

Arachidonic acid-derived eicosanoids in rheumatoid arthritis: implications and future targets

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Rheumatoid arthritis (RA) is an autoimmune articular disease associated with chronic inflammation of the joints. RA is characterized by excessive synovial proliferation, progressive joint destruction and immobility. Cell types including neutrophils, macrophages, T and B cells infiltrate the inflamed joint area and release excessive amounts of proinflammatory mediators, resulting in the persistence of inflammation and resultant synovial tissue destruction. The most prominent mediators known to play a pivotal role in initiation and progression of articular inflammation during RA include proinflammatory cytokines, such as tumor necrosis factor- α and interleukin-1, reactive oxygen radicals, growth factors and lipid mediators generated via arachidonic acid (AA) metabolism. A diverse range of lipid mediators generated via AA biosynthetic metabolism, and often referred to as eicosanoids, mediate a wide variety of physiological and pathological functions. Eicosanoids play a central role in the progression and chronic inflammation associated with RA by mediating proinflammatory actions. However, recent studies have also shown that some eicosanoids generated during cell-cell interactions may act as stop signals for inflammation and help in the resolution of inflammation. Current therapies to treat the symptoms of RA are also directed towards inhibition of key enzymes and mediators within the AA biosynthetic pathway. This review outlines the broad understanding of AA-derived eicosanoid mediators in relation to RA. Furthermore, potential future therapeutic targets within the AA metabolic pathway for the treatment of RA will be discussed.

Arachidonic acid metabolism in rheumatoid arthritis

Arachidonic acid (AA) metabolites, including prostaglandins (PGs), thromboxanes (TXs), leukotrienes (LTs), hydroxyeicosatetraenoic acids (HETEs) and lipoxins (LXs), are essential mediators involved in the pathophysiology of rheumatoid arthritis (RA). These eicosanoids, so named for the 20 carbons contained in AA, exert a broad range of pro- and/or anti-inflammatory functions at the site of RA. The first step of AA metabolism begins with the release of free arachidonate from cell membrane phospholipids by phospholipase (PL) A_2 . Subsequently, a diverse range of metabolites are generated downstream of PLA $_2$ by multiple biosynthetic pathways, including cyclooxygenase (COX), lipoxygenase (LOX) and further sequential-specific terminal enzymes that are expressed in a cell-type and tissue-specific manner (Figure 1).

Cyclooxygenase

COX is one of the major enzymes within the AA metabolic pathway and has been investigated extensively in the field of RA research. COX converts AA to PGH $_2$ which is the common substrate for physiologically active PGs and TXs, including PGE $_2$, PGD $_2$, PGF $_{2\alpha}$ and PGI $_2$

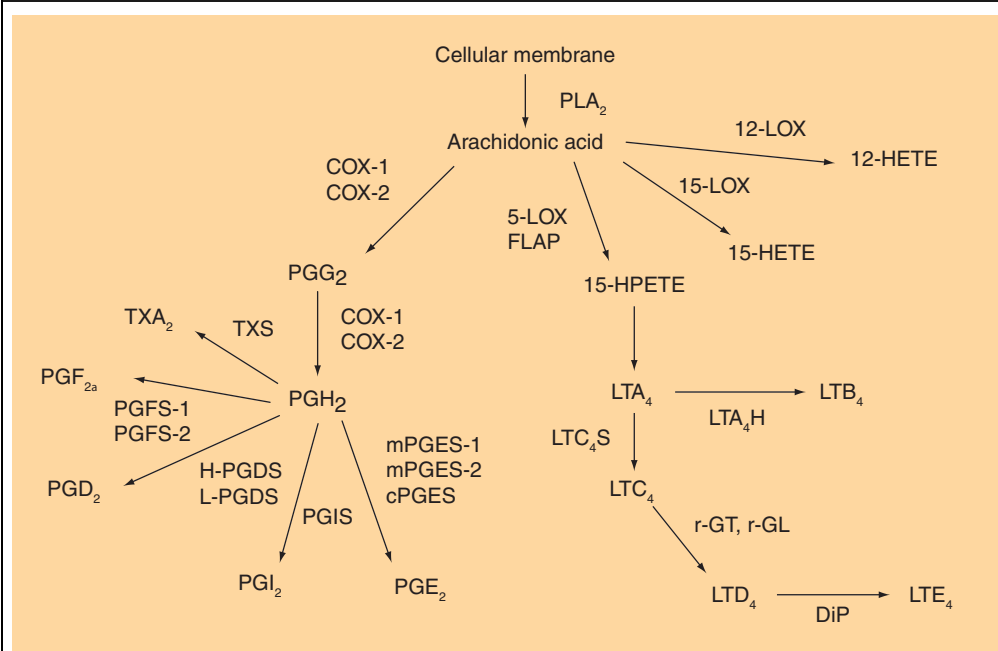
(prostacyclin) and TXA $_2$. COX exists in at least two isoforms, constitutive COX-1 and inducible COX-2 [1,2]. Most recently, a splice variant of COX-1 mRNA, retaining intron 1 and referred to by several names, including COX-3, COX-1b or COX-1v, has been described [3,4]. However, the existence and role of these remain questionable. In the synovial tissue of RA, expression of COX-2, but not COX-1, is upregulated in infiltrating mononuclear inflammatory cells, synovial fibroblasts and vascular endothelial cells in synovial sublining layers [5,6]. In addition, induction of COX-2 is observed in response to stimulation by proinflammatory cytokines such as interleukin (IL)-1 and tumor necrosis factor (TNF)- α in cultured synovial fibroblasts from patients with RA [7–9]. These observations regarding COX-2 induction in articular tissues seem to be closely associated with the clinical efficacy of COX-2-selective, as well as nonselective, inhibitors in the treatment of RA.

