New advances in the treatment of Friedreich ataxia: promises and pitfalls

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Friedreich ataxia (FRDA) is an autosomal recessive, neurodegenerative disease. It affects primarily the nervous system and the heart. Progressive gait and limb ataxia, dysarthria, loss of vibration and proprioceptive sense are characteristic neurological symptoms in FRDA. In approximately 96% of patients FRDA is caused by a triplet guanine-adenine-adenine expansion within the first intron of the FXN gene on chromosome 9q13. Increased numbers of quanineadenine-adenine repeats are suggested to interfere with FXN transcription via heterochromatin-mediated silencing and result in frataxin deficiency in FRDA. Genetic and biological studies support the role of frataxin as a multifunctional protein in iron-dependent mitochondrial pathways. Multicenter, randomizedcontrolled Phase III trials in FRDA failed to prove disease modifying properties of candidate substances until to date. Phase II studies attributed idebenone, a synthetic short chain quinine analogue of co-enzyme Q10, some clinical benefit. Recent Phase III trials, however, testing idebenone have been negative or are still ongoing. Candidate substances currently tested in small randomized controlled or open-label trials are deferiprone, a mitochondrial iron chelator that forms chemically inert molecules by binding to iron, and conventional recombinant human or carbamylated erythropoietins. Both classes of candidate substances are currently under investigation to assess their efficacy and/or safety profile in Phase II trials. Pioglitazone is a peroxisome proliferator activated receptor y molecule currently tested in a 2-year randomized, doubleblind, placebo-controlled safety and efficacy study. Preclinical candidate substances in FRDA are histone deacetylase inhibitors. Promising findings in animal models will have to be replicated in human cellular models such as reprogrammed induced pluripotent stem cells from FRDA patients. A still unmet issue in FRDA is to establish well shaped clinical study designs in small study cohorts within a reasonable time frame. Therefore, large natural history studies as well as the introduction of validated bio- and surrogate markers are essential issues for future clinical trials.

> Keywords: biomarker • deferiprone • erythropoietin • frataxin • Friedreich ataxia • idebenone

Friedreich ataxia (FRDA) is an autosomal recessive, neurodegenerative disease. It is caused by a guanine-adenine-adenine (GAA) trinucleotid expansion in intron 1 of the *FXN* gene, leading to decreased expression of the mitochondrial protein frataxin. FRDA primarily affects the nervous system and the heart. Its first description derives from Nicolas Friedreich in the second half of the 19th Century. FRDA is the most common of the recessive ataxias. It seems to be restricted to individuals from Europe, the Middle East, North Africa and India. Its prevalence appears to be closely related to the frequency of large, normal alleles for *FXN* as these are virtually absent in people from East Asia and in American Indians [1]. In Europe, heterozygous

Wolfgang Nachbauer¹ & Sylvia Boesch^{†1}

¹Department of Neurology, Medical University Innsbruck, A-6020 Innsbruck, Anichstrasse 35, Austria ¹Author for correspondence: Tel.: +43 512 5040 Fax: +43 512 5042 6286 E-mail: sylvia.boesch@i-med.ac.at



mutation carriers seem to decrease in frequency with a south to north-east gradient [2-4]. Overall prevalence is approximately one in 30,000 to one in 50,000 in most populations, carrier frequency approximately one in 85 in Caucasians [3,5,6]. The high prevalence in Western Europe may be explained by a population bottleneck in one of the so called 'ice-age refugees' in which small populations survived. A two-step model merges the duplication of an ancestral (GAA)9 allele and a second mutational event that was restricted to Indo-European and North African populations with an enlargement of the (GAA)18 allele into an unstable range of more than 34 GAA units and explains the lack of FRDA in sub-Saharan Africans [1-3,7-11]. In accordance with this hypothesis, all large normal and expanded allele carriers share a common haplotype.

Clinical presentation

The age of onset in FRDA is typically around puberty, but early and late onset variants do exist. GAA repeat length of the shorter allele correlates with age of onset, accounting for approximately 50% of phenotype variation [12]. Progressive gait and limb ataxia, dysarthria, as well as loss of vibration and proprioceptive sense are characteristic neurological symptoms in FRDA. Loss of deep tendon reflexes, extended plantar response and abnormal eye movements accomplish the spectrum of neurological features often seen in FRDA. Perception of light touch, pain and temperature may initially be normal and decrease with disease progression. Pyramidal involvement and progressive weakness becomes severe mainly in late stage disease. Ataxia and balance impairment limit mobility and create the necessity of a wheelchair approximately 15 years after disease onset [13].

