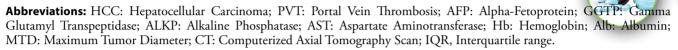
HCC with low- and normalserum alpha-fetoprotein levels



A large database of 1773 HCC patients in Turkey was examined. 41.9% had alpha-fetoprotein (AFP) levels <20 IU/ml and an additional 16.123% had values between 20-100 IU/ml. This 58% of the cohort (<100 IU/ml AFP levels) was examined in detail. 66% of patients with small (<5 cm) HCCs had low AFP, compared to 49% of patients with larger (>5 cm) HCCs. The mean diameter (MTD) of larger MTD, low AFP tumors was 8.4cm. Therefore, factors other than AFP must contribute to HCC tumor growth. Larger tumors in low AFP patients had both higher platelet levels and increased PVT percent. Linear regression analysis for both MTD and multifocality showed that platelet numbers and presence of PVT were significant variables; whereas for PVT, significant variables were albumin, alkaline phosphatase and MTD. Comparisons between patients with AFP levels <20, 20-<100, 100-<1000 and >1000 IU/ml showed the most significant tumor finding was an increase in PVT percent between each group, and to a lesser extent, MTD. Thus, low- or normal-AFP HCCs constitute the majority of patients and have slightly lower MTD and much lower PVT percent than HCCs associated with elevated blood AFP levels. New, non-AFP markers are thus needed, especially for small HCCs

Keywords: HCC, Turkey, AFP, MTD, PVT

Introduction

Alpha-fetoprotein (AFP) was first described in 1953 as a fetal form of albumin and as a marker for HCC [1-3], which opened up the relationship between oncogenesis and embryogenesis. Since then, it has become the best-described and most used marker in HCC. It has been recognized for its usefulness in prognosis [4-7], its relationship to various indices of HCC human biology [6,8,9], its use in non-biopsy HCC diagnosis [10,11], in assessment of responses to therapy [12] and more controversially, as a screening tool together with abdominal ultrasound in patients with hepatitis or cirrhosis who are known to be at risk for HCC development [13,14]. However, not all HCC tumors are AFP positive or secrete elevated amounts of AFP into the serum [15-26]. In the present study, we examined a large cohort of Turkish HCC patients, stratified by AFP status, with particular reference to the low AFP patients and examined their characteristics. We found that their tumors were smaller than those with elevated AFP levels and were much less aggressive, as judged by percent patients with PVT. Nevertheless, low AFP HCC patients also produced large size HCCs, suggesting the presence of factors other than AFP in the determination of HCC size.

Methods

Patient data

We analyzed a database of 1773 prospectively-accrued HCC patients who had full baseline tumor parameter data, including CT scan information on HCC size, number of tumor nodules and presence or absence of PVT and plasma AFP levels; complete blood count; routine blood liver function tests, (total bilirubin, GGTP, ALKP, albumin, transaminases) and patient demographics. Diagnosis was made either via tumor biopsy or according to international guidelines [6,7]. Of these patients, 1029 had low AFP levels (<100 IU/ml) and are the subject of this study. Database management conformed to legislation on privacy and this study conforms to the ethical guidelines of the Declaration of Helsinki and approval for this retrospective study on deidentified HCC patients was obtained by the Institutional Review Board.

Statistical analysis

Mean and SD for continuous variables, and relative frequency for categorical variables, were used as indices of centrality and dispersion of the distribution. For categorical variables, the Chi-

Brian I. Carr¹, Hikmet Akkiz². Oguz Üsküdar², Kendal Yalçın³, Vito Guerra⁴. Sedef Kuran², Ümit Karaoğullarından², Engin Altıntaş⁵, Ayşegül Özakyol⁶, Salih Tokmak², Tuğsan Ballı², Mehmet Yücesoy⁷, Halil İbrahim Bahçeci⁸, Abdulalh Ülkü², Tolga Akçam², Kamil Yalçın Polat⁹, Nazım Ekinci¹⁰. Halis Şimşek¹¹, Necati Örmeci¹², Abdulalh Sonsuz¹³. Mehmet Demir¹⁴, Murat Kiliç¹⁵, Ahmet Uygun¹⁶,

ClinicalPractic

square and z test for proportions were used; the Wilcoxon rank-sum (Mann-Whitney) test was to test the difference between two categories, and the Kruskal-Wallis rank test to test the difference among categories.

Linear regression model was used to evaluate the associations between maximum tumor diameter on single variables examined, while logistic regression model was used to evaluate the associations between PVT (No/Yes) or Nodule number $(1/ \ge 2)$, respectively on single variables examined.

All final multiple linear or logistic regression models was obtained with the backward stepwise method and the variables that showed associations with p<0.10 were left in the models.

When testing the null hypothesis of no association, the probability level of < error, two tailed, was 0.05.

All the statistical computations were made using STATA 10.0 Statistical Software (StataCorp). 2007. Stata Statistical Software: release 10. College Station, TX: StataCorp LP, Statistical Software (StataCorp). 2007. Stata Statistical Software: release 10. College Station, TX: StataCorp LP).

Results

Comparisons amongst AFP groups

Patients were initially evaluated according to AFP level **(TABLE 1).** There were 1773 patients in the total cohort, of whom 743 (41.91%) had AFP <20 IU/ml. The second column of 284 patients (16.02%) also had low AFP levels of 20-100 IU/ml. Thus the 2 left columns of patients with low AFP levels constituted 57.92% of the total cohort of HCC patients.

