Novel treatment strategies for antibody-mediated diseases: targeting long-lived plasma cells



Reinhard E Voll[†], Kirsten Neubert, Silke Meister, Eva Gückel & Joachim R Kalden

[†]Author for correspondence University of Erlangen-Nuremberg. Interdisciplinary Center for Clinical Research, IZKF-N2. Nikolaus-Fiebiger-Center of Molecular Medicine, Glückstrasse 6, 91054 Erlangen, Germany Tel.: +49 (0)9131 8539301 Fax: +49 (0)9131 8539311 rvoll@molmed.unierlangen.deand, University of Erlangen-Nuremberg, Department of Internal Medicine 3 (Rheumatology and Clinical Immunology), Krankenhausstrasse 12, 91054 Erlangen, Germany

'Whenever pathogenic antibodies are predominantly secreted by long-lived plasma cells, good treatment responses become unlikely.'

Autoantibodies are believed to contribute to the pathogenesis of multiple immune-mediated diseases. However, there are only a few diseases in which a critical role of antibodies for the pathogenesis has been convincingly demonstrated. A good correlation of autoantibody titers with the disease activity, and the detection of such antibodies in most patients with a certain disease or disease manifestation, as well as the detection of autoantibodies bound to damaged organ structures, may provide an initial indication of a contribution of antibodies to the pathogenesis. Further evidence might come from in vitro experiments with exposure of isolated antibodies to primary cells or cell lines. Efficacy of immune adsorption, that is, elimination of immunoglobulins by extracorporal adsorption to an affinity matrix, such as immobilized recombinant protein A, further substantiates the role of autoantibodies in a disease entity [1,2]. The induction of disease by the transfer of autoantibodies proves their importance for the pathogenesis [2,3].

Autoantibodies may cause disease by a variety of mechanisms, the most important being:

- Direct damage of cells by the antibodies binding to surface structures, and consecutive lysis of cells by antibody-dependent cellular cytotoxicity or – presumably less important – complement-mediated lysis. The different forms of autoimmune hemolytic anemia and immune thrombocytopenia are predominantly mediated by these mechanisms. Similarly, pathogenic antibodies to double-stranded (ds) DNA may crossreact with α -actinin on the surface of mesangial cells and podocytes, and, thereby, cause kidney damage [4–6];
- Modulation of cellular function by antibodies against cell surface receptors. On the one hand, antagonistic antibodies can block binding of

ligands, as is the case with antibodies directed against the acetylcholine receptors of the skeletal muscles, thereby causing symptoms of myasthenia gravis [7]. On the other hand, agonistic antibodies may induce inappropriate signaling, as has been demonstrated in Grave's disease with antibodies against the receptor of thyroid-stimulating hormone;

- Organ and tissue damage can also be caused by immune complex formation, either in the blood or *in situ* within the target organ. Complement activation with formation of C3a and C5a attract inflammatory cells. In addition, phagocyte activation via stimulating Fc receptors causes proinflammatory cytokine release. Immune complex formation appears to be critically involved in the pathogenesis of lupus nephritis and other forms of immune-mediated nephritides, including Goodpasture's syndrome [8,9];
- Autoantibodies neutralizing functionally important soluble biomolecules can induce life-threatening conditions. For instance, antibodies to coagulation factor VIII may cause severe bleedings [10];
- So-called penetrating antibodies directed against intracellular antigens appear to cross the cytoplasmic membrane of intact cells and mediate intracellular effects [11,12].

Despite important advances during the past few decades, antibody-mediated diseases are still difficult to treat. Current therapeutic approaches including high-dose glucosteroids, cvclophosphamide, azathioprin, mycophenolate mofetil, methotrexate, high-dose intravenous immunoglobulins, plasmapheresis and immunoadsorption, even in combination, often cannot induce remission of the disease. The reasons why autoantibody production can resist even high-dose chemotherapy and allogeneic stem cell transplantation was elucidated by several highly important findings from Radbruch, Manz, Slifka, Ahmed and Hiepe: long-lived plasma cells were identified as a source of antibody memory, their role in autoimmune diseases was investigated and their surprisingly high resistance to current treatments was characterized



[13–18]. Whenever pathogenic antibodies are predominantly secreted by long-lived plasma cells, good treatment responses become unlikely. Hence, the remaining autoantibodies produced by long-lived plasma cells can perpetuate the pathogenic process.