Prostaglandins

Among the PGs, PGE $_2$ is one of the most prominent proinflammatory PGs associated with rheumatoid synovitis, and high concentrations of PGE $_2$ are detectable in the synovial fluid of

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future
medicine

Figure 1. Biosynthetic pathways in arachidonic acid metabolism.

COX: Cyclooxygenase; cPGES: Cytosolic PGE synthase; DiP: Dipeptidase; FLAP: 5-LOX activating protein; HETE: Hydroxyeicosatetraenoic acid; HPETE: Hydroperoxyeicosatetraenoic acid; H-PGDS: Hematopoietic PGDS; LOX: Lipoxygenase; LT: Leukotriene; LTA₄H: LTA₄ hydrolase; LTC₄S: LTC₄ synthase; L-PGDS: Lipocalin PGD synthase; mPGES: Microsomal PGES; PG: Prostaglandin; PGDS: PGD synthase; PGFS: PGF synthase; PGES: PGE synthase; PGIS: PGI synthase; PLA₂: Phospholipase A₂; r-GT: r-glutamyl leukotrienase; r-GT: r-glutamyl transpeptidase; TX: Thromboxane; TXS: Thromboxane synthase.

patients with RA [10]. At present, inhibition of PGE₂ production and action is most closely associated with a reduction in the pain and inflammation associated with RA. Ligand-activated cells, such as synovial fibroblasts and monocytes/macrophages, in synovial tissue are likely to be a primary source of PGE₂ in the joints of patients with RA [8,11,12]. Overproduction of PGE₂ in response to proinflammatory cytokine stimuli coincides with the upregulation of COX-2 expression at sites of inflammation [8].

PGE₂ exerts most of its actions through a family of G protein-coupled receptors (GPCRs) including EP1, 2, 3 and 4. It has been reported that RA synovial fibroblasts express EP2 and 4 receptors constitutively and that PGE₂ regulates the production of cytokines and growth factors, such as parathyroid hormone-related peptide, IL-6, macrophage colony-stimulating factor (M-CSF) and vascular endothelial growth factor (VEGF), through the activation of EP2 and 4 in IL-1-stimulated synovial fibroblasts [13]. PGE₂ is also a modulator of matrix metalloproteinases (MMPs) and tissue inhibitors of MMPs (TIMPs) [14]. Furthermore, EP4 receptor-null

mice, but not EP1, 2 and 3 receptor-null mice, have been shown to have reduced incidence and severity of collagen-induced arthritis (CIA) [15]. These reports suggest that modulating EP receptor signaling is a possible therapeutic strategy for treatment of RA.

PG terminal synthases

Terminal PG synthases are involved in the conversion of PGH₂ to active PGs and TXs downstream of COX. These enzymes are PGE synthase (PGES) for PGE₂, PGDS for PGD₂, PGFS for PGF_{2α}, PGIS for PGI₂ and TXS for TXA₂, respectively. In the final step of PGE₂ biosynthesis downstream of COX, PGES specifically catalyzes the conversion of PGH₂ to PGE₂. The authors have previously reported that microsomal PGES (mPGES)-1, one of the PGES isozymes, is dramatically upregulated coordinately with COX-2 by stimulation with IL-1β and TNF-α in cultured synovial fibroblasts from RA patients [16–18]. Subsequently, it was demonstrated that mPGES-1 expression is correlated with disease activity in synovial lining cells, macrophages and fibroblasts in the

sublining, and mononuclear infiltrates and vascular endothelial cells in synovial tissues of patients with RA [19,20]. In addition, the potential importance of mPGES-1 in arthritis has been directly supported by the reduction of inflammatory and histopathological changes in CIA and collagen antibody-induced arthritis (CAIA) models in mPGES-1-null mice [21,22]. The absence of mPGES-1 appears equivalent to the absence of COX-2 in reducing PGE₂ production while preserving biosynthesis of alternate PGs. mPGES-1 might be an attractive future therapeutic target to control specifically the overproduction of PGE₂ associated with the inflammation of RA.