When the clinical characteristics of the 4 AFP groups were examined, significant differences emerged. Examination of tumor size (MTD) across the 4 AFP groups revealed no change between the lowest 2 groups, and then significant MTD increase with subsequent increase in group. Similar results were found for tumor multifocality. However, PVT increased significantly with each increase in AFP group. This was also true between the 2 low AFP groups, despite their have similar MTDs. There were significant increases in the trends for total bilirubin, GGTP, ALKP and decrease in albumin, but they were not individually significant when all 4 groups were compared Ali Demir¹⁷, Anıl Delik², with each other. Burcu Arslan²,

For the remainder of the analysis, the 2 Figen Doran², lowest AFP groups were combined as 'low AFP' Sezai Yilmaz & i.e. $\leq 100 \text{ IU/ml}$, as the tumor sizes were similar Yaman Tokat¹⁹ for these 2 lowest AFP groups. Small (<5cm MTD) and large (>5cm MTD) tumors were ¹Izmir Biomedicine and Genome Institute, then separately dichotomized according to AFP levels (**TABLE 2**). There were 750 patients in _{Department}, Adana, Turkey the small MTD group, and of these, 501 or ³Dicle üniversitesi, Turkey 66.80% of the patients had low AFP. The low Area de Bellis Medical Center, Castellana AFP subgroup with <5cm tumors had smaller Grotte, Italy tumors and significantly less PVT % (less than ⁵Mersin üniversitesi, Turkey half) than patients with smaller tumors and "Eskişehir Osmangazi üniversitesi, Turkey higher AFP, and they also had significantly 'Erciyes üniversitesi, Turkey lower mean values for platelets, total bilirubin, ⁸Fırat üniversitesi, Turkey GGTP and ALKP and higher albumin values, "istanbul memorial hastanesi, Turkey than for patients with <5 cm tumors and higher ¹⁰Dicle üniversitesi, Turkey AFP>100. The >5 cm MTD group had 385 ¹¹Hacettepe üniversitesi, Turkey or 48.92% in the low AFP subgroup. Parallel ¹²Ankara üniversitesi, Turkey differences were found for the larger tumors ¹³istanbul Cerrahpaşa üniversitesi, Turkey with respect to MTD and PVT %, as well as 14Hatay Mustafa Kemal üniversitesi, Turkey for tumor multifocality, as well as for platelets ¹⁵*Izmir Kent hastanesi, Turkey* and GGTP levels, but ALKP and bilirubin 16Haydarpasa sultan Abdülhamid eğitim levels were not significantly different when low araştırma hastanesi, Turkey and higher AFP subgroups were compared for ¹⁷Konya Necmettin Erbakan üniversitesi, Turkey the >5 cm MTD patients. For the larger MTD ¹⁸Inonu university, Malatya patients, 30.92% had PVT, despite being in the ¹⁹Florence Nightingale hospital, Istanbul low AFP subgroup (compared to 9.36% with *Author for correspondence: PVT in the low AFP small tumor group). The brianicarr@hotmail.com larger MTD patients with low AFP had a mean MTD of 8.43 cm, compared to 2.91 cm in the small MTD tumor group with low AFP, a 2.9fold increase in size, despite both tumor groups having low AFP levels. Thus, factors other than AFP likely also contribute to HCC size increase.

Comparison of small and large low AFP patients **(TABLE 2)** showed that in addition to the size differences, there were significantly more percent PVT in the patients with larger than with smaller tumors (9.36% versus 30.92%, p,0.0001), without differences in either AFP or bilirubin mean values.

Relationships of platelets, albumin, GGTP and ALKP to HCC parameters

Three of the most important and objectively measurable indices of HCC aggressiveness include: tumor size (MTD), multifocality and presence of PVT. The regression models were therefore constructed for each of these 3 parameters are shown in **TABLE 3**, and the

Clin. Pract. (2018) 15(1)

Table 1. Comparisons among AFP (IU/mL) groups in HCC	among AFP (IU/m		patients (n=1773).								
	AFP<20	20 ≤ AFP<100	$100 \le AFP < 10000$	AFP ≥ 10000				Comparisons	risons		
Parameter*	(n=743)	(n=284)	(n=603)	(n=143)	, ₽			p-value #	lue #		
	<i>(a)</i>	(q)	(C)	<i>(q)</i>		(a)vs(b)	(a)vs(c)	(a)vs(d)	(p) vs(c)	(p)sn(q)	(c)vs(d)
Platelet counts ($10^3/\mu L$)	153.20 ± 101.64	138.14 ± 80.33	166.93 ± 97.71	184.08 ± 109.50	0.0001	0.17	0.001	0.0008	0.0001	0.0001	0.13
Hemoglobin (g/dL)	12.33 ± 2.32	12.43 ± 2.30	12.13 ± 2.23	12.19 ± 2.43	0.33	ł	I	ł	ł	ł	ł
GGTP (U/L)	129.76 ± 166.72	151.86 ± 160.79	188.04 ± 185.65	200.12 ± 147.30	0.0001	0.03	<0.0001	<0.0001	0.008	0.002	0.12
ALKP (U/L)	201.05 ± 333.87	204.31 ± 196.62	231.66 ± 234.80	232.22 ± 189.11	0.0001	0.15	<0.0001	0.002	0.09	0.06	0.43
Total Bilirubin (mg/dL)	2.43 ± 3.76	2.78 ± 3.77	3.31 ± 5.34	4.03 ± 6.23	0.002	0.50	0.002	0.003	0.14	0.04	0.22
Albumin (g/dL)	3.24 ± 0.77	3.01 ± 0.70	3.01 ± 0.74	2.86 ± 0.62	0.0001	0.0001	<0.0001	<0.0001	0.84	0.08	0.05
AFP (IU/mL)	6.77 ± 4.86	47.76 ± 22.06	1584.11 ± 2066.79	63695.33 ± 113296.00	0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
MTD (cm)	5.30 ± 3.86	5.30 ± 3.29	6.57 ± 4.14	8.39 ± 4.51	0.0001	0.28	<0.0001	<0.0001	0.0001	<0.0001	<0.0001
Nodule number (%)											
Multifocality (≥ 2)	182 (28.26)	81 (32.14)	191 (36.11)	43 (36.13)	0.03 ^	0.26 $^{\diamond}$	0.004 $^{\diamond}$	0.10 ⁴	0.27 *	0.45*	o.99 ∲
PV Thrombosis (%)	120 (19.26)	64 (25.40)	189 (36.77)	61 (50.83)	<0.001^	0.05 $^{\diamond}$	<0.0001	<0.0001	0.001 [¢]	<0.0001	0.005
Cirrhosis (%)	566 (77.11)	237 (84.34)	504 (83.86)	125 (87.41)	0.001 ^	0.007	0.002	0.001 $^{\diamond}$	0.85 *	0.38 $^{\diamond}$	0.26 $^{\diamond}$
* All values: Means±Standard Deviation as continuous; Frequences and percentage (%) as categorical. ♦ Kruskal-Wallis rank test; # Wilcoxon rank-sum (Mann-Whitney) test; ^ Chi-square test;; # Test z for proportions. Abhreviations: GGTP name duramy transpentidase: ALKP Alkaline nhosobatase: AFP Alpha-feronorotein: MTD. Maximum Tumor Diameter: PVT. Portal Vein Thrombosis.	ndard Deviation as co ;;* Wilcoxon rank-sum mma glutamyl transi	ontinuous; Frequence n (Mann-Whitney) tes peptidase: ALKP Alka	t; ^ Chi-square test;; line phosphatase: AF	 as categorical. ⁺Test z for proportior ⁺P Alpha-fetoprotein: 	sı. MTD. Maxim	umor	Diameter: P ¹	/T. Portal Ve	in Thrombo	cis.	

