to suppress T-cell activation.

In vitro, galectin-1 is also known to block secretion of pro-inflammatory cytokines including IL-2, IFN γ , and TNF- α [92,94]. *In vivo* studies using inflammatory disease models concur; galectin-1 tends to skew the cytokine response to a balance towards the Th2-type [97–102] (Figure 1). In addition, treatment of both nonactivated and activated CD4⁺ and CD8⁺ T cells with recombinant galectin-1 has been reported to cause a significant increase in IL-10 mRNA and protein [103]. IL-10 is known to suppress Th1-type responses, and galectin-1 may therefore employ this mechanism for its immunoregulatory activity. In contrast to galectin-1, galectin-3 suppresses Th2 cytokine secretion in antigen-specific T-cell lines [104]. Furthermore, galectin-3 gene therapy has recently been shown to inhibit both inflammation and stromal remodeling when delivered into the chronically inflamed lungs of mice [105]. Paradoxically, galectin-3-deficient mice also appear to recruit fewer eosinophils than wild-type controls and to display a Th1 cytokine profile in an experimental model of asthma [106]. This apparent discrepancy might be explained by the different experimental strategies used by the authors of these studies (i.e., therapy versus analysis of the susceptibility of galectin-3 gene knock-out mice).

Despite the multiple effects of the exogenously added protein, galectin-1 gene disruption in mice did not apparently cause major spontaneous phenotypic abnormalities [107]. This observation suggests that different members of the galectin family can at least partially compensate for the lack of galectin-1. However, galectin-2, which is structurally related to galectin-1 and is known to share its pro-apoptotic function, clearly operates through a different pathway to galectin-1 [108]. Careful examination of *gal-1* gene deficient mice is beginning to reveal subtle but critical differences in the regulation of inflammatory responses.

Notwithstanding such functional differences, some similarities between the activities of galectins-1 and -2 do exist. For example,

galectin-2 and can shift the balance of T-cell-derived cytokines towards a Th2 profile, an *in vitro* property shared by galectin-1 [108]. Interestingly, galectin-2 appears to regulate lymphotoxin- α secretion, and subtle genetic variants of galectin-2 are reported to differentially influence the extent of inflammation during myocardial infarction [109]. In this regard, a cross-sectional genetic study performed in a British population indicated a striking correlation between plasma glucose, serum insulin and the galectin-2 genotype [110].

Although no evidence exists for the ability of other exogenously-added single-CRD galectins to affect cell survival, transfection of galectin-7 (p53-induced gene 1) in epithelial tumor cell lines did reveal its potential intracellular proapoptotic activity [111].

Regarding the immunosuppressive activities of tandem-CRD galectins, recent evidence indicates that galectin-9 can suppress the progression of experimental autoimmune encephalomyelitis by selectively killing Tim-3-positive IFN- γ -producing cells [50]. Interestingly, T-cell-mediated neuroinflammation was exacerbated in mice treated with galectin-9 small interfering (si)RNA suggesting that knocking-down galectin-9 expression during disease induction may affect the progression of the disease (Figure 1). In addition, Tsuchiyama and colleagues reported that the effect of galectin-9 inhibits the infiltration of CD8⁺ T cells in an experimental rat model of nephritis [112]. Furthermore, recent evidence indicates that galectin-9 may inhibit glomerular hypertrophy in db/db diabetic mice via inhibition of cyclin-dependent kinase inhibitors [113]. Thus, future studies are warranted to investigate the different mechanisms involved in the immunosuppressive activities of individual members of the galectin family.

Galectins & tumor-immune escape

Galectins have been shown to modulate different events of tumor progression [3]. Interestingly, expression of galectin-1 (as well as other galectins) in cancer cells positively correlates with the aggressiveness of tumors [114,115]. This suggested that secretion of galectin-1 by tumor cells may be a mechanism by which immunosuppressive microenvironments at tumor sites can be created. This hypothesis was supported using a combined *in vitro* and *in vivo* strategy: galectin-1-mediated immunoregulation clearly had an important role in tumor immune escape [116]. Local galectin-1 blockade allowed CD4⁺ and CD8⁺ tumor-specific T-cell responses to be mounted, causing a reduction in tumor mass.

Given its potent immunosuppressive effects, galectin-1 may be a useful target for therapeutic intervention in cancer.

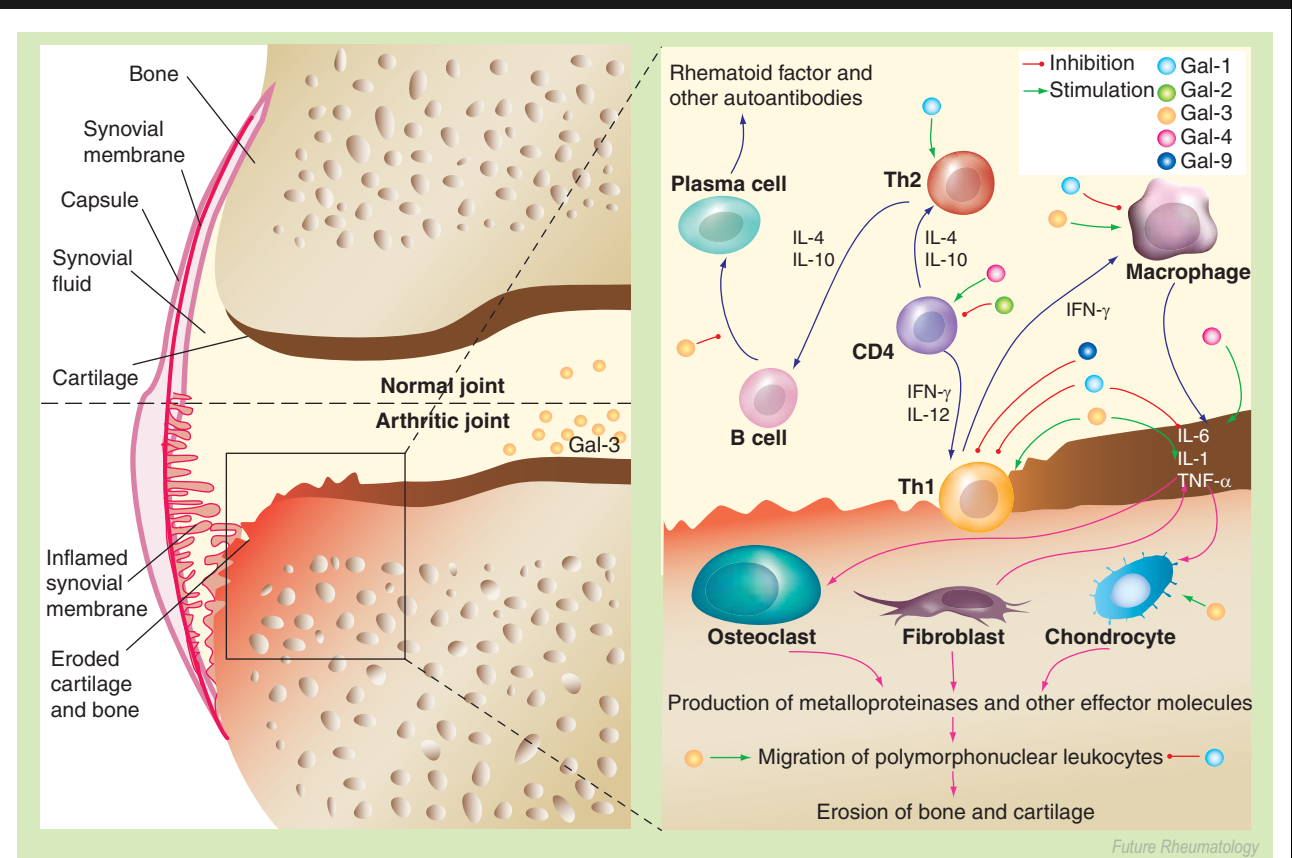
Since galectins-2, -3 and -9 have also been shown to affect T-cell survival, future studies are warranted to investigate the potential role of these proteins in tumor-cell evasion of immune responses *in vivo*.