In contrast to inhibiting proinflammatory PGE₂, elevating the levels of anti-inflammatory PGs may also prove fruitful in the treatment of arthritis. PGDS is a key enzyme for the production of PGD₂ and its metabolite, 15-deoxy-Δ¹², 14-PGJ₂ (15d-PGJ₂), that has been shown to exert anti-inflammatory/resolution effects on peripheral inflammation [23]. PGD₂ acts via at least two GPCRs, designated DP1 and DP2 receptors. It has been reported that retroviral expression of PGDS can suppress inflammatory responses in some animal models of inflammation [24,25]. In addition, 15d-PGJ₂ has an inhibitory effect on proinflammatory mediator production such as IL-1β, TNF-α, inducible nitric oxide synthase (iNOS), MMP-13 and MMP-1 via inhibition of inflammatory gene transcription by nuclear receptor activation in the articular component [26–28]. The mechanism by which 15d-PGJ₂ mediates its anti-inflammatory effects is not well known; however, recent studies suggest its ability to bind and activate peroxisome proliferator-activated receptor (PPAR)γ and activate anti-inflammatory mechanisms that help in the resolution of inflammation [29]. Furthermore, 15d-PGJ₂ also has a negative feedback effect on AA metabolic enzymes, such as COX-2 and mPGES-1, associated with the generation of proinflammatory PGE₂ [30–33]. In several studies, the anti-inflammatory effects of 15d-PGJ₂ seem to be partly associated with its pro-apoptotic effects on inflammatory cells. 15d-PGJ₂ inhibits the growth of RA synovial fibroblasts by apoptosis and also attenuates the inflammatory features, such as pannus formation and mononuclear cell infiltration, in a model of adjuvant-induced arthritis [34]. These studies suggest that 15d-PGJ₂ may be another useful therapeutic approach for inflammatory arthritis.

Lipoxygenase

Another alternative metabolism of AA occurs via the LOX enzyme, which results in the production of HETEs and LTs. LOX metabolic enzymes, including 15- and 12-LOX, yield 15- and 12-HETE, respectively, by metabolizing AA, whereas 5-LOX metabolizes AA to LTs [35]. 5-LOX metabolism of AA produces LTA₄ (an unstable metabolite), which is subsequently hydrolyzed by LTA₄ hydrolase (LTA₄H) to LTB₄, or LTC₄ by LTC₄ synthase in combination with reduced glutathione. LTC₄ is further converted to LTD₄ by γ-glutamyl leukotrienase (GL) and γ-glutamyl transpeptidase (GT), and converted subsequently to LTE₄ by dipeptidase [36,37].

Leukotrienes

LTs mediate most of their effects via LT receptors belonging to the GPCR superfamily. Cysteinyl LTs (CysLTs) including LTC₄, D₄ and E₄ bind to CysLT receptors (CysLT₁ and CysLT₂) [38], whereas LTB₄ acts by binding either to BLT₁ or BLT₂ receptors [39].

The role of CysLTs in RA is poorly understood. However, reports suggest that 5-LOX and its metabolic product, LTB₄, may play a key role in the pathophysiology of RA. Elevated levels of 5-LOX and LTB₄ have been observed in the serum, synovial fluid and synovium of the patients with RA [40,41]. One of the major pro-inflammatory actions of LTB₄ is its ability to act as a potent chemotactic factor for neutrophils [42]. Neutrophils are the predominant leukocytes present in synovial fluid. In addition, neutrophils in the inflamed joints are the major source of LTB₄ production in patients with RA [43]. Although neutrophils have long been considered as the major source of LTB₄, other cell types including macrophages, lymphocytes and mast cells also produce substantial amounts of LTB₄ under proinflammatory conditions [44–46]. A recent study demonstrated that the LTB₄ receptors BLT₁ or BLT₂ are expressed in the synovial tissues of patients with RA [47]. This study also showed that BLT₂ is the main receptor mediating the effects of LTB₄ in human RA synovial tissues.

Animal studies have also shown elevated levels of LTB₄ in the synovial fluid of carrageenan-induced arthritis and CIA, and it is a key mediator involved in the progression of CIA [42,48]. MK886, an inhibitor of LTB₄ synthesis, has been shown to decrease the articular inflammation, proinflammatory mediator production and joint destruction in a murine model of CIA [49].

Licofelone (ML3000), a dual inhibitor of the 5-LOX and COX pathways, has shown promising results in reducing joint destruction in an animal model of arthritis [50]; however, the efficacy of this and other similar drugs need further assessment in human RA.

Lipoxins

In 1971, Sir John Vane first demonstrated that the mechanism of action of aspirin and other nonsteroidal anti-inflammatory drugs (NSAIDs) is due to the inhibition of COX and subsequent inhibition of PG production. However, in recent years, new endogenous mechanisms of aspirin action have been discovered. In addition to inhibiting PG production, aspirin has the ability to acetylate COX-2 and initiate the production of novel endogenous lipids called LXs or aspirin-triggered LXs (ATL) [51]. COX-2 in the presence of aspirin catalyzes the conversion of AA to 15-HETE, which is further metabolized to 15-epi LXs by the 5-LOX pathway. LXs, including LXA₄ and LXB₄, can also be generated endogenously during cell–cell interactions between products of 15- and 5-LOX or 5- and 12-LOX. LXs act via a specific GPCR known as LXA₄ receptor (ALXR) [52].