Table 2. Comparisons be	etween HCC patients in s	Table 2. Comparisons between HCC patients in single MTD (cm) categories, dichotomized by AFP.	s, dichotomized by AFP			
	MTD<	MTD<5.0 cm		MTD≥	MTD ≥ 5.0 cm	
	AFP ≤ 100	AFP>100	÷	AFP ≤ 100	AFP>100	, ¢
Parameter*	(n=501)	(n=249)		(n=385)	(n=402)	
Platelet counts (10³/μL)	133.08 ± 78.18	150.86 ± 93.79	0.04	176.12 ± 113.39	187.99 ± 103.97	0.03
Hemoglobin (g/dL)	12.48 ± 2.33	12.07 ± 2.22	0.04	12.41 ± 2.23	12.37 ± 2.13	0.82
GGTP (U/L)	129.23 ± 181.11	176.73 ± 168.57	<0.0001	149.74 ± 159.05	203.84 ± 187.94	<0.0001
ALKP (U/L)	167.90 ± 182.51	222.46 ± 235.99	0.0001	220.51 ± 199.11	242.72 ± 237.19	0.25
Total Bilirubin (mg/dL)	2.15 ± 2.95	3.02 ± 4.36	0.0009	2.55 ± 3.78	2.98 ± 4.82	0.17
Albumin (g/dL)	3.21 ± 0.76	3.00 ± 0.79	0.0008	3.12 ± 0.76	2.98 ± 0.68	0.04
AFP (IU/mL)	16.14 ± 20.04	6629.88 ± 24234.62	<0.0001	20.72 ± 24.51	15128.16 ± 50458.48	<0.0001
MTD (cm)	2.91 ± 1.02	3.09 ± 0.96	0.04	8.43 ± 3.62	9.30 ± 3.77	<0.0001
Nodule number (%)			0.92 ^			0.002 ^
-	362 (72.84)	179 (72.47)		272 (72.15)	234 (60.15)	
2-3	135 (27.16)	68 (27.53)		104 (27.59)	154 (39.59)	
>3	0 (0.00)	0 (0:00)		1 (0.27)	1 (0.26)	
PV Thrombosis (%)	45 (9.36)	54 (22.98)	<0.001 ^	111 (30.92)	173 (46.76)	<0.001 ^
Cirrhosis (%)	394 (79.28)	208 (83.87)	0.13 ^	292 (77.04)	334 (83.29)	0.03 ^
* All values: Means±Standard Deviation as continuous; Frequer	ird Deviation as continuou: Whitney) test; ^ Chi-squ a glutamyl transpeptidase		e (%) as categorical. : AFP, Alpha-fetoprotein; A	ATD, Maximum Tumor Dian	ıcies and Percentage (%) as categorical. kaline phosphatase; AFP, Alpha-fetoprotein; MTD, Maximum Tumor Diameter; PVT, Portal Vein Thrombosis.	.sisotr