Recent data provide evidence for the beneficial effects of B-cell-targeted therapies. The chimeric anti-CD20 antibody rituximab, which induces long-lasting depletion of CD20-positive B cells, but does not attack most plasma cell populations, has been approved for the treatment of rheumatoid arthritis (RA) [19]. Despite complete depletion of peripheral blood B cells, total immunoglobulin concentrations and many autoantibody specificities are only slightly reduced. Whereas rheumatoid factor in patients with rheumatoid arthritis and antibodies against thrombocytes in patients with refractory immune thrombocytopenia are usually markedly decreased upon rituximab treatment, in patients with systemic lupus erythematosus (SLE), antibodies to dsDNA, nucleosomes, Ro/SSA and RNP/Sm are only slightly or moderately decreased [20,21]. This discrepancy could be explained by a predominant production of autoantibodies by short-lived versus long-lived plasma cells. Whereas short-lived plasma cells need to be continuously renewed from the B-cell pool, long-lived plasma cells can survive for years [15,18]. Furthermore, amelioration of disease activity by rituximab in SLE may rather depend on inhibition of B-cell functions, such as antigen presentation and cvtokine production, than on the modest reduction of autoantibody levels. Importantly, at present, placebo-controlled trials proving the efficacy of rituximab in classical antibody-mediated diseases such as SLE are still warranted.

CD22 is another B-cell antigen that can be therapeutically targeted. The humanized monoclonal antibody epratuzumab is less efficient in depleting B cells compared with rituximab; its effects may rather be due to modulation of B-cell function via ligation of the inhibitory CD22 molecule. In a small open-label trial, epratuzumab has shown durable benefits on most body systems in SLE [22]. However, there were no significant changes in the concentrations of autoantibodies or serum immunoglobulins observed.

B-lymphocyte stimulator (BLyS) and a proliferation-inducing ligand (APRIL) are closely related members of the TNF superfamily and mediate maturation, proliferation, survival and differentiation of B lymphocytes, and support plasma cell survival [23]. Overexpression of BLyS induces hypergammaglobulinemia and autoantibody production, leading to a lupus-like disease [24,25]. Moreover, BLyS serum concentrations are elevated in many patients with SLE. Hence, the humanized BLyS neutralizing antibody belimumab was tested in a clinical Phase III trial in patients with SLE. There was a slightly reduced disease activity and less flares in belimumabtreated patients. Within 1 year of treatment, the concentrations of most autoantibodies decreased to approximately 50% of their previous concentrations. However, there was no further decrease after 3 years of belimumab treatment [26]. The TACI-Ig fusion protein atacicept employs a similar mechanism of action. TACI is one of the receptor molecules for BLyS and APRIL [23]. Therefore, the TACI-Ig fusion protein neutralizes both BLyS and APRIL. In a Phase Ib placebo-controlled trial in RA patients, atacicept has been shown to be well tolerated, and there was a trend toward clinical improvement within the 3-month treatment period. Consistent with the supposed mechanism of action, in patients who had received the highest dose of atacicept, total serum IgM decreased by approximately 50%, IgM, IgG and IgA rheumatoid factors by approximately 40%, and antibodies against cyclic citrullinated peptides by approximately 25% [27]. The modest decrease in autoantibodies achieved by neutralization of BLyS and/or APRIL will not be sufficient for an efficient treatment of antibody-mediated diseases. Nevertheless, this principle may be used as add-on therapy or for the maintenance of remission.

'The proteasome inhibitor bortezomib eliminates both short-lived and the otherwise treatment-resistant long-lived plasma cells.'

All treatments mentioned above cannot efficiently deplete long-lived plasma cells, a fact that might be responsible for their limited efficacy in autoantibody-mediated diseases. In very severely affected patients not responding to conventional therapies, high-dose chemotherapy with subsequent autologous stem cell transplantation may be an option. Only if this regimen is combined with anti-thymocyte globulin treatment can long-lived plasma cells also be eliminated. This procedure might allow a 'reset' of the immune system, and can achieve long-term remissions, including disappearance of autoantibodies. However, it is connected with a substantial risk of morbidity and mortality [28].