Therapeutic potential of galectins as novel immunosuppressive agents

In a study using the DAB/1 collagen-induced arthritis model, we found a strong correlation between the apoptotic properties of galectin-1 *in vitro* and its therapeutic potential *in vivo* [97]. We demonstrated that, at the day of disease onset, a single injection of syngeneic DAB/1 fibroblasts engineered to secrete galectin-1

could abrogate clinical and histopathological manifestations of arthritis. This process involved an increased susceptibility of lymph-node cells to antigen-induced apoptosis and a shift from a Th1 to a Th2-polarised immune response. Interestingly, galectin-1-expressing fibroblasts also inhibited antigen-dependent IL-2 production in a collagen type-II-specific T-cell clone (Figure 1). In addition, in synovial tissue from juvenile RA patients, we found an interesting correlation between the levels of galectins-1 and -3 and the regulation of apoptosis [117]. Given the contrasting immunoregulatory functions of different members of the galectin family, we speculate that individual members of the galectin family might play different roles in the context of an inflamed joint during the development and resolution of RA (Figure 2).

Figure 2. Potential role of galectins in the immunopathology of rheumatoid arthritis.



The potential role of individual members of the galectin family in the context of inflamed synovial tissue. Galectins are expressed by a number of different inflammatory, stromal cells and synovial fibroblasts, and may regulate the function of these cells, thereby affecting the development of inflammatory responses. As illustrated in this diagram, galectins can behave as pro- or anti-inflammatory mediators by modulating the physiology and responses of immune cells, including macrophages, synovial fibroblasts, Th1 and Th2 cells, B cells, neutrophils and mast cells. By positively or negatively affecting the inflammatory response, galectins may indirectly influence the clinical course of rheumatoid arthritis.

GAL: Galectin; IFN: Interferon; IL: Interleukin; Th: T helper; TNF: Tumor necrosis factor.

Similarly, in T-cell-dependent animal models of liver injury [98] and inflammatory bowel disease [99], galectin-1 pretreatment has been shown to abrogate tissue damage and T-cell infiltration (Figure 1). Again the immunosuppressive mechanism involved the selective culling of antigen-activated T cells and inhibition of pro-inflammatory cytokine secretion from T cells and macrophages. Remarkably, even in a murine model of graft-versus-host disease, treatment with galectin-1 substantially suppressed inflammation without compromising the engraftment of donor cells [100]. Finally, given the potential role of galectin-1 in the maintenance of immune privilege in organs such as the eye, we have recently investigated the immunoregulatory effects of this protein in experimental autoimmune uveitis (EAU), a Th1-mediated model of retinal disease [101]. Interestingly, treatment with galectin-1 either early or late in the course of EAU was sufficient to suppress clinical ocular pathology, inhibit leukocyte infiltration and counteract pathogenic Th1 cells [101]. Administration of galectin-1 ameliorated retinal inflammation by skewing the uveitogenic response towards non-pathogenic Th2 or T regulatory (IL-10 and TGF- β)-mediated anti-inflammatory responses (Figure 1). These results highlight the ability of this endogenous lectin to counteract Th1-mediated responses through different, but potentially overlapping, anti-inflammatory mechanisms. In addition, a striking correlation was found between the levels of antiretinal galectin-1 autoantibodies in sera from uveitic patients and the severity of autoimmune retinal inflammation [118].

In addition, the ability of galectin-9 to negatively regulate Th1 responses [50], to suppress neuroinflammation [50] and to inhibit glomerular hypertrophy and nephritis [112,113] suggest that this tandem-repeat galectin may also be used as a potent target in autoimmunity and chronic inflammation.

Conclusions & future perspective

As we have seen, galectins can modulate both innate and adaptive immune responses by acting intracellularly and extracellularly, as chemokines, adhesion molecules, differentiation factors, death triggers and survival inducers. However, before the use of galectin-based therapeutic

agents can be fully realized, a more thorough understanding of the lesser studied galectins is required. To what extent is there functional redundancy and specificity of action within the galectin family? What doses are required to achieve immunosuppression and what are the tolerated 'nontoxic' doses of these sugar-binding proteins *in vivo*? What are the optimal vehicles for galectin-1 delivery and the most efficient administration routes?

The tolerogenic potency of galectin-1 is such that tumors dysregulate its expression to attain immune privilege [116]. Indeed, specific galectin-1 inhibitors might prove to be potent anticancer agents [119]. Future studies are warranted to investigate the immunosuppressive effects of other members of the galectin family, including galectins-2, -3 and -9, within tumor microenvironment.

Contrarily, synthetic glycoconjugates or lectins may prove to be excellent therapeutic immunosuppressive drugs for treating chronic inflammatory and autoimmune conditions [120–122]. Furthermore, galectin-3 gene silencing may be a very effective treatment for preventing fibrosis [13]. Our current knowledge promises a future scenario in which individual members of the galectin family may be used as immunoregulatory agents (e.g., galectins-1 and -9) or targets for anti-inflammatory drugs in autoimmune disorders, including RA. Future studies should be focussed on the careful examination of galectin-1 or -9-based immunosuppressive agents [123] or specific galectin-3 inhibitors [124] for the treatment of chronic inflammation *in vivo*.