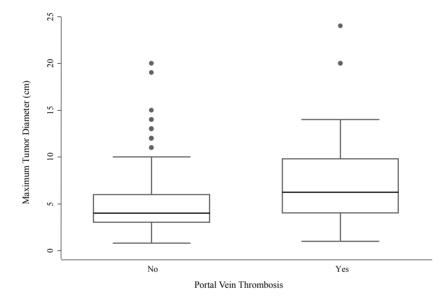
Brian I. Carr

			(A)				(B)				(C)	
Parameter	β	se(β)	٩	95% C.I.	OR	se(OR)	٩	95% C.I.	OR	se(OR)	٩	95% C.I.
Platelet counts (10³/μL)	0.013	0.002	<0.001	0.010 to 0.016	1.003	0.001	0.005	1.001 to 1.005	0.998	0.001	0.099	0.996 to 1.000
Hemoglobin (g/dL)	-0.056	0.086	0.51	-0.225 to 0.113	0.882	0.049	0.02	0.791 to 0.983	0.993	0.047	0.881	0.905 to 1.089
GGTP (U/L)	0.002	0.001	0.04	0.0001 to 0.0043	1.001	0.001	0.02	1.0002 to 1.0027	1.001	0.001	0.152	0.999 to 1.002
ALKP (U/L)	0.002	0.001	0.01	0.0006 to 0.0044	1.001	0.001	0.01	1.0003 to 1.0026	1.000	0.001	0.830	0.999 to 1.001
Total Bilirubin (mg/dL)	0.018	0.059	0.76	-0.098 to 0.133	1.055	0.036	0.11	0.988 to 1.128	0.997	0.032	0.950	0.936 to 1.064
Albumin (g/dL)	-0.175	0.247	0.48	-0.660 to 0.311	0.581	0.098	0.001	0.417 to 0.809	0.790	0.109	0.087	0.603 to 1.035
AFP (IU/mL)	0.005	0.008	0.53	-0.011 to 0.021	1.009	0.005	0.06	0.999 to 1.018	0.999	0.004	0.992	0.991 to 1.008
MTD (cm)	1	1	1	I	1.193	0.041	<0.001	1.114 to 1.276	0.972	0.029	0.335	0.917 to 1.030
Nodules number	-0.371	0.384	0.33	-1.126 to 0.384	1.507	0.373	0.10	0.928 to 2.448	ł	ł	1	ł
PV Thrombosis (yes)	2.515	0.433	<0.001	1.664 to 3.366	ł	I	1	ł	1.507	0.373	0.098	0.928 to 2.448
Cirrhosis (yes)	-0.866	0.460	0.06	-1.771 to 0.039	1.270	0.396	0.44	0.689 to 2.341	1.618	0.426	0.068	0.966 to 2.710
			(A ₁)				(B ₁)				(C ₁)	
Platelet counts (10 ³ /μL)	0.012	0.002	<0.001	0.009 to 0.015					0.998	0.001	0.070	0.996 to 1.000
ALKP (U/L)	1	1	1	1	1.001	0.0006	0.08	1.000 to 1.002	1	1	ł	I
Albumin (g/dL)	1	I	1	ł	0.605	0.108	0.005	0.426 to 0.859	I	I	I	I
MTD (cm)	ł	I	ł	I	1.187	0.042	<0.001	1.107 to 1.273	I	I	I	ł
PV Thrombosis (yes)	2.046	0.409	<0.001	1.241 to 2.851	I	I	I	I	1.571	0.401	0.077	0.953 to 2.590
# All Multiple regression final models were executed on all variables, included together in the model, and selected with stepwise method. Abbreviations: ß: coefficient; se(ß): standard error of coefficient; OR, Odds-Ratio; se(OR), standard error of Odds-Ratio; GGTP, gamma glutamyl transpeptidae; ALKP, Alkaline phosphatase; AFP, Aloha-feroprotein: MTD, Maximum Tumor Diameter: PVT. Portal Vein Thrombosis.	al models ' it; se(β): sta D. Maximu	were exec andard eri m Tumor	cuted on all ror of coeff Diameter:	l variables, included icient; OR, Odds-Rati PVT. Portal Vein Thrc	together ir io; se(OR), s mbosis.	the model, tandard erro	and selecte or of Odds-R	les, included together in the model, and selected with stepwise method. DR, Odds-Ratio; se(OR), standard error of Odds-Ratio; GGTP, gamma glut ortal Vein Thrombosis.	iod. glutamyl tra	nspeptidae; ,	ALKP, Alkalir	ie phosphatase;

significant variables are shown in **TABLE 3A-3C**. In a final model for MTD, platelets and PVT was significant (**TABLE 3A1**). The same was also found for tumor multifocality (**TABLE 3C1**). However, for PVT, the significant variables in the final model were ALKP, albumin and MTD (**TABLE 3B1**). The relationship showing an increase in PVT in larger size tumors is shown in the **FIGURE 1** box plot.

Small (<5 cm) and larger (>5 cm) MTD tumor groups were then compared, after dichotomization for the significant blood parameter variables shown in **TABLE 3**. The relationship of platelets to MTD is shown in **TABLE 4**. A significant effect of platelet dichotomization was found for MTD and PVT, but only in the larger tumors (>5 cm). In that group, the mean MTD was 3.72 cm in the thrombocytopenia subgroup, compared to 8.78 cm in the normal platelet subgroup. The relationship between blood platelet levels and MTD is also depicted in **FIGURE 2**, in which an increase in tumor size was found to be associated with a significant increase in mean platelet values, as reported in other analyses [27,28].

An examination of blood albumin levels in relation to MTD (TABLE 5) showed





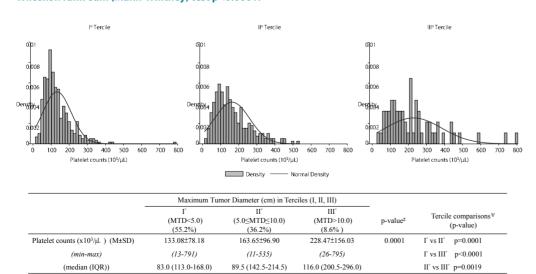


FIGURE 2.Peripheral blood platelet counts (103/ μ L) among Maximum Tumor Diameter (cm) terciles, in patients with AFP \leq 100(IU/mL). Kruskal-Wallis rank test; Mann-Whitney rank test.