Apart from biologics there has been an increasing interest in research on small molecules for the treatment of autoimmune diseases. In this context, we recently described that the proteasome inhibitor bortezomib eliminates both short-lived and the otherwise treatment-resistant long-lived plasma cells in mice [29]. Bortezomib has been approved as a second-line medication for multiple myeloma, and is currently investigated in various clinical trials for the treatment of non-Hodgkin's lymphomas and other malignancies. We and others have recently demonstrated that proteasome inhibitors induce cell death of multiple myeloma cells by activation of the terminal unfolded protein response. Our laboratory demonstrated a clear correlation between the extent of antibody production, which inherently results in defective ribosomal products and unfolded proteins, and the susceptibility of myeloma cells toward proteasome inhibition [30]. Also, normal plasma cells synthesize enormous numbers of antibodies (2000 to 10,000 molecules per second), and therefore need to degrade a large number of unfolded proteins in their proteasomes. Furthermore, Roberto Sitia and colleagues demonstrated a strong decrease of the proteasome activity during differentiation of B cells into plasma cells in vitro and in vivo, a fact that makes plasma cells even more sensitive toward proteasome inhibitors, and that may be crucial to limit the lifespan of short-lived plasma cells [31,32].

To investigate if proteasome inhibitors could be used to treat autoantibody-mediated diseases, we used bortezomib in murine models of lupus. In contrast to cyclophosphamide and dexamethasone, bortezomib was able to deplete long-lived plasma cells very efficiently from spleens and bone marrows of NZB/W F1 lupus mice. Autoantibodies to dsDNA disappeared, and lupus nephritis with proteinuria was ameliorated upon bortezomib treatment. Most importantly, bortezomib-treated lupus mice survived much longer than controls. To our knowledge, this is the first therapeutic approach that can deplete virtually all plasma cells without causing overt toxic side effects in mice. Antibodies to dsDNA and cells secreting dsDNA antibodies almost completely disappeared upon bortezomib treatment, whereas total IgG concentrations were reduced by only 50%. Based on these promising results, we are currently planning a Phase I clinical trial with bortezomib in SLE patients that are refractory to conventional therapy.

The elimination of plasma cells producing pathogenic autoantibodies represents a key therapeutic goal for efficient treatment of antibodymediated diseases. In contrast to other therapeutic regimens, bortezomib can efficiently and quite selectively eliminate plasma cells in mice. However, long-lived plasma cells secreting protective antibodies against viral or bacterial pathogens are also affected. Hence, an ideal treatment strategy would just eliminate such plasma cells secreting pathogenic antibodies. A first step in this direction was carried out in the laboratory of Tchavdar Vassilev: B cells producing autoantibodies against dsDNA were eliminated using an engineered antibody-like molecule that crosslinks the DNA-specific surface immunoglobulins on autoreactive B cells with inhibitory FcyRIIb receptors. In the MRL/lpr lupus model, the appearance of anti-DNA antibodies and the disease onset were markedly delayed by intravenous injection of this bispecific antibody-like molecule [33]. However, plasma cells themselves do not express surface immunoglobulins and, hence, cannot be directly targeted by this fascinating new approach.

'A key to a successful treatment with a long-lasting response is the depletion of pathogenic antibodies producing long-lived plasma cells.'

Taken together, new treatment strategies for antibody-mediated diseases are currently being developed, which hopefully can induce remissions with acceptable side effects in most of these difficult-to-treat patients. A key to a successful treatment with a long-lasting response is the depletion of pathogenic antibodies producing long-lived plasma cells. In particular, combinations of immune adsorption to immediately reduce autoantibody load with plasma cell depletion by proteasome inhibitors, and then maintenance therapy with conventional immunosuppressants such as glucosteroids, azathioprine and methotrexate to prevent relapses, may represent a promising approach for future clinical studies.

Financial & competing interests disclosure

This work was supported by the Interdisciplinary Center for Clinical Research (IZKF, project number N2) and the German Research Society (project VO673/31 and Collaborative Research Centers SFB 643; project B3, both to REV). REV, KN, SM and JRK are named as inventors on a patent application concerning the use of proteasome inhibitors for depletion of long-lived plasma cells. The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed.

No writing assistance was utilized in the production of this manuscript.