Acknowledgements

We give special thanks to all the colleagues who shared their important contributions in the field with us. We apologize that we could not cite many excellent studies because of space limitations. Work in the authors' laboratory is supported by grants from Mizutani Foundation for Glycoscience, Fundación Florencio Fiorini, Agencia de Promoción Científica y Tecnológica (PICT 2003-05-13787), Fundación Sales, University of Buenos Aires, Argentina (UBACYT-M091) (to Gabriel A Rabinovich) and from the Medical Research Council (G116/131) and Arthritis Research Campaign (16390) (to Christopher D Buckley). Gabriel A Rabinovich is a member of the scientific career of CONICET, Marta A Toscano, German A Bianco and Juan M Illaregui thank CONICET for the fellowships granted.

Executive summary**Galectin family**

- Structurally distinct subgroups; galectins with a single-carbohydrate recognition domain (CRD), galectins with two CRDs in tandem and chimera-type galectins that contain a single CRD fused to unusual tandem repeats of short amino acid stretches.
- Galectins are secreted by nonclassical mechanisms.
- Galectins can act in an autocrine and paracrine manner to positively or negatively regulate the inflammatory response within tissue microenvironments.

General considerations for extracellular galectin signaling

- The repertoire of glycosylated cell-surface molecules available for galectin binding is determined by the specific activities of glycosyltransferase enzymes within the cell. Some create and expose galectin ligands, whilst others mask them.

Functions of the tandem-CRD galectins

- Members of the tandem-CRD galectin subgroup have not been investigated as widely as galectin-1 (the prototypical single-CRD subgroup member) and galectin-3 (the only chimera-type subgroup member). However recent findings suggest intriguing roles for two-CRD 'tandem-repeat' galectins in the regulation of inflammation.
- Pro-inflammatory functions have been described for galectins -8 and -9 in the innate arm of the immune response.
- In the context of adaptive immunity, galectin-4 has been shown to affect T-cell activation and interleukin (IL)-6 production and galectin-9 modulates dendritic cell maturation.
- A pro-apoptotic function has also been reported for galectin-9.
- Recent evidence indicates that galectin-9 is a ligand of Tim-3, a T helper (Th)1-specific cell-surface molecule. Galectin-9 negatively regulates Th1 responses, through binding to Tim-3,

Regulation of the inflammatory response by galectin-3 (the only chimera-type galectin)

- Galectin-3-deficient mice mount a poor inflammatory response to intraperitoneal thioglycollate injection.
- Galectin-3 activates superoxide burst in neutrophils.
- This chimera-type lectin acts as a chemoattractant for monocytes and macrophages.
- It has been demonstrated that this protein is critical for phagocytic function of macrophages.
- Interestingly, galectin-3 acts intracellularly to prevent apoptosis, while exogenously added galectin-3 promotes T-cell apoptosis.

Galectin-3 in rheumatoid arthritis, bone development & fibrosis

- Galectin-3 is highly expressed together with galectin-3-binding protein (Mac-2BP/90K) at the sites of joint erosion in rheumatoid arthritis (RA) patients.
- Potentially, this protein has an antiapoptotic role in chondrocytes and osteoclasts during embryonic osteogenesis.
- Galectin-3 is a necessary factor for transforming growth factor- β -induced myofibroblast differentiation in the fibrotic response to tissue damage.

Immunosuppressive & proapoptotic functions of one-CRD galectins

- Galectin-1 can suppress acute inflammation *in vivo* by preventing neutrophil extravasation.
- Galectin-1 induces thymocyte and activated peripheral T-cell apoptosis through binding to a variety of glycoreceptors, including CD2, 7, 43 and 45.
- Galectin-1 suppresses proximal signals through the T-cell receptor and downregulates the release of pro-inflammatory cytokines from T cells.
- Galectin-2 promotes apoptosis of activated T cells and induces a bias toward a Th2 response *in vitro*.

Galectin-1 & tumor immune escape

- Tumors dysregulate the expression of galectin-1 to attain an immune-privileged microenvironment.
- The ability of other members of the galectin family (e.g., galectins-2, -3 and -9) to suppress T-cell responses and their high levels in certain tumor types suggest the potential contribution of these proteins to tumor-cell evasion of immune responses.

Therapeutic potential of galectins as novel immunosuppressants

- Galectin-1 suppresses pathology in several T-cell-dependent animal models of disease, including collagen-induced arthritis, experimental autoimmune encephalomyelitis, experimental autoimmune uveitis, concanavalin a-induced hepatitis, inflammatory bowel disease and graft-versus-host disease.
- In most cases, galectin-1 achieved immunosuppression by specific culling of activated T cells, together with immune deviation to a Th2-type response.
- Interestingly, galectin-9 suppresses experimental autoimmune encephalomyelitis by specifically killing Tim-3-positive Th1 pathogenic cells.