Non-neurological features in FRDA include hypertrophic cardiomyopathy and diabetes mellitus. Heart disease can be severe and can cause premature death, particularly in early-onset cases. Electrocardiography shows widespread T-wave inversion in virtually all patients. Conduction disturbances occur in approximately 10%. Supraventricular ectopic beats and atrial fibrillation are occasionally detectable and increase the risk of thromboembolic complications and may lead to heart failure. Echocardiography detects left ventricular hypertrophy in 50-65% of FRDA patients, showing increasing incidence in patients with larger GAA alleles [12,14]. Severe cardiomyopathy with progressive deterioration of left ventricular ejection fraction and chronic heart failure, however, is rare. The prevalence of foot deformities and scoliosis in FRDA is high [15,16]. Pes cavus can be seen in > 90% of patients, whereas range-of-motion limitations in other joints are uncommon in early FRDA. Secondary to immobility and spasticity a range of joint abnormalities may though occur. Scoliosis is seen in approximately 60% of FRDA patients, associated with severe, progressive hyperkyphosis [17,18]. Progressive curves are seen before the age of 10 years, whereas nonprogressive curves tend to present during or after puberty. Patients should be carefully screened and surgical treatment should be considered. Diabetes mellitus or impaired fastening glucose may be part of the clinical spectrum of FRDA. Oral glucose testing is therefore recommended annually [13].

Neuropathology

Characteristic neuropathological features in FRDA are atrophy of dorsal root ganglia and thinning of dorsal roots. Especially large myelinated fibers of the dorsal column tend to be sparse in FRDA. A reduction of spinal cord diameters is therefore evident and seems to be most pronounced in the thoracic region [19,20]. In addition, neuropathology in FRDA reveals subsequent atrophy of the dentate nucleus, as well as spinocerebellar and corticospinal tracts. Histopathological studies indicate for neuronal iron dysmetabolism and inappropriate myelination in FRDA [20]. Atrophy of the cerebellar hemispheres and the vermis cerebelli, however, remains mild and might only be seen in later disease stages. Peripheral neuropathy in FRDA may primarily be related to hypomyelination by deficient interaction between axons and Schwann cells. Axonal degeneration of peripheral nerves is present in FRDA but remains slow in disease progression [21].

Diagnostic criteria, clinical assessment & rating scales

Diagnostic criteria for FRDA have been suggested by Anita Harding in the 1980s [22]. They stipulate a slowly progressive, recessive ataxia that is not explained by other primary causes. Ataxia should start before the age of 25 years and is characterized by absent tendon reflexes of the lower limbs as well as dysarthria that occur within 5 years of disease onset [22]. These criteria provide for a sensitivity of 63% and a positive predictive value of 96% [23]. Lower GAA repeat numbers go along with a later onset and slower course of disease, as well as retained tendon reflexes [12].