LXs mediate stop signals that help in the resolution of inflammation. In a number of animal models of inflammation, LXs have been shown to limit neutrophil chemotaxis, adherence and transmigration [53], inhibit eosinophil migration [54], inhibit superoxide anion generation [55] and attenuate the production of proinflammatory mediators such as TNF- α [56] and LTD₄ [57].

Knowledge of the role of these novel anti-inflammatory lipid mediators in RA is poorly understood to date; however, studies suggest they have therapeutic potential for counteracting inflammation. LXA₄ has been shown to decrease the levels of IL-1 β -induced IL-6, IL-8 and MMP-3 in RA synovial fibroblasts [58]. In addition, LXA₄ has also been shown to stimulate TIMP-2 in human synovial fibroblasts [59]. Furthermore, treatment of neutrophils isolated from RA patients with ionophore A23187 results in elevated levels of LXs [60], further suggesting the anti-inflammatory/resolution effects of these lipid mediators.

Apart from LXs, aspirin treatment of human tissues *in vitro* and murine tissues *in vivo* have been shown to generate novel 17-*R*-hydroxy series docosanoids involved in limiting inflammation. These novel mediators are the oxygenated products derived from omega-3

polyunsaturated fatty acids. Since these mediators are generated during the resolution phase of inflammation, they are termed resolvins [61].

Efficacy of NSAIDs for the treatment of RA

NSAIDs act by inhibiting COX activity and are used extensively for the management of pain, swelling and stiffness in RA [62]. Nonselective or traditional NSAIDs, including indomethacin, ibuprofen and naproxen, although effective in the management of pain associated with RA, are associated with a number of gastrointestinal and renal side effects as these drugs inhibit both COX-1 and -2 activity and inhibit the production of PGs and TXs [63]. Since PGs generated by COX-1 are vital for the maintenance of gastrointestinal and renal function, selective COX-2 inhibitors (COXIBs), including celecoxib and rofecoxib, were developed and used extensively for the treatment of RA and other inflammatory and painful diseases. Despite improved symptoms, clinical trials have demonstrated that use of selective COXIBs may lead to an increased risk of cardiovascular events [64–66].

These studies have raised several questions regarding the balance of safety and efficacy for these drugs in the treatment of inflammatory diseases, including RA. New targets within the AA metabolism are being identified to overcome the possible adverse effects associated with selective COX-2 inhibition. Since studies using mPGES-1-deficient mice reveal resistance to pain, arthritis and fever, selective mPGES-1 inhibitors are the most promising. However, no specific inhibitor of this enzyme is currently available.

Future perspective

Eicosanoids have been implicated as key mediators involved in the inflammatory symptoms of RA, particularly pain, stiffness and swelling. NSAIDs and COXIBs are used extensively for the treatment of RA, even in this era of biologic therapies. However, when one looks at the history of NSAIDs and COXIBs, more drugs have failed than succeeded. One example of this is that, of the three COXIBs indicated for the treatment of RA released since 1999, only one remains on the market. The current issues include ongoing gastrointestinal risk that, while reduced by several strategies, including co-administration of proton pump inhibitors or misoprostol and the use of COX-2-specific inhibitors, have not eliminated serious gastrointestinal events associated with these medications. Co-administration of low-dose aspirin,

whose use is quite frequent in patients with RA, especially in light of the increased cardiovascular risk in this disease, increases gastrointestinal toxicity. All of these drugs have renal effects, including increased blood pressure, which may contribute to long-term issues with cardiovascular adverse events. The cardiovascular issues continue to be troublesome and it cannot be concluded from current data that any drug that inhibits COX-2, whether nonspecific or specific, has no adverse effects.

The fact that the COX isozymes sit relatively high up in the biosynthetic pathway and lead to inhibition of all PGs make inhibition of these enzymes rather crude tools if indeed the desire is to inhibit the production or action of only one PG or block its actions. The current task is to

devise new therapeutic tools to control the chronic inflammation associated with RA. More specific targeting of the eicosanoid pathway may allow for benefit, while minimizing the risks. As research progresses to provide evidence of the specific eicosanoids involved in inflammation and anti-inflammation aimed specifically at altering this balance, more targeted therapies will become possible. At present, inhibition of PGE₂ production and action seems most likely to treat the pain and inflammation of RA. Promoting increased levels of anti-inflammatory eicosanoids may also prove fruitful in the treatment of RA. These strategies may be useful for symptom modification or possibly in disease modification. At present, the potential adverse consequences of alternate treatment strategies remain unclear.

Executive summary

Cyclooxygenase

- Cyclooxygenase (COX) is mainly comprised of two isozymes, constitutive COX-1 and inducible COX-2.
- Induction of COX-2 expression has been observed in inflamed synovial tissues of rheumatoid arthritis (RA) patients with the disease-related pattern.