Bibliography

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

1. Leffler H, Carlsson S, Hedlund M, Qian Y, Poirier F: Introduction to galectins. *Glycoconj. J.* 19(7–9), 433–440 (2004).
- **Succinct overview of many aspects of the galectin field.**
2. Cooper DN: Galectinomics: finding themes in complexity. *Biochim. Biophys. Acta* 1572(2–3), 209–231 (2002).
- **Comprehensive review of the emerging themes of the complex field of galectins. In particular, the biochemical and structural features of galectins in diverse animal species.**
3. Liu FT, Rabinovich GA: Galectins as modulators of tumor progression. *Nat. Rev. Cancer* 5(1), 29–41 (2005).
4. Giudicelli V, Lutowski D, Levi-Strauss M *et al.*: Is human galectin-1 activity modulated by monomer/dimer equilibrium? *Glycobiology* 7(3), 8–10 (1997).
5. Ahmad N, Gabius HJ, Andre S *et al.*: Galectin-3 precipitates as a pentamer with synthetic multivalent carbohydrates and forms heterogeneous cross-linked complexes. *J. Biol. Chem.* 279(12), 10841–10847 (2004).
6. Brewer CF: Binding and cross-linking properties of galectins. *Biochim. Biophys. Acta* 1572(2–3), 255–262 (2002).
7. Sacchetti JC, Baum LG, Brewer CF: Multivalent protein-carbohydrate interactions. A new paradigm for supermolecular assembly and signal transduction. *Biochemistry* 40(10), 3009–3015 (2001).
- **Describes a new paradigm by which binding and cross-linking of multivalent carbohydrates with multivalent lectins can affect signal transduction in biological systems.**
8. Liu FT, Patterson RJ, Wang JL: Intracellular functions of galectins. *Biochem. Biophys. Acta* 1572(2–3), 263–273 (2002).
- **Focusses on lectins inside the cell and their participation in fundamental intracellular processes, such as pre-mRNA splicing and protection from apoptosis.**
9. Vyakarnam A, Dagher SF, Wang JL, Patterson RJ: Evidence for a role for galectin-1 in pre-mRNA splicing. *Mol. Cell Biol.* 17(8), 4730–4737 (1997).
10. Dagher SF, Wang JL, Patterson RJ: Identification of galectin-3 as a factor in pre-mRNA splicing. *Proc. Natl Acad. Sci. USA* 92(4), 1213–1217 (1995).
11. Yang RY, Hsu DK, Liu FT: Expression of galectin-3 modulates T-cell growth and apoptosis. *Proc. Natl Acad. Sci. USA* 93(13), 6737–6742 (1996).
- **First paper demonstrating the anti-apoptotic function of galectin-3.**
12. Yang RY, Hsu DK, Yu L, Ni J, Liu FT: Cell cycle regulation by galectin-12, a new member of the galectin superfamily. *J. Biol. Chem.* 276(23), 20252–20260 (2001).
13. Henderson NC, Mackinnon AC, Farnworth SL *et al.*: Galectin-3 regulates myofibroblast activation and hepatic fibrosis. *Proc. Natl Acad. Sci. USA* 103(13), 5060–5065 (2006).
- **Elegant study demonstrating a critical role of galectin-3 in the inhibition of myofibroblast activation and hepatic fibrosis, with therapeutic implications in the prevention and treatment of liver fibrosis.**
14. Delacour D, Cramm-Behrens CI, Drobecq H, *et al.*: Requirement for galectin-3 in apical protein sorting. *Curr. Biol.* 16(4), 408–414 (2006).
15. Openo KP, Kadrofske MM, Patterson RJ, Wang JL: Galectin-3 expression and subcellular localization in senescent human fibroblasts. *Exp. Cell Res.* 255(2), 278–290 (2000).
16. Davidson PJ, Li SY, Lohse AG *et al.*: Transport of galectin-3 between the nucleus and the cytoplasm. I. Conditions and signals for nuclear import. *Glycobiology* 16(7), 602–611 (2006).
17. Li SY, Davidson PJ, Lin NY *et al.*: Transport of galectin-3 between the nucleus and the cytoplasm. II. Identification of the signal for nuclear transport. *Glycobiology* 16(7), 612–622 (2006).
- **References [15–17] provide the molecular basis of the intracellular shuttling of galectin-3 between the nucleus and the cytoplasm.**
18. Cooper DN, Barondes SH: Evidence for export of a muscle lectin from cytosol to extracellular matrix and for a novel secretory mechanism. *J. Cell. Biol.* 110(5), 1681–1691 (1990).
- **Pioneer study describing the unorthodox mechanism of secretion of galectin-1.**
19. Sato S, Hughes RC: Control of Mac-2 surface expression on murine macrophage cell lines. *Eur. J. Immunol.* 24(1), 216–221 (1994).
- **Pioneer study on the regulation of galectins by inflammatory stimuli.**
20. Zhu WQ, Ochieng J: Rapid release of intracellular galectin-3 from breast carcinoma cells by fetuin. *Cancer Res.* 61(5), 1869–1873 (2001).
21. Seelenmeyer C, Weghlingel S, Tews I *et al.*: Cell surface counter receptors are essential components of the unconventional export machinery of galectin-1. *J. Cell Biol.* 171(2), 373–381 (2005).
22. Rubinstein N, Ilarregui JM, Toscano MA, Rabinovich GA: The role of galectins in the initiation, amplification and resolution of the inflammatory response. *Tissue Antigens* 64(1), 1–12 (2004).
23. Gabius HJ: Cell surface glycans: the why and how of their functionality as biochemical signals in lectin-mediated information transfer. *Crit. Rev. Immunol.* 26(1), 43–79 (2006).
- **Critical review on the role of protein-glycan interactions in biological systems.**
24. Bianco GA, Toscano MA, Ilarregui JM, Rabinovich GA: Impact of protein-glycan interactions in the regulation of autoimmunity and chronic inflammation. *Autoimmun. Rev.* 5(5), 349–356 (2006).
25. Amano M, Galvan M, He J, Baum LG: The ST6Gal I sialyltransferase selectively modifies N-glycans on CD45 to negatively regulate galectin-1-induced CD45 clustering, phosphatase modulation, and T cell death. *J. Biol. Chem.* 278(9), 7469–7475 (2003).
26. Galvan M, Tsuboi S, Fukuda M, Baum LG: Expression of a specific glycosyltransferase enzyme regulates T cell death mediated by galectin-1. *J. Biol. Chem.* 275(22), 16730–16737 (2000).
- **[25] and [26] describe the role of specific glycosyltransferases in modulating susceptibility to galectin-1-induced cell death.**
27. Almkvist J, Dahlgren C, Leffler H, Karlsson A: Activation of the neutrophil nicotinamide adenine dinucleotide phosphate oxidase by galectin-1. *J. Immunol.* 168(8), 4034–4041 (2002).
28. Matarrese P, Tinari A, Mormone E *et al.*: Galectin-1 sensitizes resting human T lymphocytes to Fas (CD95)-mediated cell death via mitochondrial hyperpolarization, budding, and fission. *J. Biol. Chem.* 280(8), 6969–6985 (2005).
29. Endharti AT, Zhou YW, Nakashima I, Suzuki H: Galectin-1 supports survival of naive T cells without promoting cell proliferation. *Eur. J. Immunol.* 35(1), 86–97 (2005).
30. Stillman BN, Hsu DK, Pang M *et al.*: Galectin-3 and galectin-1 bind distinct cell surface glycoprotein receptors to induce T cell death. *J. Immunol.* 176(2), 778–789 (2006).

