Clinical applications of IFN-α blockade in systemic lupus erythematosus

"...there is ample evidence that IFN- α is overproduced by many patients with active SLE ... and may have direct effects..."

The immune system relies on a wide variety of cytokines for controlling infections, and also for dampening the inflammatory reaction in order to avoid excessive tissue damage. Among these cytokines, those of the type I interferon (IFN) family are essential mediators of antivirus defense [1]. In fact, the type I IFNs were detected as a result of their protective effects against established viruses infecting more cells. Thus, for many years, IFN- α 2a and -2b have been successfully used in the therapy of chronic viral hepatitis [2]. In addition, its blockade of cell proliferation, proapoptotic effects [3] and anti-angiogenic properties have led to the use of IFN-a in melanoma patients [4]. However, under such therapy, autoimmunity is not uncommon [5,6] and viral infections associated with IFN release can lead to flares of autoimmune diseases [7]. This involvement of IFN- α in immuno-inflammatory events makes it an interesting target in rheumatology.

All type I IFNs use the same receptor, consisting of the IFN-aR1 and -R2 chains [8,9], which bind to the Janus kinases, Jak1 and Tyk2, respectively, for transducing their signals [10,11]. By contrast, IFN-y, the single type II IFN, employs Jak1 and Jak2 for its signal transduction. While the pathway of IFN-γ ends in the phosphorylation of serine and tyrosine and subsequent nuclear localization of dimers of signal transducer and activator of transcription (STAT) 1, IFN- α mainly induces formation of a heterotrimer of STAT1 and STAT2 with IFN regulatory factor-9 [12]. In addition, IFN- α induces the phosphorylation of STAT4 dimers, which play an important role in enabling IFN- γ production [13,14], and leads to the presence of phosphorylated STAT1 dimers in macrophages [15]. These transcription factors directly lead to the various effects exerted by IFNs.

There is significant evidence that some of these effects are problematic in systemic lupus erythematosus (SLE). The therapeutic application of type I IFN, but also IFN-y, may provoke drug-induced lupus [16,17]. Moreover, mouse model data suggest an influence of type I IFNs on lupus [18], even though IFN- γ appears to be predominant in the mouse [19]. How could this happen? There are several pieces of evidence that may help us to understand these effects. First, IFNs lead to cell death, both by preparing them for apoptosis and by stimulating killer cells. In SLE, dead cells may not be sufficiently removed [20], thus exposing the body to an overload of autoantigens [21]. Second, IFNs activate dendritic cells, which can then effectively present these autoantigens [22]. Third, IFNs may also keep autoimmune T cells alive and promote antibody production by B cells.

In contrast with their proposed role in lupus pathogenesis, type I IFNs are normally produced when cells become infected with a virus and they contribute to an effective host response. Intracellular viral RNA binds intracellular RIG I-like helicases, activating IFN regulatory factors and NF- κ B [23]. Alternatively, Toll-like receptors of uninfected dendritic cells, which detect viral RNA or DNA in their endolysosomes, induce the same signaling events [24] that lead to potent type I IFN induction [25]. Indeed, in the peripheral blood, plasmacytoid dendritic cells are the main producers of IFN- α [26].

In SLE there is an abundance of immune complexes containing DNA and/or RNA. Like viruses, these immune complexes can trigger Toll-like receptors, inducing IFN- α production as well as the production of other cytokines such as TNF [27]. Along this line, there is ample evidence that IFN- α is overproduced by many patients with active SLE [28–30] and may have direct effects, as evidenced by changes in mRNA patterns [31-33] and by increased expression of STAT1, which itself is an IFN-dependent gene [34]. Thus, type I IFNs likely constitute a positive feedback loop in SLE pathogenesis,



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wherein viral infection and immune complexes induce IFNs, the effects of which increase the amount of available autoantigen, foster the production of affinity-maturated autoantibodies and make IFN signaling even more effective.