categories, all with A				<u> </u>		
	MTD<5	.0 (cm)		MTD ≥	5.0 (cm)	
Parameter*	Platelets<125 (10 ³ /μL)	Platelets ≥ 125 (10 ³ /µL)	pΨ	Platelets<125 (10 ³ /µL)	Platelets ≥ 125 (10 ³ /µL)	pψ
	(n=220)	(n=177)		(n=120)	(n=202)	
Platelet counts (10 ³ /µL)	83.53 ± 24.60	194.66 ± 78.21	<0.0001	81.85 ± 25.14	232.14 ± 108.17	<0.0001
Hemoglobin (g/dL)	12.12 ± 2.30	12.94 ± 2.30	0.0009	11.98 ± 2.21	12.64 ± 2.22	0.02
GGTP (U/L)	123.69 ± 185.70	134.80 ± 176.96	0.72	110.91 ± 126.15	169.30 ± 170.59	0.01
ALKP (U/L)	146.62 ± 117.82	192.34 ± 237.53	0.77	193.74 ± 119.17	234.15 ± 227.47	0.78
Total Bilirubin (mg/dL)	2.36 ± 2.73	1.82 ± 3.19	<0.0001	3.33 ± 3.53	2.22 ± 3.88	<0.0001
Albumin (g/dL)	3.05 ± 0.71	3.41 ± 0.77	<0.0001	2.92 ± 0.67	3.23 ± 0.79	0.0007
AFP (IU/mL)	16.49 ± 20.00	16.94 ± 21.68	0.30	19.31 ± 20.55	21.67 ± 27.18	0.35
MTD (cm)	2.89 ± 1.00	3.04 ± 1.00	0.16	3.72 ± 3.06	8.78 ± 3.97	0.009
Nodule number (%)			0.70 ^			0.52 ^
1	159 (72.94)	126 (71.19)		86 (73.50)	138 (68.66)	
2-3	59 (27.06)	51.(28.81)		31 (26.50)	62 (30.85)	
>3	0 (0.00)	0 (0.00)		0 (0.00)	1 (0.50)	
PV Thrombosis (%)	20 (9.13)	23 (14.29)	0.12 ^	28 (23.73)	71 (37.37)	0.01 ^
Cirrhosis (%)	201 (91.36)	115 (66.47)	<0.001 ^	113 (95.76)	137 (68.16)	<0.001 ′

Table 4. Comparisons among HCC patients between platelet groups (<125/ \ge 125) in single MTD

* All values: Means±Standard Deviation as continuous; Frequences and Percentage (%) as categorical. [•] Wilcoxon rank-sum (Mann-Whitney) test; ^ Chi-square test.

Abbreviations: GGTP, gamma glutamyl transpeptidase; ALKP, Alkaline phosphatase; AFP, Alpha-fetoprotein; MTD, Maximum Tumor Diameter; PVT, Portal Vein Thrombosis; CRP, C-Reactive Protein.

Table 5. Compari categories in patie						
	MTD<	5.0 (cm)		MTD ≥	: 5.0 (cm)	
Parameter*	Albumin ≤ 3.5 (g/dL)	Albumi n>3.5 (g/dL)	pψ	Albumin ≤3.5 (g/dL)	Albumin >3.5 (g/dL)	pψ
	(n=247)	(n=146)		(n=221)	(n=100)	
Platelet counts (10 ³ / µL)	124.12 ± 79.69	147.38 ± 74.55	0.0001	170.53 ± 121.42	189.39 ± 94.03	0.004
Hemoglobin (g/dL)	11.77 ± 2.19	13.61 ± 2.10	<0.0001	11.72 ± 2.01	13.91 ± 1.97	<0.0001
GGTP (U/L)	144.41 ± 199.61	91.77 ± 123.27	0.001	160.45 ± 142.47	120.43 ± 196.17	0.001
ALKP (U/L)	187.20 ± 205.53	117.58 ± 91.06	0.0001	232.51 ± 205.71	188.18 ± 178.10	0.04
Total Bilirubin (mg/ dL)	2.52 ± 3.33	1.21 ± 1.34	0.0003	3.14 ± 4.26	0.98 ± 0.82	<0.0001
Albumin (g/dL)	2.75 ± 0.52	4.00 ± 0.33	<0.0001	2.70 ± 0.48	4.04 ± 0.33	<0.0001
AFP (IU/mL)	17.99 ± 21.54	14.53 ± 19.27	0.26	21.93 ± 24.18	17.48 ± 25.67	0.004
MTD (cm)	2.96 ± 1.01	2.93 ± 1.01	0.78	8.71 ± 3.94	7.71 ± 2.97	0.02
Nodule number (%)			0.47 ^			0.08 ^
1	173 (70.61)	108 (73.97)		145 (66.51)	78 (78.79)	
2-3	72 (29.39)	38 (26.03)		72 (33.03)	21 (21.21)	
>3	0 (0.00)	0 (0.00)		1 (0.46)	0 (0.00)	
PV Thrombosis (%)	30 (12.45)	13 (9.49)	0.38 ^	75 (35.71)	24 (24.74)	0.06 ^
Cirrhosis (%)	215 (87.04)	99 (69.23)	<0.001 ^	187 (85.78)	62 (62.00)	<0.001 ′

* All values: Means±Standard Deviation as continuous; Frequences and Percentage (%) as categorical. ⁴ Wilcoxon rank-sum (Mann-Whitney) test; ^ Chi-square test.

Abbreviations: GGTP, gamma glutamyl transpeptidase; ALKP, Alkaline phosphatase; AFP, Alphafetoprotein; MTD, Maximum Tumor Diameter; PVT, Portal Vein Thrombosis; CRP, C-Reactive Protein.