31. Perone MJ, Larregina AT, Schufesky WJ *et al.*: Transgenic galectin-1 induces maturation of dendritic cells that elicit contrasting responses in naive and activated T cells. *J. Immunol.* 176(12), 7207–7220 (2006).
32. Lahm H, Andre S, Hoefflich A *et al.*: Comprehensive galectin fingerprinting in a panel of 61 human tumor cell lines by RT-PCR and its implications for diagnostic and therapeutic procedures. *J. Cancer Res. Clin. Oncol.* 127(6), 375–386 (2001).
33. Rabinovich GA, Castagna L, Landa C *et al.*: Regulated expression of a 16-kd galectin-like protein in activated rat macrophages. *J. Leukoc. Biol.* 59(3), 363–370 (1996).
34. Rabinovich GA, Iglesias MM, Modesti NM *et al.*: Activated rat macrophages produce a galectin-1-like protein that induces apoptosis of T cells: biochemical and functional characterization. *J. Immunol.* 169(10), 4831–4840 (1998).
35. Blaser C, Kaufmann M, Muller C *et al.*: β -galactoside-binding protein secreted by activated T cells inhibits antigen-induced proliferation of T cells. *Eur. J. Immunol.* 28(8), 2311–2319 (1998).
- **First description of the hypothesis that galectin-1 may act as an autocrine negative regulatory signal in the immune system.**
36. Fuertes MB, Molinero LL, Toscano MA *et al.*: Regulated expression of galectin-1 during T cell activation involves Lck and Fyn kinases and signaling through MEK1/ERK, p38 MAP kinase and p70S6 kinase. *Mol. Cell. Biochem.* 267(1–2), 177–185 (2004).
37. Joo HG, Goedegebuure PS, Sadanaga N *et al.*: Expression and function of galectin-3, a β -galactoside-binding protein in activated T lymphocytes. *J. Leukoc. Biol.* 69(4), 555–564 (2001).
38. Voss PG, Tsay YG, Wang JL: Galectin-3: differential accumulation of distinct mRNAs in serum-stimulated mouse 3T3 fibroblasts. *Glycoconj. J.* 11(4), 353–362 (1994).
39. Sugimoto N, Oida T, Hirota K *et al.*: Foxp3-dependent and independent molecules specific for CD25⁺ CD4⁺ natural regulatory T cells revealed by DNA microarray analysis. *Int. Immunol.* (2006) (In Press).
40. Yoshida H, Imaizumi T, Kumagai M *et al.*: Interleukin-1 beta stimulates galectin-9 expression in human astrocytes. *Neuroreport* 12(17), 3755–3758 (2001).
41. Imaizumi T, Kumagai M, Sazaki N *et al.*: Interferon-gamma stimulates the expression of galectin-9 in cultured human endothelial cells. *J. Leukoc. Biol.* 72(3), 486–491 (2002).
42. Fasshauer M, Klein J, Lossner U *et al.*: Negative regulation of adipose-expressed galectin-12 by isoproterenol, tumor necrosis factor- α insulin and dexamethasone. *Eur. J. Endocrinol.* 147(4), 553–559 (2002).
43. Abedin MJ, Kashio Y, Seki M *et al.*: Potential roles of galectins in myeloid differentiation into three different lineages. *J. Leukoc. Biol.* 73(5), 650–656 (2003).
44. Nishi N, Shoji H, Seki M *et al.*: Galectin-8 modulates neutrophil function via interaction with integrin α M. *Glycobiology* 13(11), 755–763 (2003).
45. Matsumoto R, Hirashima M, Kita H, Gleich GJ: Biological activities of ecalectin: a novel eosinophil-activating factor. *J. Immunol.* 168(4), 1961–1967 (2002).
- **Describes the role of galectin-9 in leukocyte chemotaxis with critical implications in the development of inflammation.**
46. Dai SY, Nakagawa R, Itoh A *et al.*: Galectin-9 induces maturation of human monocyte-derived dendritic cells. *J. Immunol.* 175(5), 2974–2981 (2005).
47. Hokama A, Mizoguchi E, Sugimoto K *et al.*: Induced reactivity of intestinal CD4⁺ T cells with an epithelial cell lectin, galectin-4, contributes to exacerbation of intestinal inflammation. *Immunity* 20(6), 681–693 (2004).
- **Demonstrates a critical role of galectin-4 in T-cell activation and inflammation.**
48. Wada J, Ota K, Kumar A *et al.*: Developmental regulation, expression, and apoptotic potential of galectin-9, a β -galactoside binding lectin. *J. Clin. Invest.* 99(10), 2452–2461 (1997).
49. Kashio Y, Nakamura K, Abedin MJ *et al.*: Galectin-9 induces apoptosis through the calcium-calpain-caspase-1 pathway. *J. Immunol.* 170(7), 3631–3636 (2003).
- **References [48] and [49] demonstrate the ability of galectin-9 to kill immature thymocytes and peripheral T cells.**
50. Zhu C, Anderson AC, Schubart A *et al.*: The Tim-3 ligand galectin-9 negatively regulates T helper type 1 immunity. *Nat. Immunol.* 6(12), 1245–1252 (2005)
- **Elegant study demonstrating *in vitro* and *in vivo* that galectin-9 acts as a specific ligand for Tim-3 on the surface of T-helper (Th)1 cells and negatively regulates Th1-mediated immune responses.**
51. Arbel-Goren R, Levy Y, Ronen D, Zick Y: Cyclin-dependent kinase inhibitors and JNK act as molecular switches, regulating the choice between growth arrest and apoptosis induced by galectin-8. *J. Biol. Chem.* 280(19), 19105–19114 (2005).
52. Yang RY, Hsu DK, Yu L *et al.*: Cell cycle regulation by galectin-12, a new member of the galectin superfamily. *J. Biol. Chem.* 276(23), 20252–20260. (2001)
- **References [51] and [52] clearly demonstrate the ability of galectin-8 and galectin-12 (tandem-repeat galectins) in the regulation of cell cycle progression.**
53. Hotta K, Funahashi T, Matsukawa Y *et al.*: Galectin-12, an adipose-expressed galectin-like molecules possessing apoptosis-inducing activity. *J. Biol. Chem.* 276(36), 34089–34097 (2001).
54. Colnot C, Ripoché MA, Milon G *et al.*: Maintenance of granulocyte numbers during acute peritonitis is defective in galectin-3-null mutant mice. *Immunology* 94(3), 290–296 (1998).
55. Hsu DK, Yang RY, Pan Z *et al.*: Targeted disruption of the galectin-3 gene results in attenuated peritoneal inflammatory responses. *Am. J. Pathol.* 156(3), 1073–1083 (2000).
- **Studies in [54] and [55] were the first to show a role for galectin-3 in the regulation of inflammation *in vivo*.**
56. Kuwabara I, Liu FT: Galectin-3 promotes adhesion of human neutrophils to laminin. *J. Immunol.* 156(10), 3939–3944 (1996).
57. Sato S, Ouellet N, Pelletier I *et al.*: Role of galectin-3 as an adhesion molecule for neutrophil extravasation during streptococcal pneumonia. *J. Immunol.* 168(4), 1813–1822 (2002).
58. Karlsson A, Follin P, Leffler H, Dahlgren C: Galectin-3 activates the NADPH-oxidase in exudated but not peripheral blood neutrophils. *Blood* 91(9), 3430–3438 (1998).
59. Fernandez GC, Ilarregui JM, Rubel CJ *et al.*: Galectin-3 and soluble fibrinogen act in concert to modulate neutrophil activation and survival: involvement of alternative MAPK pathways. *Glycobiology* 15(5), 519–527 (2005).
60. Sano H, Hsu DK, Apgar JR *et al.*: Critical role of galectin-3 in phagocytosis by macrophages. *J. Clin. Invest.* 112(3), 389–397 (2003).
61. Sano H, Hsu DK, Yu L *et al.*: Human galectin-3 is a novel chemoattractant for monocytes and macrophages. *J. Immunol.* 165(4), 2156–2164 (2000).
62. Swarte VV, Mebius RE, Joziase DH *et al.*: Lymphocyte triggering via L-selectin leads to enhanced galectin-3-mediated binding to dendritic cells. *Eur. J. Immunol.* 28(9), 2864–2871 (1998).