For the assessment of ataxia in FRDA patients and the monitoring of disease progression three appropriate scales - the 'International Cooperative Ataxia Rating Scale (ICARS)' [24], the 'Friedreich Ataxia Rating Scale' (FARS) [25], as well as the recently validated 'Scale for the Assessment and Rating of Ataxia (SARA)' [26] may be used. SARA is composed out of eight items yielding in a total numeric score from 0 (no ataxia) to 40 (severe ataxia). Initially SARA was invented for rating of autosomal dominant ataxia. Validity and reliability of the scale in FRDA have been shown recently [27]. Mean time to administer SARA in patients is 14.2 ± 7.5 min (range: 5–40 min). Single SARA items comprise gait (score 0-8), stance (score 0-6), sitting (score 0-4), speech disturbance (score 0-6), finger chase (score 0-4), nose-finger test (score 0-4), fast alternating hand movements (score 0-4) and heel-shin slide (score 0-4). Limb kinetic functions are rated independently for both sides. The mean score is introduced in the total score. SARA scores significantly correlate with ICARS and FARS total scores [27]. FARS is a disease specific scale for rating ataxia severity in FRDA and thus also considers symptoms especially occurring in FRDA. FARS consists of five separate sub-scores (bulbar system, upper extremities, lower extremities, peripheral system and upright stability) and results in a score from 0 to 125 with higher scores representing more severe ataxia. Performance measures that have been modified for the use in FRDA have been introduced into the FARS. These measures include a 9-hole peg test for fine motor coordination, a timed 25-foot walk for ambulation, a speech test using the phrase 'PATA', and a low-contrast letter acuity vision test. Additionally 'activities of daily living' (ADL) and a 'functional disability scale' can be implemented into the FARS, resulting in a maximum score of 167 points [28,29]. Progression of FRDA has been measured with FARS, showing more valid data in a 2-year period than after 1 year [30]. ICARS is a widely used semi-quantitative ataxia rating scale designed to represent the classic features of ataxia. ICARS is a hundred percent scale with higher percentages indicating more severe ataxia. Postural and stance disturbances (32%) and limb ataxia (52%) engage the largest parts of this scale. The compartmentalization makes it possible to determine sub-scores [24]. ICARS has shown high inter-rater reliability in genetically confirmed ataxias [31]. Usefulness of ICARS for interventional trials was questioned recently concerning practicability and sub-scale structure [32]. Additionally several overlapping ICARS rating items have shown to lead to inconsistent rating [33].

Genetics & pathophysiology

Genetic testing of FRDA is available on a routine basis. In approximately 96% of patients FRDA is caused by a homozygous triplet GAA expansion within the first intron of the *FXN* gene on chromosome 9q13 [34]. Repeat expansions within the *FXN* gene can range from 66 repeats (normal is <40 repeats) up to more than 1000 GAA repeats. The majority of expanded alleles contain between 600 and 1200 GAA repeats. There is an inverse correlation between the age at onset, severity of disease and associated systemic symptoms with the size of the smaller GAA repeat expansion probably reflecting residual frataxin expression from the respective allele [5,6,35–37]. Approximately 4% of individuals with FRDA are compound heterozygote for a GAA expansion in one *FXN* allele and a point mutation in

the other allele resulting in distinct phenotype severity [6,32,35,38]. Penetrance is complete in homozygotes and in compound heterozygotes [34]. Tissue mosaics as often found in mitochondrial disorders may also contribute to an individual clinical phenotype [39,40].

Unlike other triplet repeat diseases such as polyglutamine expansion and RNA toxicity diseases, GAA expansions in *FXN* are intronic and do not alter the frataxin protein sequence. GAA repeat expansion leads to triplex DNA formation which may interfere with the transcription of the *FXN* gene [41–44] and heterochromatin-mediated silencing [45], resulting in limited production of frataxin. Frataxin is highly conserved across species with homologs in bacteria, yeast, plants, and animals. In humans, frataxin mRNA is translated into a precursor protein containing 210 amino acids (frataxin^{1–210}) [46,47]. Post-translational processing generates at least two frataxin isoforms (frataxin^{42–210} and frataxin^{81–210}). Recent reports suggest both isoforms relevant to FRDA pathophysiology [48].