Accordingly, one would expect to see beneficial effects of therapeutic interference with this feedback loop by means of blocking the type I IFN receptor or (presumably several) type I IFNs. Indeed, such an approach has been used in patients with SLE [35,36]. In a first safety trial, the anti-IFN- α monoclonal antibody MEDI-545 dampened the IFN signature in a dose-dependent way, not only in peripheral blood but also in skin samples [36]. Moreover, other proinflammatory molecules, such as TNF, were likewise down-modulated, which is consistent with the idea of interrupting a feedback loop. Compared with placebo, this approach appeared to reduce both overall SLE activity, as measured by the SLE disease activity index, and the number of SLE flares [35]. Most importantly, no obvious safety signals were reported [35].

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Therefore, IFN- α blockade is being tested in controlled clinical trials in SLE [101], and given the similar lines of evidence, it will also be tested in polymyositis/dermatomyositis [37–39]. Such trials will ultimately prove or disprove this concept. However, for the time being, there is some room for educated guessing: given that subsets of patients display the classical IFN signature, one would expect these patients to be more likely to respond to IFN receptor blockade. In view of the association of type I IFN levels with disease activity, one might also argue that a type I IFN blockade would probably diminish activity rather than cure the disease. Under these circumstances, type I IFN receptor blockade may be an interesting option, along with other agents, in treating severe lupus flares or in maintaining remission following flares.

On the other hand, safety will have to be monitored very carefully in longer term therapy and with particular vigilance with regard to virus infections and even tumor surveillance. While blockade of both the type I and type II IFN systems would likely show dramatic effects in this regard [40,41], incomplete blockade of type I IFN is probably more benign. Nevertheless, both severe acute and chronic virus infections could cause problems, and it remains to be seen how much IFN blockade is feasible from this perspective.

In conclusion, the type I IFN system is a plausible target for therapeutic blockade for the treatment of severe SLE, and probably for autoimmune myositides treatment as well. Clinical trial data are still very limited, but there is hope that this concept will realize success as a feasible and effective therapy for some patients with a severe SLE flare or ongoing moderate activity associated with evidence of IFN pathway activation. Even after efficacy and safety have been secured, there is a need for a more profound understanding of the exact pathophysiological processes that involve IFNs.

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Bibliography

Papers of special note have been highlighted as: • of interest

- of considerable interest
- Stetson DB, Medzhitov R: Type I interferons in host defense. *Immunity* 25, 373–381 (2006).
- Very good review on the role of type I interferons (IFNs) in host defence.
- 2 Davis GL: Recombinant α-interferon treatment of non-A, and non-B (type C) hepatitis: review of studies and

recommendations for treatment. J. Hepatol. 11(Suppl. 1), S72–S77 (1990).

- 3 Chawla-Sarkar M, Lindner DJ, Liu YF et al.: Apoptosis and interferons: role of interferon-stimulated genes as mediators of apoptosis. Apoptosis 8, 237–249 (2003).
- 4 Kirkwood JM, Ernstoff MS: Role of interferons in the therapy of melanoma. J. Invest. Dermatol. 95, S180–S184 (1990).
- 5 Raanani P, Ben-Bassat I: Immune-mediated complications during interferon therapy in hematological patients. *Acta Haematol.* 107, 133–144 (2002).
- 6 Wilson LE, Widman D, Dikman SH, Gorevic PD: Autoimmune disease complicating antiviral therapy for hepatitis C virus infection. *Semin. Arthritis Rheum.* 32, 163–173 (2002).
- 7 Zandman-Goddard G, Shoenfeld Y: Infections and SLE. *Autoimmunity* 38, 473–485 (2005).
- Novick D, Cohen B, Rubinstein M: The human interferon α/β receptor: characterization and molecular cloning. *Cell* 77, 391–400 (1994).