Prostaglandins

- Elevated levels of proinflammatory prostaglandin (PG)E₂ are observed in the synovial fluid of patients with RA.
- Microsomal PGE synthase (mPGES)-1 is expressed in correlation with the extent of disease activity in synovial tissues of patients with RA.
- Deletion of the PGE₂ biosynthetic enzyme and its receptor action have been demonstrated to reduce inflammatory and histopathological changes in arthritis animal models.
- Anti-inflammatory PGs, such as PGD₂ and 15-deoxy-Δ¹², 14-PGJ₂, may be a useful therapeutic approach for inflammatory arthritis.

Lipoxygenase

- Lipoxygenase (LOX) metabolic enzymes, including 15- and 12-LOX, yield 15-hydroxyeicosatetraenoic acid (HETE) and 12-HETE, respectively, by metabolizing arachidonic acid, whereas 5-LOX metabolizes arachidonic acid to leukotrienes (LTs).

Leukotrienes

- Elevated levels of LTB₄ have been observed in the serum, synovial fluid and synovium of patients with RA.
- Inhibitors of LTB₄ synthesis have been shown to decrease articular inflammation, proinflammatory mediator production and joint destruction in animal models of arthritis.

Lipoxins

- Aspirin has the ability to acetylate COX-2 and initiate the production of novel endogenous lipids called lipoxins.
- Lipoxins mediate stop signals that help in the resolution of inflammation.

Nonsteroidal anti-inflammatory drugs

- Recent clinical trials have demonstrated that use of selective COX-2 inhibitors can lead to an increased risk of cardiovascular events.
- mPGES-1 is an attractive therapeutic target for RA disease.

Bibliography

Papers of special note have been highlighted as either of interest (•) or of considerable interest (••) to readers.