Despite intensive research, the exact physiological functions of frataxin remain the subject of debate. Genetic and biological studies support a pivotal role of frataxin as a multifunctional protein in different irondependent mitochondrial pathways [34,49]. Frataxin is suggested to act as a mitochondrial iron chaperone [50] or as an iron sensor regulating the iron-sulfur (Fe-S) cluster biogenesis [51]. In vitro studies suggest that iron binding may trigger the oligomerization of frataxin and lead to radical scavenging of toxic iron in a bioavailable form in consequence [52]. Both monomeric and oligomeric forms of frataxin were shown to interact with various potential iron acceptors. In vitro frataxin was shown to interact with ferrochelatase and to provide the iron that is needed in the last step of heme synthesis [53,54]. Frataxin may also interact with mitochondrial aconitase, a Fe-S-containing protein, which protects against the disassembly of the Fe-S cluster by facilitating iron transfer to aconitase [55]. Moreover, both monomeric and oligomeric forms of frataxin were proposed to be the iron donor protein for de novo Fe-S cluster biosynthesis [51,56-61]. Fe-S clusters are critical prosthetic groups present in proteins involved in essential cellular processes ranging from nuclear genome stability to protein translation in mitochondrial metabolism [62]. De novo Fe-S cluster assembly, a mitochondrial process in eukaryontes, relies on the assembly of a Fe-S cluster on a scaffold protein (ISCU) from inorganic iron and sulfur, followed by the transfer of the scaffold bound Fe-S cluster to the target apoproteins. Both the synthesis and the final transfer to apoproteins require the help of additional proteins [63]. The exact function of these proteins is currently unknown. Still, in vitro iron loaded human frataxin has been shown to deliver iron to ISCU [50]. The iron-donor function of frataxin has recently been challenged in vitro since it behaves as an iron-dependent inhibitor of Fe-S cluster assembly through specific interaction with scaffold proteins in kinetic studies of Fe-S cluster biosynthesis [51]. Impaired mitochondrial iron handling in FRDA and mitochondrial iron accumulation causes in consequence an impairment of respiratory chain function and contributes to increased oxidative stress and cellular damage [64-66]. Loss of iron sulfur proteins including the respiratory chain complexes I, II, III and aconitase result in reduced ATP generation as confirmed in FRDA ataxia patients by MRI [67]. However, the level of importance of an oxidative stress component in FRDA has been discussed controversially. Still, deficient mitochondrial ATP production in tissue that depends on oxidative phosphorylation such as the nervous system is likely to be responsible for cellular dysfunction and cell death. Moreover, if oxidative stress via dysfunctional lipid peroxidation, impaired scavenging of superoxide radicals and reduced ATP formation may play a role in disease progression of FRDA remains to be elucidated.

Biomarker & surrogate markers Frataxin

Assays to quantitatively measure the amount of frataxin protein have been established recently [68,69]. Frataxin levels are usually quantified per µg total protein content to allow standardized measures. Absolute frataxin levels, however, vary considerably in recently published studies, which may be addressed to distinct cell lysis protocols and frataxin protein detection methods. Compared with healthy controls FRDA patients show a mean residual frataxin expression of 20-35% [68-71]. The amount of frataxin expression is inversely correlated to the number of GAA repeats. Late onset FRDA patients therefore display higher frataxin levels ranging from 40-90% of control values [70]. Until to date frataxin measurements have mainly been performed out of isolated peripheral blood mononuclear cells and cultured cell lines of FRDA patients. Recent reports also suggest buccal cells, whole blood and skeletal muscle [71,72] as appropriate specimens for frataxin detection. Still, considering frataxin's important role as a biomarker in clinical trials the range of frataxin levels in normal controls, FRDA carriers and FRDA patients in different tissues should be studied in depth.

Markers of oxidative stress

Oxidative stress is caused by the presence of any of a number of reactive oxygen species, which the cell is unable to counterbalance. This may result in damage to one or more biomolecules including DNA, RNA, proteins and lipids. Detection of oxidative stress may be based on DNA/RNA damage, lipid peroxidation, protein oxidation or the detection of reactive oxygen species in general. In principle, a sole assay addressing oxidative stress may be rather nonspecific for the detection of neurodegenerative cell loss in FRDA patients. Moreover, reactive oxygen species markers in patients may be afflicted by multiple confounders during clinical trials.

MRI techniques

MRI has been used as a biomarker in a first clinical FRDA trial by measuring iron content in the dendate nucleus [73]. In addition to special iron detection methods, new MRI techniques such as voxel-based morphometry, fiber tracking or MR-spectroscopy are prone to become valid in vivo surrogate markers in future clinical studies. Several nontherapeutic studies using voxel-based morphometry have revealed a correlation of patients' clinical scores and disease duration with brain white matter atrophy [74-76]. Detection of neurochemical patterns in specific brain areas using MR-spectroscopy was suggested to differentiate distinct ataxias [77]. MRI scans with higher field force (3-5 tesla) will provide for better spatial differentiation. The validity of MRI-techniques as a biomarker in therapeutic trials, however, warrants further investigation.