- 9 Lutfalla G, Holland SJ, Cinato E et al.: Mutant U5A cells are complemented by an interferon-α β receptor subunit generated by alternative processing of a new member of a cytokine receptor gene cluster. EMBO J. 14, 5100–5108 (1995).
- 10 Rodig SJ, Meraz MA, White JM *et al.*: Disruption of the *Jak1* gene demonstrates obligatory and nonredundant roles of the Jaks in cytokine-induced biologic responses. *Cell* 93, 373–383 (1998).
- 11 Karaghiosoff M, Neubauer H, Lassnig C et al.: Partial impairment of cytokine responses in Tyk2-deficient mice. *Immunity* 13, 549–560 (2000).
- 12 Darnell JE Jr, Kerr IM, Stark GR: Jak-STAT pathways and transcriptional activation in response to IFNs and other extracellular signaling proteins. *Science* 264, 1415–1421 (1994).
- 13 Nguyen KB, Watford WT, Salomon R *et al.*: Critical role for STAT4 activation by type 1 interferons in the interferon-γ response to viral infection. *Science* 297, 2063–2066 (2002).
- 14 Longman RS, Braun D, Pellegrini S, Rice CM, Darnell RB, Albert ML: Dendritic-cell maturation alters intracellular signaling networks, enabling differential effects of IFN-α/β on antigen crosspresentation. *Blood* 109, 1113–1122 (2007).
- 15 Park C, Li S, Cha E, Schindler C: Immune response in Stat2 knockout mice. *Immunity* 13, 795–804 (2000).
- 16 Ioannou Y, Isenberg DA: Current evidence for the induction of autoimmune rheumatic manifestations by cytokine therapy. *Arthritis Rheum.* 43, 1431–1442 (2000).
- Comprehensive review on cytokine and cytokine-directed therapy causing autoimmune phenotypes.
- 17 Graninger WB, Hassfeld W, Pesau BB, Machold KP, Zielinski CC, Smolen JS: Induction of systemic lupus erythematosus by interferon-γ in a patient with rheumatoid arthritis. J. Rheumatol. 18, 1621–1622 (1991).
- 18 Santiago-Raber ML, Baccala R, Haraldsson KM *et al.*: Type-I interferon receptor deficiency reduces lupus-like disease in NZB mice. *J. Exp. Med.* 197, 777–788 (2003).
- 19 Theofilopoulos AN, Koundouris S, Kono DH, Lawson BR: The role of IFN-γ in systemic lupus erythematosus: a challenge to the Th1/Th2 paradigm in autoimmunity. *Arthritis Res.* 3, 136–141 (2001).
- 20 Herrmann M, Voll RE, Zoller OM, Hagenhofer M, Ponner BB, Kalden JR: Impaired phagocytosis of apoptotic cell material by monocyte-derived macrophages

from patients with systemic lupus erythematosus. *Arthritis Rheum.* 41, 1241–1250 (1998).