	able 6. Comparisons HCC patient between categories of GGTP (\leq 100/>100) in single MTC ategories, in patients with AFP \leq 100 (IU/mL).										
	MTD<5			MTD ≥	5.0 (cm)						
Parameter*	GGTP ≤ 100 (U/L)	GGTP>100 (U/L)	pψ	GGTP ≤ 100 (U/L)	GGTP>100 (U/L)	pψ					
	(n=126)	(n=78)		(n=106)	(n=77)						
Platelet counts (10 ³ /µL)	137.51 ± 78.60	150.26 ± 109.46	0.57	166.66 ± 100.95	211.26 ± 126.87	0.006					
Hemoglobin (g/ dL)	12.21 ± 2.38	12.47 ± 2.24	0.61	12.20 ± 2.27	12.17 ± 1.98	0.72					
GGTP (U/L)	47.43 ± 24.88	261.38 ± 238.38	< 0.0001	52.47 ± 25.32	283.64 ± 168.33	<0.0001					
ALKP (U/L)	108.11 ± 59.50	262.95 ± 257.67	< 0.0001	149.96 ± 99.87	317.99 ± 254.41	<0.0001					
Total Bilirubin (mg/dL)	1.91 ± 2.93	2.53 ± 2.99	0.008	2.23 ± 3.06	3.02 ± 4.59	0.07					
Albumin (g/dL)	3.17 ± 0.80	2.90 ± 0.71	0.01	3.16 ± 0.86	2.81 ± 0.58	0.004					
AFP (IU/mL)	16.61 ± 18.41	19.85 ± 25.08	0.50	15.95 ± 22.26	26.58 ± 28.69	0.003					
MTD (cm)	2.96 ± 1.03	2.94 ± 1.02	0.80	8.01 ± 3.15	9.15 ± 3.79	0.03					
Nodule number (%)			0.36 ^			0.12 ^					
1	76 (60.32)	42 (53.85)		64 (62.14)	39 (50.65)						
2-3	50 (39.68)	36 (46.15)		39 (37.86)	38 (49.35)						
>3	0 (0.00)	0 (0.00)		0 (0.00)	0 (0.00)						
PV Thrombosis (%)	14 (11.20)	12 (15.58)	0.37 ^	27 (27.00)	33 (45.21)	0.01 ^					
Cirrhosis (%)	103 (82.40)	61 (78.21)	0.46 ^	79 (75.96)	59 (77.63)	0.79 ^					

* All values: Means±Standard Deviation as continuous; Frequences and Percentage (%) as categorical. ψ Wilcoxon rank-sum (Mann-Whitney) test; ^ Chi-square test.

Abbreviations: GGTP, gamma glutamyl transpeptidase; ALKP, Alkaline phosphatase; AFP, Alphafetoprotein; MTD, Maximum Tumor Diameter; PVT, Portal Vein Thrombosis; CRP, C-Reactive Protein.

that albumin appeared to have a significant protective effect on MTD in larger tumors, and a lesser protective effect on large tumor PVT and multifocality. Dichotomization according to blood GGTP values (**TABLE 6**) also showed a significant effect for MTD and PVT, but only in larger (≥ 5 cm) MTD patients. In this subgroup of larger tumors with higher GGTP levels, there were also significantly higher platelet counts and ALKP levels, and significantly lower albumin levels. A similar dichotomization using ALKP values however, showed no difference in MTD, PVT or multifocality (data not shown).

Discussion

There are several noteworthy findings from this analysis. Amongst them are the high (42% or 58%, depending on AFP cutoff) of negative or low AFP HCC patients (**TABLE 1**). This has been found in several different reports [15-21]. It is thus unsurprising that AFP has been a poor screening tool, especially for small size HCCs. In addition, we found that large HCCs also occurred in absent or low range AFP patients (**TABLE 2**). Thus, factors other than AFP, such as cell growth and inflammatory cytokines must be postulated to be important in the growth of large size HCCs. Another finding was the high percent (19-50%) of patients with PVT, even in the lowest AFP group (19%). As AFP level group and thus MTD increased, so did GGTP mean values and to a lesser extent, ALKP. GGTP has been found to be a useful tumor marker by others, especially in low AFP HCCs [21,29,30-33] and so, to a lesser extent has ALKP [34]. Platelets were found to be associated with an increase in MTD, especially for the larger size tumors, but there was an association between MTD and platelet numbers (TABLE 4 and FIGURE 2), as noted elsewhere [27,28] and there was a greater percent of PVT in the larger tumors (TABLES 2, 4 and FIGURE 1). The reason for this is unclear. Whether it is that tumors beyond a certain diameter have a larger number of invasive (? stem) cells, or whether the growth and inflammatory factors that produce larger tumors also produce more PVT, has not been closely explored, to our knowledge. However, consistent with these ideas, lower albumin, which is associated with increased systemic inflammation in the Glasgow Index [35,36], is also associated with larger MTD and increased PVT (TABLE 5), although with statistical significance only for the larger tumors.

Furthermore, a weakness in this study is that survival data was not available from this newlyformed collaborative group.

Various markers have been suggested for low- or normal-AFP HCCs. These include, but are not limited to: DCP [37,38], Glypican-3 [39], fucosylated serum paraoxonase 1 [40] and alpha-1 acid glycoprotein [41], TEMs [42], Dickkopf-1 [43] and EpCam stained circulating tumor cells [44] and staining of HCCs [45], C-reactive protein [46], alpha-L-fucosidase [47] and hepatic miRs [48]. With such a high percent of small and large HCCs having low AFP levels, novel biomarkers are clearly needed. Much effort has recently focused on combinations of markers, such as AFP plus AFP-L3 plus DCP for use in surveillance [49]. Even in patients with very low AFP (<20 ng/ml), sensitivity was over 65% using these 3 marker combinations [50,51]. However, the different clinical HCC phenotypes for these marker combinations has not been explored in detail, except for the association of increased incidence of PVT in patients with high DCP levels [52].

Disclosure statement

The authors declare no conflict of interest. All authors have read and agree with this paper.

This work is supported in part by NIH grant CA 82723 (B.I.C).

REFERENCES

Abelev GI, Perova SD, Khramkova NI, Postnikova ZA, Irlin IS. Production of embryonic alpha-globulin by the transplantable mouse hepatomas. *Transplantation.* 1, 174-180 (1963).

Tatarinov YS. Detection of embryospecific alpha-globulin in the blood sera of patients with primary liver tumour. *Vop. Med. Khim.* 10, 90-91 (1964).

Yang SL, Liu LP, Yang S, et al. Preoperative serum α -fetoprotein and prognosis after hepatectomy for hepatocellular carcinoma. *Br. J. Surg.* 103, 716-724 (2016).

Blank S, Wang Q, Fiel MI, et al. Assessing prognostic significance of preoperative alpha-fetoprotein in hepatitis B-associated hepatocellular carcinoma: normal is not the new normal. *Ann. Surg. Oncol.* 21, 986-994 (2014).