Cardiac MRI and spectroscopy are adequate methods for detection of interventricular septum thickness, left ventricular mass and estimation of mitochondrial function [78,79]. Their relevance as a biomarker, however, is disputable as cardiomyopathy is not evident in all FRDA patients and ataxia severity lacks correlation with cardiac hypertrophy. Application of a cardiac MRI in clinical trials will therefore be limited to therapeutic compounds addressing cardiac involvement in FRDA.

Clinical rating scales

Precision in ataxia rating is limited because of the semi-quantitative approach in all ataxia rating scales. Moreover, both inter- and intra-rater variability as well as ceiling effects in case of patients with more advanced disease stages contribute to a lack of accuracy in clinical rating. New tools and improvement of rating scales will be necessary for small but significant clinical benefit in future trials [80]. The introduction of quantitative clinical measurements (based on existing clinical ataxia symptoms) using electronic-based portable devices would therefore be helpful. Additionally, functional scores and speech assessment have to be developed to improve clinical outcome measures.

Clinical studies

Most therapeutic approaches in FRDA focus on antioxidant treatment to protect mitochondria from oxidative stress and iron accumulation. Alternatively, therapy strategies also target enhancing the transcription or protein stabilization of frataxin. Nevertheless, until to date randomized-controlled trials in FRDA are sparse. The following paragraphs provide an overview of candidate substances tested for the use in FRDA, ongoing trials and future perspectives. PubMed was searched for articles published before March 2011 including the search terms 'Friedreich ataxia' in combination with 'therapy' and 'treatment'. Studies conducted before availability of genetic testing [34] were given low priority. Ongoing clinical trials were identified via the database of the US NIH [201].

Idebenone

The basic pathophysiological approach of impaired mitochondrial function due to oxidative stress has led to broad application of antioxidant agents for the treatment of FRDA. The majority of clinical therapeutic studies were carried out using idebenone, a synthetic short chain quinine analogue of co-enzyme Q10. It acts as an electron carrier in the mitochondrial respiratory chain and therefore allocates the production of ATP. Besides, idebenone inhibits lipid-peroxidation, serves as a free radical scavenger and may therefore additionally prevent mitochondria from oxidative damage [81]. After absorption idebenone is rapidly metabolized and conjugated into several metabolites showing dose proportional pharmacokinetics in healthy human subjects up to 2250 mg/day [82,83]. However, cerebrospinal fluid analysis suggest idebenone to be less distributed to the brain than to other tissues [84].

Several clinical trials have been conducted with idebenone in FRDA patients. A majority, however, are lacking controlled or blinded study designs. A single-center randomized placebo-controlled trial (Phase II) assessed idebenone in 47 children using different dosages of idebenone (5, 15 and 45 mg/kg). Primary end point was a change in 8-hydroxy-2'-deoxyguanosine, a urinary marker of oxidative stress, after 6 months of treatment. Secondary outcome measures included ataxia rating (ICARS and FARS) and measures of ADL. Whereas the primary outcome was failed, a subgroup analysis of ambulatory patients showed significant improvement in ICARS. Moreover, higher dosages of idebenone were suggested to be superior compared with the lower dosage and placebo [28]. Two subsequent multicenter, randomized-controlled Phase III trials (MICONOS and IONIA) were carried out to assess efficacy of idebenone treatment for neurological and cardiac symptoms in FRDA. IONIA included seventy ambulatory children (age 8-18 years) with genetic proven FRDA. Participants were randomized into three treatment arms (placebo, low-dose and high-dose idebenone). Primary outcome was change in ICARS score after 6 months.

Secondary outcomes included changes in FARS rating, performance measures and ADL. IONIA failed to show significant improvement compared with placebo in each of the two end points [85]. Moreover, idebenone did not decrease left ventricular hypertrophy or alter cardiac function [86]. Upcoming results of the MICONOS trial (more than 200 ambulatory as well as wheelchair bound FRDA patients for a study duration of 1 year) will enlighten the issue if larger studies of longer duration are prone to assess the therapeutic potential of idebenone in FRDA [87]. The primary outcome of the MICONOS study is absolute change in ICARS from baseline to year one. Secondary outcome measures include reduction of left ventricular mass index detected by MRI and echocardiography as well as the improvement in peak workload assessed by a modified exercise test.