- 21 Casciola-Rosen L, Rosen A, Petri M, Schlissel M: Surface blebs on apoptotic cells are sites of enhanced procoagulant activity: implications for coagulation events and antigenic spread in systemic lupus erythematosus. *Proc. Natl Acad. Sci. USA* 93, 1624–1629 (1996).
- 22 Banchereau J, Pascual V: Type I interferon in systemic lupus erythematosus and other autoimmune diseases. *Immunity* 25, 383–392 (2006).
- Substantial review on IFN-α in systemic lupus erythematosus (SLE), but misinterprets the role of TNF in our opinion.
- 23 Takeuchi O, Akira S: MDA5/RIG-I and virus recognition. *Curr. Opin. Immunol.* 20, 17–22 (2008).
- 24 Kawai T, Akira S: Toll-like receptor and RIG-I-like receptor signaling. Ann. NY Acad. Sci. 1143, 1–20 (2008).
- 25 Honda K, Takaoka A, Taniguchi T: Type I interferon [corrected] gene induction by the interferon regulatory factor family of transcription factors. *Immunity* 25, 349–360 (2006).
- 26 Siegal FP, Kadowaki N, Shodell M et al.: The nature of the principal type 1 interferon-producing cells in human blood. *Science* 284, 1835–1837 (1999).
- Background paper on IFN and plasmacytoid dendritic cells.
- 27 Aringer M, Crow MK: A bridge between interferon-α and tumor necrosis factor in lupus. J. Rheumatol. 35, 1473–1476 (2008).
- 28 Hooks JJ, Moutsopoulos HM, Geis SA, Stahl NI, Decker JL, Notkins AL: Immune interferon in the circulation of patients with autoimmune disease. *N. Engl. J. Med.* 301, 5–8 (1979).
- Classical paper on IFN in SLE.
- 29 Ytterberg SR, Schnitzer TJ: Serum interferon levels in patients with systemic lupus erythematosus. *Arthritis Rheum.* 25, 401–406 (1982).
- 30 Dall'era MC, Cardarelli PM, Preston BT, Witte A, Davis JC: Type I interferon correlates with clinical and serologic manifestations of systemic lupus erythematosus. Ann. Rheum. Dis. 64(12), 1692–1697 (2005).
- 31 Bennett L, Palucka AK, Arce E *et al.*: Interferon and granulopoiesis signatures in systemic lupus erythematosus blood. *J. Exp. Med.* 197, 711–723 (2003).

- 32 Kirou KA, Lee C, George S et al.: Coordinate overexpression of interferon-α-induced genes in systemic lupus erythematosus. Arthritis Rheum. 50, 3958–3967 (2004).
- 33 Baechler EC, Batliwalla FM, Karypis G et al.: Interferon-inducible gene expression signature in peripheral blood cells of patients with severe lupus. Proc. Natl Acad. Sci. USA 100, 2610–2615 (2003).
- 34 Karonitsch T, Feierl E, Steiner CW et al.: Activation of the interferon-γ signaling pathway in systemic lupus erythematosus peripheral blood mononuclear cells. Arthritis Rheum. 60, 1463–1471 (2009).
- 35 Wallace DJ, Petri M, Olsen N *et al.*: MEDI-545, an anti-interferon α monoclonal antibody, shows evidence of clinical activity in systemic lupus erythematosus. *Arthritis Rheum.* 56(9 Suppl.), S526–S527 (2007) (Abstract).
- 36 Yao Y, Richman L, Higgs BW et al.: Neutralization of interferon-α/β-inducible genes and downstream effect in a Phase I trial of an anti-interferon-α monoclonal antibody in systemic lupus erythematosus. Arthritis Rheum. 60, 1785–1796 (2009).
- First paper on an IFN-α blockade therapeutic trial in SLE.
- 37 Somani AK, Swick AR, Cooper KD, McCormick TS: Severe dermatomyositis triggered by interferon β-1a therapy and associated with enhanced type I interferon signaling. *Arch. Dermatol.* 144, 1341–1349 (2008).
- 38 Baechler EC, Bauer JW, Slattery CA *et al.*: An interferon signature in the peripheral blood of dermatomyositis patients is associated with disease activity. *Mol. Med.* 13, 59–68 (2007).
- 39 Niewold TB, Kariuki SN, Morgan GA, Shrestha S, Pachman LM: Elevated serum interferon-α activity in juvenile dermatomyositis: associations with disease activity at diagnosis and after thirty-six months of therapy. *Arthritis Rheum.* 60, 1815–1824 (2009).
- 40 Meraz MA, White JM, Sheehan KC et al.: Targeted disruption of the Stat1 gene in mice reveals unexpected physiologic specificity in the JAK-STAT signaling pathway. *Cell* 84, 431–442 (1996).
- 41 Durbin JE, Hackenmiller R, Simon MC, Levy DE: Targeted disruption of the mouse *Stat1* gene results in compromised innate immunity to viral disease. *Cell* 84, 443–450 (1996).

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101 ClinicalTrials.gov http://clinicaltrials.gov