63. Acosta-Rodriguez EV, Montes CL, Motran CC *et al.*: Galectin-3 mediates IL-4-induced survival and differentiation of B cells: functional cross-talk and implications during *Trypanosoma cruzi* infection. *J. Immunol.* 172(1), 493–502 (2004).
64. Fukumori T, Takenaka Y, Yoshii T *et al.*: CD29 and CD7 mediate galectin-3-induced type II T-cell apoptosis. *Cancer Res.* 63(23), 8302–8311 (2003).
- **First study describing the role of exogenous galectin-3 in the induction of T-cell apoptosis.**
65. Hahn HP, Pang M, He J *et al.*: Galectin-1 induces nuclear translocation of endonuclease G in caspase- and cytochrome c-independent T cell death. *Cell Death Differ.* 11(12), 1277–1286 (2004).
66. Ohshima S, Kuchen S, Seemayer CA *et al.*: Galectin 3 and its binding protein in rheumatoid arthritis. *Arthritis Rheum.* 48(10), 2788–2795 (2003).
- **First study demonstrating that galectin-3 and galectin-3-binding protein may represent novel markers of disease activity in rheumatoid arthritis patients.**
67. Hoyer KK, Pang M, Gui D *et al.*: An anti-apoptotic role for galectin-3 in diffuse large B-cell lymphomas. *Am. J. Pathol.* 164(3), 893–902 (2004).
68. Nakahara S, Oka N, Raz A: On the role of galectin-3 in cancer apoptosis. *Apoptosis* 10(2), 267–275 (2005).
69. Seki T, Selby J, Haupt T, Winchester R: Use of differential subtraction method to identify genes that characterize the phenotype of cultured rheumatoid arthritis synoviocytes. *Arthritis Rheum.* 41(8), 1356–1364 (1998).
70. Neidhart M, Rethage J, Kuchen S *et al.*: Retrotransposable L1 elements expressed in rheumatoid arthritis synovial tissue: association with genomic DNA hypomethylation and influence on gene expression. *Arthritis Rheum.* 43(12), 2634–2647 (2000).
71. Neidhart M, Zaucke F, von Knoch R *et al.*: Galectin-3 is induced in rheumatoid arthritis synovial fibroblasts after adhesion to cartilage oligomeric matrix protein. *Ann. Rheum. Dis.* 64(3), 419–424 (2005).
72. Colnot C, Sidhu SS, Balmain N, Poirier F: Uncoupling of chondrocyte death and vascular invasion in mouse galectin 3 null mutant bones. *Dev. Biol.* 229(1), 203–214 (2001).
73. Ortega N, Behonick DJ, Colnot C, Cooper DN, Werb Z: Galectin-3 is a downstream regulator of matrix metalloproteinase-9 function during endochondral bone formation. *Mol. Biol. Cell* 16(6), 3028–3039 (2005).
74. Kadrofske MM, Openo KP, Wang JL: The human LGALS3 (galectin-3) gene: determination of the gene structure and functional characterization of the promoter. *Arch. Biochem. Biophys.* 349(1), 7–20 (1998).
75. Rabinovich GA, Sotomayor CE, Riera CM, Bianco I, Correa SG: Evidence of a role for galectin-1 in acute inflammation. *Eur. J. Immunol.* 30 (5), 1331–1339 (2000).
76. Correa SG, Sotomayor CE, Aoki MP, Maldonado CA, Rabinovich GA: Opposite effects of galectin-1 on alternative metabolic pathways of L-arginine in resident, inflammatory and activated macrophages. *Glycobiology* 13(2), 119–128 (2003).
77. La M, Cao TV, Cerchiaro G *et al.*: A novel biological activity for galectin-1: inhibition of leukocyte-endothelial cell interactions in experimental inflammation. *Am. J. Pathol.* 163(4), 1505–1515 (2003).
78. Dias-Baruffi M, Zhu H, Cho M *et al.*: Dimeric galectin-1 induces surface exposure of phosphatidylserine and phagocytic recognition of leukocytes without inducing apoptosis. *J. Biol. Chem.* 278(42), 41282–41293 (2003).
- **Describes the ability of galectin-1 to promote phosphatidylserine exposure thus favoring phagocytic recognition and homeostasis.**
79. Rabinovich GA, Baum LG, Tinari N *et al.*: Galectins and their ligands: amplifiers, silencers or tuners of the inflammatory response? *Trends Immunol.* 23(6), 313–320 (2002).
- **Comprehensive review on the role of galectins in the inflammatory response.**
80. Perillo NL, Uittenbogaart CH, Nguyen JT, Baum LG: Galectin-1, an endogenous lectin produced by thymic epithelial cells, induces apoptosis of human thymocytes. *J. Exp. Med.* 185(10), 1851–1858 (1997).
81. Perillo NL, Pace KE, Seilhamer JJ, Baum LG: Apoptosis of T cells mediated by galectin-1. *Nature* 378(6558), 736–739 (1995).
- **References [80] and [81] describe for the first time the ability of galectin-1 to induce apoptosis of immature thymocytes and peripheral T cells.**
82. Rabinovich GA, Modesti NM, Castagna LF, Landa CA, Riera CM, Sotomayor CE: Specific inhibition of lymphocyte proliferation and induction of apoptosis by CLL-1, a beta-galactoside-binding lectin. *J. Biochem.* 122(2), 365–373 (1997).
83. Lenardo MJ: The molecular regulation of lymphocyte apoptosis. *Semin. Immunol.* 9(1), 1–5 (1997).
84. He J, Baum LG: Presentation of galectin-1 by extracellular matrix triggers T cell death. *J. Biol. Chem.* 279(6), 4705–4712 (2004).
85. He J, Baum LG: Endothelial cell expression of galectin-1 induced by prostate cancer cells inhibits T-cell transendothelial cell migration. *Lab. Invest.* (2006) (In Press).
86. Walzel H, Blach M, Hirabayashi J, Kasai KI, Brock J: Involvement of CD2 and CD3 in galectin-1 induced signaling in human Jurkat T-cells. *Glycobiology* 10(2), 131–140 (2000).
87. Pace KE, Hahn HP, Pang M, Nguyen JT, Baum LG: CD7 delivers a pro-apoptotic signal during galectin-1-induced T cell death. *J. Immunol.* 165(5), 2331–2334 (2000).
88. Pace KE, Lee C, Stewart PL, Baum LG: Restricted receptor segregation into membrane microdomains occurs on human T cells during apoptosis induced by galectin-1. *J. Immunol.* 163(7), 3801–3811 (1999).
89. Lanteri M, Giordanengo V, Hiraoka N *et al.*: Altered T cell surface glycosylation in HIV-1 infection results in increased susceptibility to galectin-1-induced cell death. *Glycobiology* 13(12), 909–918 (2003).
90. Rappl G, Abken H, Muche JM *et al.*: CD4⁺CD7⁺ leukemic T cells from patients with Sezary syndrome are protected from galectin-1-triggered T cell death. *Leukemia* 16(5), 840–845 (2002).
91. Cabrera PV, Amano M, Mitoma J *et al.*: Haploinsufficiency of C2GnT-I glycosyltransferase renders T lymphoma cells resistant to cell death. *Blood* (2006) (In Press).
92. Rabinovich GA, Ariel A, Hershkoviz R *et al.*: Specific inhibition of T-cell adhesion to extracellular matrix and proinflammatory cytokine secretion by human recombinant galectin-1. *Immunology* 97(1), 100–106 (1999).
93. Rabinovich GA, Ramhorst RE, Rubinstein N, *et al.*: Induction of allogenic T-cell hyporesponsiveness by galectin-1-mediated apoptotic and non-apoptotic mechanisms. *Cell Death Differ.* 9(6), 661–670 (2002).
94. Vespa GN, Lewis LA, Kozak KR *et al.*: Galectin-1 specifically modulates TCR signals to enhance TCR apoptosis but inhibit IL-2 production and proliferation. *J. Immunol.* 162(2), 799–806 (1999).
95. Chung CD, Patel VP, Moran M, Lewis LA, Miceli MC: Galectin-1 induces partial TCR zeta-chain phosphorylation and antagonizes processive TCR signal transduction. *J. Immunol.* 165(7), 3722–3729 (2000).
- **References [94] and [95] support a role for galectin-1 in the regulation of proximal T-cell receptor signals.**