Bibliography

- Hershko AY, Naparstek Y: Removal of pathogenic autoantibodies by immunoadsorption. *Ann. NY Acad. Sci.* 1051, 635–646 (2005).
- Petkova SB, Konstantinov KN, Sproule TJ, Lyons BL, Awwami MA, Roopenian DC: Human antibodies induce arthritis in mice deficient in the low-affinity inhibitory IgG receptor Fc γ RIIB. J. Exp. Med. 203, 275–280 (2006).
- Terato K, Hasty KA, Reife RA, Cremer MA, Kang AH, Stuart JM: Induction of arthritis with monoclonal antibodies to collagen. *J. Immunol.* 148, 2103–2108 (1992).
- Deocharan B, Qing X, Lichauco J, Putterman C: α-actinin is a cross-reactive renal target for pathogenic anti-DNA antibodies. *J. Immunol.* 168, 3072–3078 (2002).
- Mostoslavsky G, Fischel R, Yachimovich N et al.: Lupus anti-DNA autoantibodies cross-react with a glomerular structural protein: a case for tissue injury by molecular mimicry. *Eur. J. Immunol.* 31, 1221–1227 (2001).
- Smoyer WE, Mundel P, Gupta A, Welsh MJ: Podocyte α-actinin induction precedes foot process effacement in experimental nephrotic syndrome. *Am. J. Physiol.* 273, F150–F157 (1997).
- Conti-Fine BM, Milani M, Kaminski HJ: Myasthenia gravis: past, present, and future. *J. Clin. Invest.* 116, 2843–2854 (2006).
- Mohan C, Datta SK: Lupus: key pathogenic mechanisms and contributing factors. *Clin. Immunol. Immunopathol.* 77, 209–220 (1995).
- Nangaku M, Couser WG: Mechanisms of immune-deposit formation and the mediation of immune renal injury. *Clin. Exp. Nephrol.* 9, 183–191 (2005).
- Holme PA, Brosstad F, Tjonnfjord GE: Acquired haemophilia: management of bleeds and immune therapy to eradicate autoantibodies. *Haemophilia* 11, 510–515 (2005).
- Ehrenstein MR, Katz DR, Griffiths MH et al.: Human IgG anti-DNA antibodies deposit in kidneys and induce proteinuria in SCID mice. *Kidney Int.* 48, 705–711 (1995).
- Koren E, Koscec M, Wolfson-Reichlin M et al.: Murine and human antibodies to native DNA that cross-react with the A and D SnRNP polypeptides cause direct injury of cultured kidney cells. J. Immunol. 154, 4857–4864 (1995).

- Manz RA, Radbruch A: Plasma cells for a lifetime? *Eur. J. Immunol.* 32, 923–927 (2002).
- Miller JJ 3rd, Cole LJ: Resistance of long-lived lymphocytes and plasma cells in rat lymph nodes to treatment with prednisone, cyclophosphamide, 6-mercaptopurine, and actinomycin D. *J. Exp. Med.* 126, 109–125 (1967).
- Slifka MK, Ahmed R: Long-lived plasma cells: a mechanism for maintaining persistent antibody production. *Curr. Opin. Immunol.* 10, 252–258 (1998).
- Slifka MK, Antia R, Whitmire JK, Ahmed R: Humoral immunity due to long-lived plasma cells. *Immunity* 8, 363–372 (1998).
- Hoyer BF, Moser K, Hauser AE *et al.*: Short-lived plasmablasts and long-lived plasma cells contribute to chronic humoral autoimmunity in NZB/W mice. *J. Exp. Med.* 199, 1577–1584 (2004).
- Manz RA, Thiel A, Radbruch A: Lifetime of plasma cells in the bone marrow. *Nature* 388, 133–134 (1997).
- Kessel A, Rosner I, Toubi E: Rituximab: beyond simple B cell depletion. *Clin. Rev. Allergy Immunol.* 34, 74–79 (2008).
- 20. Cambridge G, Leandro MJ, Teodorescu M *et al.*: B cell depletion therapy in systemic lupus erythematosus: effect on autoantibody and antimicrobial antibody profiles. *Arthritis Rheum.* 54, 3612–3622 (2006).
- Looney RJ, Anolik JH, Campbell D *et al.*: B cell depletion as a novel treatment for systemic lupus erythematosus: a Phase I/II dose-escalation trial of rituximab. *Arthritis Rheum.* 50, 2580–2589 (2004).
- Dorner T, Kaufmann J, Wegener WA, Teoh N, Goldenberg DM, Burmester GR: Initial clinical trial of epratuzumab (humanized anti-CD22 antibody) for immunotherapy of systemic lupus erythematosus. *Arthritis Res. Ther.* 8, R74 (2006).
- Schneider P: The role of APRIL and BAFF in lymphocyte activation. *Curr. Opin. Immunol.* 17, 282–289 (2005).
- 24. Pers JO, Daridon C, Devauchelle V *et al.*: BAFF overexpression is associated with autoantibody production in autoimmune diseases. *Ann. NY Acad. Sci.* 1050, 34–39 (2005).
- Stohl W, Xu D, Kim KS *et al.*: BAFF overexpression and accelerated glomerular disease in mice with an incomplete genetic predisposition to systemic lupus erythematosus. *Arthritis Rheum.* 52, 2080–2091 (2005).