1. Xie WL, Chipman JG, Robertson DL, Erikson RL, Simmons DL: Expression of a mitogen-responsive gene encoding prostaglandin synthase is regulated by mRNA splicing. *Proc. Natl Acad. Sci. USA* 88(7), 2692–2696 (1991).
2. Kujubu DA, Fletcher BS, Varnum BC, Lim RW, Herschman HR: *J. Biol. Chem.* 266(20), 12866–12872 (1991).
3. Chandrasekharan NV, Dai H, Roos KL *et al.*: TIS10, a phorbol ester tumor promoter-inducible mRNA from Swiss 3T3 cells, encodes a novel prostaglandin synthase/cyclooxygenase homologue. *Proc. Natl Acad. Sci. USA* 99(21), 13926–13931 (2002).
4. Qin N, Zhang SP, Reitz TL, Mei JM, Flores CM: Cloning, expression, and functional characterization of human cyclooxygenase-1 splicing variants: evidence for intron 1 retention. *J. Pharmacol. Exp. Ther.* 315(3), 1298–1305 (2005).
5. Kang RY, Freire-Moar J, Sigal, E, Chu CQ: Cloning, expression, and functional characterization of human cyclooxygenase-1 splicing variants: evidence for intron 1 retention. *Br. J. Rheumatol.* 35(8), 711–718 (1996).
6. Siegle I, Klein T, Backman JT, Saal JG, Nusing RM, Fritz P: *Arthritis Rheum.* 41(1), 122–129 (1998).
7. Angel J, Berenbaum F, Le Denmat C, Nevalainen T, Masliah J, Fournier C: Expression of cyclooxygenase 1 and cyclooxygenase 2 in human synovial tissue: differential elevation of cyclooxygenase 2 in inflammatory joint diseases. *Eur. J. Biochem.* 226(1), 125–131 (1994).
8. Crofford LJ, Wilder RL, Ristimaki AP *et al.*: Cyclooxygenase-1 and -2 expression in rheumatoid synovial tissues. Effects of interleukin-1 β , phorbol ester, and corticosteroids. *J. Clin. Invest.* 93(3), 1095–1101 (1994).
9. Hulkower KI, Wertheimer SJ, Levin W *et al.*: Interleukin-1 β induces cytosolic phospholipase A₂ and prostaglandin H synthase in rheumatoid synovial fibroblasts. Evidence for their roles in the production of prostaglandin E₂. *Arthritis Rheum.* 37(5), 653–661 (1994).
10. Egg D, Gunther R, Herold M, Kerschbaumer F: Prostaglandins E₂ and F₂ α concentrations in the synovial fluid in rheumatoid and traumatic knee joint diseases. *Z. Rheumatol.* 39(5–6), 170–175 (1980).
11. Crofford LJ: COX-2 in synovial tissues. *Osteoarthr. Cartil.* 7(4), 406–408 (1999).
12. Kawai S, Nishida S, Kato M *et al.*: Comparison of cyclooxygenase-1 and -2 inhibitory activities of various nonsteroidal anti-inflammatory drugs using human platelets and synovial cells. *Eur. J. Pharmacol.* 347(1), 87–94 (1998).
13. Inoue H, Takamori M, Shimoyama Y, Ishibashi H, Yamamoto, S, Koshihara Y: Regulation by PGE₂ of the production of interleukin-6, macrophage colony stimulating factor, and vascular endothelial growth factor in human synovial fibroblasts. *Br. J. Pharmacol.* 136(2), 287–295 (2002).
- **Interesting work on prostaglandin (PG)E₂ regulation.**
14. Masuko-Hongo K, Sato T, Nishioka K: Chemokines differentially induce matrix metalloproteinase-3 and prostaglandin E₂ in human articular chondrocytes. *Clin. Exp. Rheumatol.* 23(1), 57–62 (2005).
15. McCoy JM, Wicks JR, Audoly LP: The role of prostaglandin E₂ receptors in the pathogenesis of rheumatoid arthritis. *J. Clin. Invest.* 110(5), 651–658 (2002).
- **Interesting piece of work on PGE₂ signaling via the EP4 receptor using EP4 knockout mice.**
16. Kojima F, Naraba H, Sasaki Y, Okamoto R, Koshino T, Kawai S: Coexpression of microsomal prostaglandin E synthase with cyclooxygenase-2 in human rheumatoid synovial cells. *J. Rheumatol.* 29(9), 1836–1842 (2002).
17. Stichtenoth DO, Thoren S, Bian H, Peters-Golden M, Jakobsson PJ, Crofford LJ: Microsomal prostaglandin E synthase is regulated by proinflammatory cytokines and glucocorticoids in primary rheumatoid synovial cells. *J. Immunol.* 167(1), 469–474 (2001).
- **First report on induction of microsomal PGE synthase (mPGES)-1 in rheumatoid arthritis synovial fibroblasts.**
18. Kojima F, Naraba H, Sasaki Y, Beppu M, Aoki H, Kawai S: Prostaglandin E₂ is an enhancer of interleukin-1 β -induced expression of membrane-associated prostaglandin E synthase in rheumatoid synovial fibroblasts. *Arthritis Rheum.* 48(10), 2819–2828 (2003).
- **First study to show the positive feedback mechanism of mPGES-1 induction by PGE₂.**
19. Westman M, Korotkova M, af Klint E *et al.*: Expression of microsomal prostaglandin E synthase 1 in rheumatoid arthritis synovium. *Arthritis Rheum.* 50(6), 1774–1780 (2004).
20. Murakami M, Nakashima K, Kamei D *et al.*: Cellular prostaglandin E₂ production by membrane-bound prostaglandin E synthase-2 via both cyclooxygenases-1 and -2. *J. Biol. Chem.* 278(39), 37937–37947 (2003).
21. Trebino CE, Stock JL, Gibbons CP *et al.*: Impaired inflammatory and pain responses in mice lacking an inducible prostaglandin E synthase. *Proc. Natl Acad. Sci. USA* 100(15), 9044–9049 (2003).
- **First report to define the role of mPGES-1 in pain and inflammatory responses using mPGES-1 knockout mice.**
22. Kamei D, Yamakawa K, Takegoshi Y *et al.*: Reduced pain hypersensitivity and inflammation in mice lacking microsomal prostaglandin E synthase-1. *J. Biol. Chem.* 279(32), 33684–33695 (2004).
23. Gilroy DW, Colville-Nash PR, Willis D, Chivers J, Paul-Clark MJ, Willoughby DA: Inducible cyclooxygenase may have anti-inflammatory properties. *Nat. Med.* 5(6), 698–701 (1999).
24. Ando M, Murakami Y, Kojima F *et al.*: Retrovirally introduced prostaglandin D₂ synthase suppresses lung injury induced by bleomycin. *Am. J. Respir. Cell Mol. Biol.* 28(5), 582–591 (2003).
25. Murakami Y, Akahoshi T, Hayashi I *et al.*: Inhibition of monosodium urate monohydrate crystal-induced acute inflammation by retrovirally transfected prostaglandin D synthase. *Arthritis Rheum.* 48(10), 2931–2941 (2003).
26. Ji JD, Cheon H, Jun JB *et al.*: Effects of peroxisome proliferator-activated receptor- γ (PPAR- γ) on the expression of inflammatory cytokines and apoptosis induction in rheumatoid synovial fibroblasts and monocytes. *J. Autoimmun.* 17(3), 215–221 (2001).
27. Fahmi H, Di Battista JA, Pelletier JP, Mineau F, Ranger P, Martel-Pelletier J: Peroxisome proliferator-activated receptor γ activators inhibit interleukin-1 β -induced nitric oxide and matrix metalloproteinase 13 production in human chondrocytes. *Arthritis Rheum.* 44(3), 595–607 (2001).
28. Fahmi H, Pelletier JP, Di Battista JA, Cheung HS, Fernandes JC, Martel-Pelletier J: Peroxisome proliferator-activated receptor γ activators inhibit MMP-1 production in human synovial fibroblasts likely by reducing the binding of the activator protein 1. *Osteoarthr. Cartil.* 10(2), 100–108 (2002).