Meguro M, Mizuguchi T, Nishidate T, et al. Prognostic roles of preoperative α -fetoprotein and des- γ carboxy prothrombin in hepatocellular carcinoma patients. *World J. Gastroenterol.* 21, 4933-4945 (2015).

Nomura F, Ohnishi K, Tanabe Y. Clinical features and prognosis of hepatocellular carcinoma with reference to serum alpha-fetoprotein levels. Analysis of 606 patients. *Cancer.* 64, 1700-1707 (1989).

Kudo A, Matsumura S, Ban D, Irie T, et al. Does the preoperative alphafetoprotein predict the recurrence and mortality after hepatectomy for hepatocellular carcinoma without macrovascular invasion in patients with normal liver function? *Hepatol. Res.* 44, E437-E446 (2014).

Liu C, Xiao GQ, Yan LN, et al. Value of α-fetoprotein in association with clinicopathological features of hepatocellular carcinoma. *World J. Gastroenterol.* 19, 1811-1819 (2013).

An SL, Xiao T, Wang LM, et al. Prognostic Significance of

Preoperative Serum Alpha- fetoprotein in Hepatocellular Carcinoma and Correlation with Clinicopathological Factors: a Single-center Experience from China. *Asian Pac. J. Cancer Prev.* 16, 4421-4427 (2015).

El-Serag HB, Kanwal F, Davila JA, Kramer J, Richardson P. A new laboratory-based algorithm to predict development of hepatocellular carcinoma in patients with hepatitis C and cirrhosis. *Gastroenterol.* 146, 1249-1255 (2014).

Omata M, Cheng AL, Kokudo N, et al. Asia-Pacific clinical practice guidelines on the management of hepatocellular carcinoma: a 2017 update. *Hepatol Int.* 11, 317-370 (2017).

Memon K, Kulik L, Lewandowski RJ, et al. Alpha-fetoprotein response correlates with EASL response and survival in solitary hepatocellular carcinoma treated with transarterial therapies: a subgroup analysis. *J. Hepatol.* 56, 1112-1120 (2012).

Worland T, Harrison B, Delmenico L, Dowling D. Hepatocellular Carcinoma Screening Utilising Serum Alpha-Fetoprotein Measurement and Abdominal Ultrasound Is More Effective than Ultrasound Alone in Patients with Non-viral Cirrhosis. *J. Gastrointest. Cancer.* (2017).

Yang JD, Dai J, Singal AG, Gopal P, et al. Improved Performance of Serum Alpha-Fetoprotein for Hepatocellular Carcinoma Diagnosis in HCV Cirrhosis with Normal Alanine Transaminase. *Cancer Epidemiol. Biomarkers Prev.* 26, 1085-1092 (2017).

Lok AS, Sterling RK, Everhart JE, Wright EC, et al. HALT-C Trial Group. Des-gamma-carboxy prothrombin and alpha-fetoprotein as biomarkers for the early detection of hepatocellular carcinoma. *Gastroenterol.* 138, 493-502 (2010).

Farinati F, Marino D, De Giorgio M, et al. Diagnostic and prognostic role

of alpha-fetoprotein in hepatocellular carcinoma: both or neither? *Am. J. Gastroenterol.* 101, 524-532 (2006).

Hamamura K, Shiratori Y, Shiina S, et al. Unique clinical characteristics of patients with hepatocellular carcinoma who present with high plasma desgamma-carboxy prothrombin and low serum alpha-fetoprotein. *Cancer.* 88, 1557-1564 (2000).

Furui J, Furukawa M, Kanematsu T. The low positive rate of serum alpha-fetoprotein levels in hepatitis C virus antibody-positive patients with hepatocellular carcinoma. *Hepatogastroenterol.* 42, 445-449 (1995).

Nomura F, Ohnishi K, Tanabe Y. Clinical features and prognosis of hepatocellular carcinoma with reference to serum alpha-fetoprotein levels. Analysis of 606 patients. *Cancer.* 64, 1700-1707 (1989).

Carr BI, Guerra V. Low Alpha-Fetoprotein Levels Are Associated with Improved Survival in Hepatocellular Carcinoma Patients with Portal Vein Thrombosis. *Dig. Dis. Sci.* 61, 937-947 (2016).

Carr BI, Guerra V, Giannini EG, Farinati F, et al. Low alpha-fetoprotein HCC and the role of GGTP. *Int. J. Biol. Markers.* 29, e395-e402 (2014).

Hu J, Xu Y, Shen ZZ, et al. High expressions of vascular endothelial growth factor and platelet-derived endothelial cell growth factor predict poor prognosis in alpha-fetoproteinnegative hepatocellular carcinoma patients after curative resection. *J. Cancer Res. Clin. Oncol.* 135, 1359-1367 (2009).

Sawabu N, Wakabayashi T, Ozaki K, et al. Serum tumor markers in patients with hepatocellular carcinoma: diagnosis of alpha-fetoprotein -low or -negative patients. *Gastroenterol. Jpn.* 20, 209-215 (1985).

Sangiovanni A, Del Ninno E, Fasani P, et al. Increased survival of cirrhotic patients with a hepatocellular carcinoma detected during surveillance. *Gastroenterol.* 126, 1005-1014 (2004).

Giannini EG, Marenco S, Borgonovo G, Savarino V. Italian Liver Cancer (ITA. LI.CA) group. Alpha-fetoprotein has no prognostic role in small hepatocellular carcinoma identified during surveillance in compensated cirrhosis. *Hepatol.* 56, 1371-1379 (2012).

Zhang XF, Qi X, Meng B, et al. Prognosis evaluation in alpha-fetoprotein negative hepatocellular carcinoma after hepatectomy: comparison of five staging systems. *Eur. J. Surg. Oncol.* 36, 718-724 (2010).

Carr BI, Guerra V Hepatocellular carcinoma size: platelets, γ -glutamyl transpeptidase, and alkaline phosphatase. *Oncol.* 85, 153-159 (2013).