- Furie R, Petri M, Weisman MH *et al.*: Belimumab (fully human monoclonal antibody to BLys) improved or stabilized systemic lupus erythematosus (SLE) disease activity and reduced flare rate during 3 years of therapy. *Ann. Rheum. Dis.* 67, 53 (2008).
- 27. Tak PP, Thurlings RM, Rossier C *et al.*: Atacicept in patients with rheumatoid arthritis: results of a multicenter, phase Ib, double-blind, placebo-controlled, dose-escalating, single- and repeated-dose study. *Arthritis Rheum.* 58, 61–72 (2008).
- Radbruch A, Muehlinghaus G, Luger EO et al.: Competence and competition: the challenge of becoming a long-lived plasma cell. *Nat. Rev. Immunol.* 6, 741–750 (2006).
- Neubert K, Meister S, Moser K *et al*.: The proteasome inhibitor bortezomib depletes plasma cells and protects mice with lupus-like disease from nephritis. *Nat. Med.* (2008) (Epub ahead of print).
- Meister S, Schubert U, Neubert K *et al*.: Extensive immunoglobulin production sensitizes myeloma cells for proteasome inhibition. *Cancer Res.* 67, 1783–1792 (2007).
- Cenci S, Mezghrani A, Cascio P *et al.*: Progressively impaired proteasomal capacity during terminal plasma cell differentiation. *Embo. J.* 25, 1104–1113 (2006).
- Cascio P, Oliva L, Cerruti F *et al.*: Dampening Ab responses using proteasome inhibitors following *in vivo* B cell activation. *Eur. J. Immunol.* 38, 658–667 (2008).
- Tchorbanov AI, Voynova EN, Mihaylova NM *et al.*: Selective silencing of DNA-specific B lymphocytes delays lupus activity in MRL/lpr mice. *Eur. J. Immunol.* 37, 3587–3596 (2007).

Affiliations

Reinhard E Voll University of Erlangen-Nuremberg, Interdisciplinary Center for Clinical Research, IZKF-N2, Nikolaus-Fiebiger-Center of Molecular Medicine, Glückstrasse 6, 91054 Erlangen, Germany Tel.: +49 (0)9131 8539301 Fax: +49 (0)9131 8539301 Fax: +49 (0)9131 8539311 rvoll@molmed.uni-erlangen.de and, University of Erlangen-Nuremberg, Dependence of External Medicine 2

Department of Internal Medicine 3 (Rheumatology and Clinical Immunology), Krankenhausstrasse 12, 91054 Erlangen, Germany

- Kirsten Neubert University of Erlangen-Nuremberg, Interdisciplinary Center for Clinical Research, IZKF- N2, Nikolaus-Fiebiger-Center of Molecular Medicine, Glückstrasse 6, 91054 Erlangen, Germany
- Silke Meister University of Erlangen-Nuremberg, Interdisciplinary Center for Clinical Research,

IZKF- N2, Nikolaus-Fiebiger-Center of Molecular Medicine, Glückstrasse 6, 91054 Erlangen, Germany

- Eva Gückel University of Erlangen-Nuremberg, Interdisciplinary Center for Clinical Research, IZKF- N2, Nikolaus-Fiebiger-Center of Molecular Medicine, Glückstrasse 6, 91054 Erlangen, Germany
- Joachim R Kalden University of Erlangen-Nuremberg, Department of Internal Medicine 3 (Rheumatology and Clinical Immunology), Krankenhausstrasse 12, 91054 Erlangen, Germany