29. Ricote M, Li AC, Willson TM, Kelly CJ, Glass CK: The peroxisome proliferator-activated receptor- γ is a negative regulator of macrophage activation. *Nature* 391(6662), 79–82 (1998).
30. Tsubouchi Y, Kawahito Y, Kohno M, Inoue K, Hla T, Sano H: Feedback control of the arachidonate cascade in rheumatoid synoviocytes by 15-deoxy- Δ (12,14)-prostaglandin J_2 . *Biochem. Biophys. Res. Commun.* 283(4), 750–755 (2001).
31. Farrajota K, Cheng S, Martel-Pelletier J *et al.*: Inhibition of interleukin-1 β -induced cyclooxygenase 2 expression in human synovial fibroblasts by 15-deoxy- Δ 12,14-prostaglandin J_2 through a histone deacetylase-independent mechanism *Arthritis Rheum.* 52(1), 94–104 (2005).
32. Fahmi H, Pelletier JP, Mineau F, Martel-Pelletier J: 15d-PGJ(2) is acting as a 'dual agent' on the regulation of COX-2 expression in human osteoarthritic chondrocytes. *Osteoarthr. Cartil.* 10(11), 845–848 (2002).
33. Cheng S, Afif H, Martel-Pelletier J *et al.*: Activation of peroxisome proliferator-activated receptor γ inhibits interleukin-1 β -induced membrane-associated prostaglandin E_2 synthase-1 expression in human synovial fibroblasts by interfering with Egr-1. *J. Biol. Chem.* 279(21), 22057–22065 (2004).
34. Kawahito Y, Kondo M, Tsubouchi Y *et al.*: 15-deoxy- Δ (12,14)-PGJ(2) induces synoviocyte apoptosis and suppresses adjuvant-induced arthritis in rats. *J. Clin. Invest.* 106(2), 189–197 (2000).
- **Interesting study focusing on apoptotic effects of 15-deoxy- Δ 12,14-PGJ $_2$ in arthritis.**
35. Williams KI, Higgs GA: Eicosanoids and inflammation. *J. Pathol.* 156(2), 101–110 (1988).
36. Peters-Golden M, Brock TG: 5-lipoxygenase and FLAP. *Prostaglandins Leukot. Essent. Fatty Acids* 69(2–3), 99–109 (2003).
37. Samuelsson B: The discovery of the leukotrienes. *Am. J. Respir. Crit. Care Med.* 161(2 Pt 2), S2–S6 (2000).
38. Busse W, Kraft M: Cysteinyl leukotrienes in allergic inflammation: strategic target for therapy. *Chest* 127(4), 1312–1326 (2005).
39. Yokomizo T, Izumi T, Chang K, Takuwa Y, Shimizu T: A G-protein-coupled receptor for leukotriene B_4 that mediates chemotaxis. *Nature* 387(6633), 620–624 (1997).
40. Sawazaki Y: Leukotriene B_4 , leukotriene C_4 and prostaglandin E_2 in the serum, synovial fluid and synovium in patients with rheumatoid arthritis. *Nippon Ika Daigaku Zasshi* 56(6), 559–564 (1989).
41. Bonnet C, Bertin P, Cook-Moreau J, Chable-Rabinovitch H, Treves R, Rigaud M: Lipoxygenase products and expression of 5-lipoxygenase and 5-lipoxygenase-activating protein in human cultured synovial cells. *Prostaglandins* 50(3), 127–135 (1995).
42. Herlin T, Fogh K, Hansen ES *et al.*: 15-HETE inhibits leukotriene B_4 formation and synovial cell proliferation in experimental arthritis. *Agents Actions* 29(1–2), 52–53 (1990).
43. Moilanen E, Alanko J, Nissila M, Hamalainen M, Isomaki H, Vapaatalo H: Eicosanoid production in rheumatoid synovitis. *Agents Actions* 28(3–4), 290–297 (1989).
44. Fels AO, Pawlowski NA, Cramer EB, King TK, Cohn ZA, Scott WA: Human alveolar macrophages produce leukotriene B_4 . *Proc. Natl Acad. Sci. USA* 79(24), 7866–7870 (1982).
45. Jakobsson PJ, Steinhilber D, Odlander B, Radmark O, Claesson HE, Samuelsson B: On the expression and regulation of 5-lipoxygenase in human lymphocytes. *Proc. Natl Acad. Sci. USA* 89(8), 3521–3525 (1992).
46. Ford-Hutchinson AW: Leukotriene B_4 in inflammation. *Crit. Rev. Immunol.* 10(1), 1–12 (1990).
47. Hashimoto A, Endo H, Hayashi I *et al.*: Differential expression of leukotriene B_4 receptor subtypes (BLT $_1$ and BLT $_2$) in human synovial tissues and synovial fluid leukocytes of patients with rheumatoid arthritis. *J. Rheumatol.* 30(8), 1712–1718 (2003).
48. Griffiths RJ, Pettipher ER, Koch K *et al.*: Leukotriene B_4 plays a critical role in the progression of collagen-induced arthritis. *Proc. Natl Acad. Sci. USA* 92(2), 517–521 (1995).