Carr BI, Guerra V, Giannini EG, et al. Significance of platelet and AFP levels and liver function parameters for HCC size and survival. *Int. J. Biol. Markers.* 29, e215-e223 (2014).

Carr BI, Pancoska P, Branch RA. Significance of increased serum GGTP levels in HCC patients. *Hepatogastroenterol.* 57, 869-874 (2010).

Izumi M. Isolation, characterization and clinical evaluation of the gammaglutamyltransferase associated with hepatocellular carcinoma. *Acta Med. Okayama.* 39, 19-33 (1985).

Xu K, Meng XY, Wu JW, et al. Diagnostic value of serum gammaglutamyl transferase isoenzyme for hepatocellular carcinoma: a 10-year study. *Am. J. Gastroenterol.* 87, 991-995 (1992).

Yao DF, Huang ZW, Chen SZ, et al. Diagnosis of hepatocellular carcinoma by quantitative detection of hepatomaspecific bands of serum gammaglutamyltransferase. *Am. J. Clin. Pathol.* 110, 743-749 (1998).

Yao D, Jiang D, Huang Z, Lu J, Tao Q, Yu Z, Meng X. Abnormal expression

of hepatoma specific gamma-glutamyl transferase and alteration of gamma-glutamyl transferase gene methylation status in patients with hepatocellular carcinoma. *Cancer.* 88, 761-769 (2000).

Yu MC, Chan KM, Lee CF, Lee YS, et al. Alkaline phosphatase: does it have a role in predicting hepatocellular carcinoma recurrence? *J. Gastrointest. Surg.*15, 1440-1449 (2011).

Kinoshita A, Onoda H, Imai N, Iwaku A, et al. Comparison of the prognostic value of inflammationbased prognostic scores in patients with hepatocellular carcinoma. *Br. J. Cancer.* 107, 988-993 (2012).

Pinato DJ, Stebbing J, Ishizuka M, Khan SA, et al. A novel and validated prognostic index in hepatocellular carcinoma: the inflammation based index(IBI). *J. Hepatol.* 57(5), 1013-1020 (2012).

Fujiyama S, Morishita T, Hashiguchi O, Sato T. Plasma abnormal prothrombin (des-gamma-carboxy prothrombin) as a marker of hepatocellular carcinoma. *Cancer.* 61, 1621-1628 (1988).

Ji J, Wang H, Li Y, Zheng L. Diagnostic Evaluation of Des-Gamma-Carboxy Prothrombin versus α -Fetoprotein for Hepatitis B Virus-Related Hepatocellular Carcinoma in China: A Large-Scale, Multicentre Study. *PLoS One.* 11(4), e0153227 (2016).

Li B, Liu H, Shang HW, Li P, Li N, Ding HG. Diagnostic value of glypican-3 in alpha fetoprotein negative hepatocellular carcinoma patients. *Afr. Health Sci.* 13, 703-709 (2013).

Shu H, Li W, Shang S, Qin X, et al. Diagnosis of AFP-negative earlystage hepatocellular carcinoma using Fuc-PON1. *Discov. Med.* 23, 163-168 (2017).

Bachtiar I, Kheng V, Wibowo GA, Gani RA. Alpha-1-acid glycoprotein as potential biomarker for alphafetoprotein-low hepatocellular carcinoma. BMC Res. Notes. 3, 319 (2010).

Mao L, Wang Y, Wang D, et al. TEMs but not DKK1 could serve as complementary biomarkers for AFP in diagnosing AFP-negative hepatocellular carcinoma. *PLoS One.* 12(9), e0183880 (2017).

Yang H, Chen GD, Fang F, et al. Dickkopf-1: as a diagnostic and prognostic serum marker for early hepatocellular carcinoma. *Int. J. Biol. Markers.* 28, 286-297 (2013).

Sun YF, Xu Y, Yang XR, et al. Circulating stem cell-like epithelial cell adhesion molecule-positive tumor cells indicate poor prognosis of hepatocellular carcinoma after curative resection. *Hepatol.* 57, 1458-1468 (2013).

Yamashita T, Forgues M, Wang W, et al. EpCAM and alpha-fetoprotein expression defines novel prognostic subtypes of hepatocellular carcinoma. *Cancer Res.* 68, 1451-1461 (2008).

She S, Xiang Y, Yang M, et al. C-reactive protein is a biomarker of AFPnegative HBV-related hepatocellular carcinoma. *Int. J. Oncol.* 47, 543-554 (2015).

Gan Y, Liang Q, Song X. Diagnostic value of alpha-L-fucosidase for hepatocellular carcinoma: a metaanalysis. *Tumour Biol.* 35, 3953-3960 (2014).

Ghosh A, Ghosh A, Datta S, et al. Hepatic miR-126 is a potential plasma biomarker for detection of hepatitis B virus infected hepatocellular carcinoma. *Int. J. Cancer.* 138, 2732-2744 (2016).

Johnson P, Pirrie SJ, Cox TF, et al. The Detection of Hepatocellular Carcinoma Using a Prospectively Developed and Validated Model Based on Serological Biomarkers. *Cancer Epidemiol. Biomarkers Prev.* 23, 144-153 (2014).

Best J, Bilgi H, Heider D, et al. The GALAD scoring algorithm based on

AFP, AFP-L3, and DCP significantly improves detection of BCLC early stage hepatocellular carcinoma. *Z. Gastroenterol.* 54, 1296-1305 (2016).

Choi JY, Jung SW, Kim HY, et al. Diagnostic value of AFP-L3 and PIVKA-

II in hepatocellular carcinoma according to total-AFP. *World J. Gastroenterol.* 19, 339-346 (2013).

Koike Y, Shiratori Y, Sato S, et al. Des-gamma-carboxy prothrombin as a useful predisposing factor for the development of portal venous invasion in patients with hepatocellular carcinoma: a prospective analysis of 227 patients. *Cancer.* 91, 561-569 (2001).