Numerous earlier studies using idebenone 5 mg/kg/ day showed significant impact on cardiac hypertrophy determined by echocardiography [88–91], although only one study was conducted as a randomized-controlled trial [91]. Still, these results could not be replicated in other studies [92,93]. Open label studies investigating clinical effects of idebenone treatment in FRDA patients showed decrease in ICARS Score [92] and stabilization of motor symptoms [94].

Until to date no multicenter, randomized-controlled trial showing idebenone to have clinical benefit on neurological symptoms in FRDA has been reported. Positive effects of idebenone on cardiac hypertrophy are still lacking clinical significance [95]. By all means idebenone therapy has been shown to be safe and well tolerated as solely low side effects of high dosages have been reported in FRDA patients of all ages [28,87,96]. Benefit of idebenone treatment in FRDA to stabilize disease progression may additionally depend on disease stage and age at initiation of idebenone treatment [92,94], which has been insufficiently considered in clinical trials so far. Table 1 gives an overview of recent idebenone trials in FRDA.

Other antioxidants

Apart from idebenone co-enzyme Q10 and vitamin E have been suggested for antioxidant treatment of FRDA. Co-enzyme Q10 plays a role in mitochondrial ATP production. Vitamin E is a naturally occurring lipid soluble antioxidant. The combination of co-enzyme Q10 and vitamin E revealed improvement of energy metabolism in cardiac- and skeletal muscle assessed with MRI in an open label trial on ten FRDA patients. Moreover changes in ICARS score suggested stabilization of disease progression as compared with a cross sectional group of FRDA patients [97,98]. Evidence for L-carnitin is based on one double-blind crossover placebo-controlled trial, though conducted in a heterogeneous study population

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Table 1. Recent idebenone trials in Friedreich ataxia and their main neurological and cardiac findings.						
Trial	Study design	No. of patients	Study duration (months)	Main neurological findings	Main cardiac findings	Ref.
Rustin <i>et al.</i> (1999)	Open-label	3	4–6	N/A	Decrease of left-ventricular mass index	[88]
Schols <i>et al.</i> (2001)	Randomized- controlled	9	1.5	No changes in clinical ataxia rating	No changes in echocardiographic measures	[93]
Artuch <i>et al.</i> (2002)	Open-label	9	12	Reduction in ICARS score after 3 months, correlation between idebenone serum values and ICARS difference after 12 months	No changes in echocardiographic measurements	[92]
Hausse <i>et al.</i> (2002)	Open-label	38	24	N/A	20% reduction in left ventricular mass and improvement in shortening fraction	[89]
Mariotti <i>et al.</i> (2003)	Randomized- controlled	28	12	No improvement in clinical rating	Significant reduction of interventricular septal thickness and left ventricular mass	[91]
Buyse <i>et al.</i> (2003)	Open-label	8	12	Progressive ataxia	Reduction of left ventricular mass index	[90]
Di Prospero <i>et al.</i> (2007)	Randomized- controlled	48	6	Higher doses associated with neurological improvement	N/A	[96]
Ribai <i>et al.</i> (2007)	Open-label	104	6–84	Worsening of ICARS score over time	Decrease of left ventricular mass index without improvement of cardiac function	[117]
Pineda <i>et al.</i> (2008)	Open-label	24	36–60	Stabilization in pediatric patients, progression in adult patients	No progression of cardiomyopathy in pediatric and adult patients	[94]
Lynch <i>et al.</i> (2010), Lagedrost <i>et al.</i> (2011)	Randomized- controlled	70	24	No significant alterations	No improvement of echocardiographic or ECG parameters	[85,86]
ECG: Electrocardiography; ICARS: International Cooperative Ataxia Rating Scale; N/A: Not available.						

of ataxias. FRDA patients showed improvement of coordination and muscle tone [99]. A randomized placebocontrolled trial in FRDA patients could not confirm these results. No improvement on echocardiographic or neurological features could be achieved [100].