49. Canetti CA, Leung BP, Culshaw S, McInnes IB, Cunha FQ, Liew FY: IL-18 enhances collagen-induced arthritis by recruiting neutrophils via TNF- α and leukotriene B_4 . *J. Immunol.* 171(2), 1009–1015 (2003).
50. Gay RE, Neidhart M, Pataky F, Tries S, Laufer S, Gay S: Dual inhibition of 5-lipoxygenase and cyclooxygenases-1 and-2 by ML3000 reduces joint destruction in adjuvant arthritis. *J. Rheumatol.* 28(9), 2060–2065 (2001).
51. Claria J, Serhan CN: Aspirin triggers previously undescribed bioactive eicosanoids by human endothelial cell-leukocyte interactions. *Proc. Natl Acad. Sci. USA* 92(21), 9475–9479 (1995).
52. Fiore S, Maddox JF, Perez HD, Serhan CN: Identification of a human cDNA encoding a functional high affinity lipoxin A_4 receptor. *J. Exp. Med.* 180(1), 253–260 (1994).
53. Devchand PR, Arita M, Hong S *et al.*: Human ALX receptor regulates neutrophil recruitment in transgenic mice: roles in inflammation and host defense. *FASEB J.* 17(6), 652–659 (2003).
54. Bandeira-Melo C, Bozza PT, Diaz BL *et al.*: Cutting edge: lipoxin (LX) A_4 and aspirin-triggered 15-epi-LXA $_4$ block allergen-induced eosinophil trafficking. *J. Immunol.* 164(5), 2267–2271 (2000).
55. Levy BD, Fokin VV, Clark JM, Wakelam MJ, Petasis NA, Serhan CN: Polyisoprenyl phosphate (PIPP) signaling regulates phospholipase D activity: a 'stop' signaling switch for aspirin-triggered lipoxin A_4 . *FASEB J.* 13(8), 903–911 (1999).
56. Wu SH, Lu C, Dong L, Zhou GP, He ZG, Chen ZQ: Lipoxin A_4 inhibits TNF- α -induced production of interleukins and proliferation of rat mesangial cells. *Kidney Int.* 68(1), 35–46 (2005).
57. McMahon B, Stenson C, McPhillips F, Fanning A, Brady HR, Godson C: Lipoxin A_4 antagonizes the mitogenic effects of leukotriene D_4 in human renal mesangial cells. Differential activation of MAP kinases through distinct receptors. *J. Biol. Chem.* 275(36), 27566–27575 (2000).
58. Sodin-Semrl S, Taddeo B, Tseng D, Varga J, Fiore S: Lipoxin A_4 inhibits IL-1 β -induced IL-6, IL-8, and matrix metalloproteinase-3 production in human synovial fibroblasts and enhances synthesis of tissue inhibitors of metalloproteinases. *J. Immunol.* 164(5), 2660–2666 (2000).
- **One of the first reports on the role of lipoxins in arthritis.**
59. Sodin-Semrl S, Spagnolo A, Barbaro B, Varga J, Fiore S: Lipoxin A_4 counteracts synergistic activation of human fibroblast-like synoviocytes. *Int. J. Immunopathol. Pharmacol.* 17(1), 15–25 (2004).
60. Thomas E, Leroux JL, Blotman F, Chavis C: Conversion of endogenous arachidonic acid to 5,15-diHETE and lipoxins by polymorphonuclear cells from patients with rheumatoid arthritis. *Inflamm. Res.* 44(3), 121–124 (1995).
61. Serhan CN, Hong S, Gronert K *et al.*: *J. Exp. Med.* 196(8), 1025–1037 (2002).
- **Interesting overview of the anti-inflammatory properties of resolvins.**
62. Crofford LJ: COX-2: where are we in 2003? – Specific cyclooxygenase-2 inhibitors and aspirin-exacerbated respiratory disease. *Arthritis. Res. Ther.* 5(1), 25–27 (2003).

63. Meade EA, Smith WL, DeWitt DL: Differential inhibition of prostaglandin endoperoxide synthase (cyclooxygenase) isozymes by aspirin and other non-steroidal anti-inflammatory drugs. *J. Biol. Chem.* 268(9), 6610–6614 (1993).
64. Bresalier RS, Sandler RS, Quan H *et al.*: Cardiovascular events associated with rofecoxib in a colorectal adenoma chemoprevention trial. *N. Engl. J. Med.* 352(11), 1092–1102 (2005).
- **One of the earliest clinical trials on cardiovascular events associated with rofecoxib.**
65. Solomon SD, McMurray JJ, Pfeffer MA *et al.*: Cardiovascular risk associated with celecoxib in a clinical trial for colorectal adenoma prevention. *N. Engl. J. Med.* 352(11), 1071–1080 (2005).

- **One of the earliest clinical trials on the cardiovascular effects of celecoxib.**
66. White WB, Faich G, Borer JS, Makuch RW: Cardiovascular thrombotic events in arthritis trials of the cyclooxygenase-2 inhibitor celecoxib. *Am. J. Cardiol.* 92(4), 411–418 (2003).

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