96. Demetriou M, Granovsky M, Quaggin S, Dennis JW: Negative regulation of T-cell activation and autoimmunity by Mgat5 *N*-glycosylation. *Nature* 409(6821), 733–739 (2001).
- **Elegant study demonstrating both *in vitro* and *in vivo* the impact of *N*-glycosylation in the regulation T-cell responses and autoimmunity.**
97. Rabinovich GA, Daly G, Dreja H *et al.*: Recombinant galectin-1 and its genetic delivery suppress collagen-induced arthritis via T cell apoptosis. *J. Exp. Med.* 190(3), 385–398 (1999).
- **Demonstrates the therapeutic potential of galectin-1 gene delivery in an experimental model of rheumatoid arthritis and the ability of this protein to ameliorate inflammatory disease by promoting T-cell apoptosis and Th2 cytokine bias.**
98. Santucci L, Fiorucci S, Cammilleri F *et al.*: Galectin-1 exerts immunomodulatory and protective effects on concanavalin A-induced hepatitis in mice. *Hepatology* 31(2), 399–406 (2000).
99. Santucci L, Fiorucci S, Rubinstein N *et al.*: Galectin-1 suppresses experimental colitis in mice. *Gastroenterology* 124(5), 1381–1394 (2003).
100. Baum LG, Blackall DP, Arias-Magallano S *et al.*: Amelioration of graft versus host disease by galectin-1. *Clin. Immunol.* 109(3), 295–307 (2003).
101. Toscano MA, Commodaro AG, Ilarregui JM, *et al.*: Galectin-1 suppresses autoimmune retinal disease by promoting concomitant T helper (Th)2- and T regulatory mediated anti-inflammatory responses. *J. Immunol.* 176(10), 6323–6332 (2006).
102. Offner H, Celnik B, Bringman TS *et al.*: Recombinant human beta-galactoside binding lectin suppresses clinical and histological signs of experimental autoimmune encephalomyelitis. *J. Neuroimmunol.* 28(2), 177–184 (1990).
103. van der LJ, van den BA, Blokzijl T *et al.*: Dimeric galectin-1 induces IL-10 production in T-lymphocytes: an important tool in the regulation of the immune response. *J. Pathol.* 204(5), 511–518 (2004).
104. Cortegano I, del P, V, Cardaba B *et al.*: Galectin-3 down-regulates IL-5 gene expression on different T cell types. *J. Immunol.* 161 (1), 385–389 (1998).
105. Lopez E, del PV, Miguel T *et al.*: Inhibition of chronic airway inflammation and remodeling by galectin-3 gene therapy in a murine model. *J. Immunol.* 176(3), 1943–1950 (2006).
106. Zuberi RI, Hsu DK, Kalayci O *et al.*: Critical role for galectin-3 in airway inflammation and bronchial hyperresponsiveness in a murine model of asthma. *Am. J. Pathol.* 165(6), 2045–2053 (2004).
- **Reference [104–106] provide the cellular bases for a role of galectin-3 in the regulation of allergic inflammation.**
107. Poirier F: Roles of galectins *in vivo*. *Biochem. Soc. Symp.* 69, 95–103 (2002).
108. Sturm A, Lensch M, Andre S *et al.*: Human galectin-2: novel inducer of T cell apoptosis with distinct profile of caspase activation. *J. Immunol.* 173(6), 3825–3837 (2004).
109. Ozaki K, Inoue K, Sato H *et al.*: Functional variation in LGALS2 confers risk of myocardial infarction and regulates lymphotoxin-alpha secretion *in vitro*. *Nature* 429(6987), 72–75 (2004).
- **References [108] and [109] are pioneer studies demonstrating the first functional role of galectin-2 in the regulation of inflammation.**
110. Christensen MB, Lawlor DA, Gaunt TR *et al.*: Genotype of galectin-2 (LGALS2) is associated with insulin-glucose profile in the British Women's Heart and Health Study. *Diabetologia* 49(4), 673–677 (2006).
111. Kuwabara I, Kuwabara Y, Yang RY *et al.*: Galectin-7 (PIG1) exhibits pro-apoptotic function through JNK activation and mitochondrial cytochrome c release. *J. Biol. Chem.* 277(5), 3487–3497 (2002).
112. Tsuchiyama Y, Wada JU, Zhang H. *et al.*: Efficacy of galectins in the amelioration of nephrotoxic serum nephritis in Wistar Kyoto rats. *Kidney Int.* 58(5), 1941–1952 (2000).
113. Baba M, Wada J, Eguchi J *et al.*: Galectin-9 inhibits glomerular hypertrophy in db/db diabetic mice via cell-cycle-dependent mechanisms. *J. Am. Soc. Nephrol.* 16(11), 3222–3234 (2005).
114. Danguy A, Camby I, Kiss R: Galectins and cancer. *Biochim. Biophys. Acta* 1572(2–3), 285–293 (2002).
115. van den Brule FA, Califice S, Castronovo V: Expression of galectins in cancer: a critical review. *Glycoconj. J.* 19(7–8), 537–542 (2004).
- **References [114] and [115] are comprehensive reviews on the expression and role of galectins in cancer.**
116. Rubinstein N, Alvarez M, Zwirner NW *et al.*: Targeted inhibition of galectin-1 gene expression in tumor cells results in heightened T cell-mediated rejection; a potential mechanism of tumor-immune privilege. *Cancer Cell* 5 (3), 241–251 (2004).
- **Demonstration that inhibition of galectin-1 gene expression in tumor cells results in heightened T-cell mediated tumor rejection, thereby validating galectin-1 as a target for cancer therapy in an experimental model.**
117. Harjacek M, Diaz-Cano S, De Miguel M *et al.*: Expression of galectins-1 and -3 correlates with defective mononuclear cell apoptosis in patients with juvenile idiopathic arthritis. *J. Rheumatol.* 28(8), 1914–1922 (2001).
118. Romero MD, Muiño JC, Bianco GA *et al.*: Circulating anti-galectin-1 antibodies are associated with the severity of ocular disease in autoimmune and infectious uveitis. *Invest. Ophthalmol. Vis. Sci.* 47(4), 1550–1556 (2006).
119. Rabinovich GA, Cumashi A, Bianco GA *et al.*: Synthetic lactulose amines: novel class of anticancer agents that induce tumor-cell apoptosis and inhibit galectin-mediated homotypic cell aggregation and endothelial cell morphogenesis. *Glycobiology* 16(3), 210–220 (2006).
120. Dube DH, Bertozzi CR: Glycans in cancer and inflammation – potential for therapeutics and diagnostics. *Nat. Rev. Drug Discov.* 4(6), 477–488 (2005).
- **Outstanding review discussing the impact of differential cellular glycosylation in health and disease and the development of novel therapeutic targets in inflammatory processes and cancer.**
121. Rabinovich GA: Apoptosis as a target for gene therapy in rheumatoid arthritis. *Mem. Inst. Oswaldo Cruz.* 95(1), 225–233 (2000).
122. Blixt O, Head S, Mondala T *et al.*: Printed covalent glycan array for ligand profiling of diverse glycan binding proteins. *Proc. Natl Acad. Sci. USA* 101(49), 17033–17038 (2004).
123. Battig P, Saudan P, Gunde T, Bachmann MF: Enhanced apoptotic activity of a structurally optimized form of galectin-1. *Mol. Immunol.* 41(1), 9–18 (2004).
124. Tejler J, Lefler H, Nilsson UJ: Synthesis of *O*-galactosyl aldoximes as potent LacNAc-mimetic galectin-3 inhibitors. *Bioorg. Med. Chem. Lett.* 15(9), 2343–2345 (2005).