Deferiprone

The pathophysiological evidence of mitochondrial iron accumulation in FRDA has led to the consideration of implementing iron chelation as a therapeutic strategy. The iron chelator deferiprone forms chemically inert molecules by binding to iron. In contrast to conventional iron chelators deferiprone is capable to cross the blood brain barrier and target mitochondria [101]. Deferiprone was tested in a first open label trial on nine adult FRDA patients over 6 months. Magnetic resonance imaging revealed significant decrease of iron content in the dentate nucleus [73]. Moreover, modest clinical improvement as measured by the ICARS scale was found. Hematopoietic or neurological side effects did not occur within this dosage regimen. However, *in vitro* data suggest loss of aconitase activity by excessive iron chelation [102]. Results of a 1 year randomized, double-blind, placebo-controlled Phase II study with the primary outcome safety and tolerability are underway.

Combined therapy with idebenone & deferiprone

A prospective open-label single-arm clinical trial in 20 FRDA patients combining idebenone and deferiprone for 11 months revealed improvement in iron deposition in deep cerebellar nuclei measured by MRI. While cardiomyopathy improved significantly, neurological functions remained stable. Side effects comprised neutropenia and the reduction of plasma iron parameters [103].

Pioglitazone

Pioglitazone is a peroxisome proliferator activated receptor γ molecule. It induces the expression of enzymes involved in mitochondrial metabolism including superoxide dismutase. Pioglitazone crosses the blood brain barrier and is suggested to improve antioxidant defense mechanisms. A 2-year randomized, double-blind, placebo-controlled safety and efficacy study in FRDA with the primary out-come measure of stabilization in ICARS is currently ongoing.

Erythropoietins

Due to their neuroprotective capacities erythropoietins have received considerable attention within the last years [104,105]. In vitro erythropoietin incubation of isolated lymphocytes and fibroblasts of FRDA patients has led to upregulation of the protein frataxin without affecting mRNA expression [106,107]. Open label studies in FRDA patients administering subcutaneously recombinant human erythropoietin (rhuEPO) have been conducted in different dosages and regimens. Continuous rhuEPO application (three-times weekly) showed frataxin upregulation of 27% (range: 15-63%) after 2 months in ten patients. Moreover, reduction of oxidative stress parameters as measured by peroxides and 8-hydroxy-2'deoxyguanosine were shown [108]. A further open label extension study on eight out of these ten patients revealed clinical improvement as measured by SARA and FARS rating after a study period of 6 months [109]. Intermittent high dose application of rhuEPO in monthly intervals resulted in a cumulative long lasting increase in frataxin levels without clinical improvement in two open label studies on FRDA patients [110,111]. Due to the well known hematopoietic stimulation potential of erythropoietin close meshed monitoring of blood cell count is essential. Elevated hemoglobin and hematocrit levels require phlebotomies, especially in continuous dosing regimens. Though carbamylated erythropoietin could provide the benefit of increase in frataxin levels [112] without affecting red blood cell count or iron metabolism. Currently a first clinical Phase II trial using carbamylated erythropoietin in FRDA is underway.

Histone deacetylase inhibitors

Histone deacetylase (HDAC) inhibitors are promising candidate substances for the future treatment as they may directly reverse the primary cause deficient frataxin expression in FRDA. HDAC inhibitors may act on DNA transcription by reversing heterochromatinmediated silencing of the *FXN* gene with subsequent increase of frataxin mRNA expression and protein in lymphoid cell lines of FRDA patients [45]. HDAC inhibitors have also been shown to increase frataxin expression in brain and heart in a KIKI mouse model for FRDA [113]. Moreover, FRDA disease phenotype could partially be reversed in a GAA repeat expansion YG8R mouse model after 5 months of HDAC inhibitor tor treatment [114]. In murine models HDAC inhibitors were well tolerated [113–115], their safety and efficacy profile in humans, however, requires further investigation. Preclinical trials are currently underway.

Future perspective

Promising advances for the treatment of FRDA rely on several pathophysiological considerations (Figure 1). The prevention of oxidative stress through an improvement of mitochondrial respiratory chain function and the allocation of ATP production underlies the use of idebenone, a synthetic short chain quinine analogue of co-enzyme Q10. Unfortunately, after some hope during Phase II clinical trials, results of recent Phase III clinical trials using idebenone to achieve clinical benefit were somehow disappointing [85]. Also deferiprone, an intracellular iron chelator entered into a clinical Phase II studies. While findings in MRI surrogates appeared to be promising, secondary clinical outcome measures did not detect consistent benefit [73]. A second Phase II study is currently addressing safety and tolerability and will provide for further data. Recent advances in the understanding of mitochondrial iron metabolism point to a complex interaction between mitochondrial iron content and frataxin function. In the light of these findings the role of iron chelators as a long-term therapeutic approach in FRDA will have to be reconsidered. Pioglitazone is suggested to improve antioxidant defense mechanisms and is currently tested in a 2-year randomized, double-blind, placebo-controlled safety and efficacy study in FRDA. Erythropoietins are candidate substances in FRDA therapy. Multimodal properties of erythropoietins may explain laboratory findings, namely an upregulation of cellular frataxin content. They may also explain clinical improvement through symptomatic effects on muscle strength and endurance, similar to enhanced motor performance in athletes with illicit use. Erythropoietins are also prone to influence frataxin function per se considering their multiple functions on intracellular and mitochondrial iron handling. Thus, conventional erythropoietins have considerable short-comings in their safety profile because of their hematopoietic side effects. Therefore nonhemotopoietic derivatives of erythropoietin such as carbamylated erythropoietin may have a role in future FRDA therapy [112]. A clinical Phase II study with carbamylated erythropoietin in FRDA is currently under way. Only recently, preclinical studies using HDAC inhibitors in cell cultures and in mouse models provided for promising data in FRDA [45,113]. Reversing heterochromatin-mediated silencing of the FTX gene with subsequent increase of frataxin is an elegant and

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associated relevant therapeutic point of application. GAA: Guanine-adenine-adenine; HDAC: Histone deacetylase; rhuEPO: Recombinant human erythropoietin.

Disease modifying treatment options may address either neuroprotection or neurorescuing, since the regeneration potency in neuronal tissue appears to be remote. Therefore large natural history studies, validated biomarkers and disease progression markers are pivotal in FRDA. A large natural history study has been launched in Europe with support of the European Community only recently (European Friedreich Ataxia Consortium for Translational Studies). Remarkable efforts have been made to validate scales for the assessment of ataxic symptoms. Thus, semi-quantitative measures remain prone to be afflicted by subjective interpretation. Therefore, the development of quantitative tools for the assessment of ataxia is of importance for clinical studies. The design of future clinical studies in FRDA will be challenged by the following issues:

• To develop tailored study schedules in small study populations for substance testing with appropriate tools and in applicable time frames;

• To bear in mind that clinical effects of candidate substances may not reverse neuronal cell loss in advanced disease stage;

• To give patients afflicted by rare disease the right to participate in clinical trials regardless of their disease stage.

To date, these issues represent major pitfalls in disease-modifying therapeutic trials in FRDA.

tempting therapeutic approach. Still, considering limited experience in epigenetic therapy approaches in general and concerns about the specificity of HDAC inhibitors *in vivo* have to be overcome. New techniques such as induced pluripotent stem cells from FRDA patients [116] that are re-differentiated to neuronal cell lines will have major impact on candidate substance development and testing in the near future. For the first time candidate substances will be developed in human disease specific cell lines and adapted for special needs in distinct neuro-degenerative conditions.

Financial & competing interest disclosure

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Executive summary

- Therapeutic approaches in Friedreich ataxia (FRDA) focus on antioxidant treatment, as well as on enhancing frataxin transcription or protein stabilization.
- Until to date no multicenter randomized controlled trial showed clinical benefit of idebenone on neurological symptoms in FRDA. A recent Phase III trial in pediatric FRDA patients failed to show changes in neurological rating scales, as well as cardiac symptoms.
- Erythropoietins and deferiprone have been suggested as appropriate candidate substances by open label studies and *in vitro* findings. Ongoing Phase II trials will provide for additional information on safety and efficacy on these compounds in the near future.
- Reversing heterochromatin-mediated silencing of the frataxin gene as shown *in vitro* by histone deacetylase inhibitors is a new and elegant therapeutic approach.
- New techniques such as induced pluripotent stem cells of FRDA patients will have major impact on candidate substance development and testing.
- Clinical trials will have to use appropriate tools and consider applicable study time frames for efficacy of candidate substances. The right of patients in rare disease to participate in clinical studies even in advanced disease stages has to be taken into account for future study designs.

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