Affiliations

- *Greg Parsonage*
University of Birmingham, Department of Rheumatology, Division of Immunity & Infection, Institute of Biomedical Research, Edgbaston, B15 2TT, UK
Tel.: +44 121 414 6777;
Fax: +44 121 414 5475

- *Emily Trebilcock*
University of Birmingham, Department of Rheumatology, Division of Immunity & Infection, Institute of Biomedical Research, Edgbaston, B15 2TT, UK
Tel.: +44 121 414 6777;
Fax: +44 121 414 5475
- *Marta A Toscano*
Hospital de Clinicas "Jose de San Martin", Division of Immunogenetics, Faculty of Medicine, University of Buenos Aires, Buenos Aires, Argentina
Tel.: +54 115 950 8755;
Fax: +54 115 950 8758
- *Germán A Bianco*
Hospital de Clinicas "Jose de San Martin", Division of Immunogenetics, Faculty of Medicine, University of Buenos Aires, Buenos Aires, Argentina
Tel.: +54 115 950 8755;
Fax: +54 115 950 8758
- *Juan M Ilarregui*
Hospital de Clinicas "Jose de San Martin", Division of Immunogenetics, Faculty of Medicine, University of Buenos Aires, Buenos Aires, Argentina
Tel.: +54 115 950 8755;
Fax: +54 115 950 8758
- *Christopher D Buckley*
University of Birmingham, Department of Rheumatology, Division of Immunity & Infection, Institute of Biomedical Research, Edgbaston, B15 2TT, UK
Tel.: +44 121 414 6777;
Fax: +44 121 414 5475
- *Gabriel A Rabinovich*
Hospital de Clínicas "José de San Martín", Rabinovich, División Inmunogenética, Facultad de Medicina, Universidad de Buenos Aires, Av. Córdoba 2351, 3 Piso (C1120), Ciudad de Buenos Aires, Argentina
Tel.: +54 115 950 8755;
Fax: +54 115 950 8758;
gabyrabi@ciudad.com.ar